



40 **zoo**reach  
Zoo Outreach Organisation  
Years

Open Access

Building evidence for conservation globally  
**Journal of  
Threatened  
Taxa**

10.11609/jott.2026.18.2.28262-28454  
[www.threatenedtaxa.org](http://www.threatenedtaxa.org)

26 February 2026 (Online & Print)  
18(2): 28262-28454  
ISSN 0974-7907 (Online)  
ISSN 0974-7893 (Print)



ISSN 0974-7907 (Online); ISSN 0974-7893 (Print)

Publisher  
**Wildlife Information Liaison Development Society**  
www.wild.zooreach.org

Host  
**Zoo Outreach Organization**  
www.zooreach.org

Srivari Illam, No. 61, Karthik Nagar, 10th Street, Saravanampatti, Coimbatore, Tamil Nadu 641035, India  
Registered Office: 3A2 Varadarajulu Nagar, FCI Road, Ganapathy, Coimbatore, Tamil Nadu 641006, India  
Ph: +91 9385339863 | [www.threatenedtaxa.org](http://www.threatenedtaxa.org)  
Email: [sanjay@threatenedtaxa.org](mailto:sanjay@threatenedtaxa.org)

#### EDITORS

##### Founder & Chief Editor

**Dr. Sanjay Molur**

Wildlife Information Liaison Development (WILD) Society & Zoo Outreach Organization (ZOO),  
Coimbatore, Tamil Nadu 641006, India

##### Assistant Editor

**Dr. Chaithra Shree J.**, WILD/ZOO, Coimbatore, Tamil Nadu 641006, India

##### Managing Editor

**Mr. B. Ravichandran**, WILD/ZOO, Coimbatore, Tamil Nadu 641006, India

##### Associate Editors

**Dr. Mandar Paingankar**, Government Science College Gadchiroli, Maharashtra 442605, India

**Dr. Ulrike Streicher**, Wildlife Veterinarian, Eugene, Oregon, USA

**Ms. Priyanka Iyer**, ZOO/WILD, Coimbatore, Tamil Nadu 641006, India

##### Board of Editors

**Dr. Russel Mittermeier**

Executive Vice Chair, Conservation International, Arlington, Virginia 22202, USA

##### Prof. Mewa Singh Ph.D., FASC, FNA, FNASC, FNAPsy

Ramanna Fellow and Life-Long Distinguished Professor, Biopsychology Laboratory, and  
Institute of Excellence, University of Mysore, Mysuru, Karnataka 570006, India; Honorary  
Professor, Jawaharlal Nehru Centre for Advanced Scientific Research, Bangalore; and Adjunct  
Professor, National Institute of Advanced Studies, Bangalore

##### Stephen D. Nash

Scientific Illustrator, Conservation International, Dept. of Anatomical Sciences, Health Sciences  
Center, T-8, Room 045, Stony Brook University, Stony Brook, NY 11794-8081, USA

##### Dr. Fred Pluthero

Toronto, Canada

##### Dr. Priya Davidar

Sigur Nature Trust, Chadapatti, Mavinhalla PO, Nilgiris, Tamil Nadu 643223, India

##### Dr. John Fellowes

Honorary Assistant Professor, The Kadoorie Institute, 8/F, T.T. Tsui Building, The University of  
Hong Kong, Pokfulam Road, Hong Kong

##### Prof. Dr. Mirco Solé

Universidade Estadual de Santa Cruz, Departamento de Ciências Biológicas, Vice-coordenador  
do Programa de Pós-Graduação em Zoologia, Rodovia Ilhéus/Itabuna, Km 16 (45662-000)  
Salobrinho, Ilhéus - Bahia - Brasil

##### Dr. Rajeev Raghavan

Professor of Taxonomy, Kerala University of Fisheries & Ocean Studies, Kochi, Kerala, India

##### English Editors

**Mrs. Mira Bhojwani**, Pune, India

**Dr. Fred Pluthero**, Toronto, Canada

##### Copy Editors

**Ms. Usha Madgunaki**, Zooreach, Coimbatore, India

**Ms. Trisa Bhattacharjee**, Zooreach, Coimbatore, India

**Ms. Paloma Noronha**, Daman & Diu, India

##### Web Development

**Mrs. Latha G. Ravikumar**, ZOO/WILD, Coimbatore, India

##### Typesetting

**Mrs. Radhika**, Zooreach, Coimbatore, India

**Mrs. Geetha**, Zooreach, Coimbatore, India

#### Fundraising/Communications

**Mrs. Payal B. Molur**, Coimbatore, India

#### Subject Editors 2021–2023

##### Fungi

Dr. B. Shivaraju, Bengaluru, Karnataka, India

Dr. R.K. Verma, Tropical Forest Research Institute, Jabalpur, India

Dr. Vatsavaya S. Raju, Kakatiya University, Warangal, Andhra Pradesh, India

Dr. M. Krishnappa, Jnana Sahyadri, Kuvempu University, Shimoga, Karnataka, India

Dr. K.R. Sridhar, Mangalore University, Mangalagangothri, Mangalore, Karnataka, India

Dr. Gunjan Biswas, Vidyasagar University, Midnapore, West Bengal, India

Dr. Kiran Ramchandra Ranadive, Annasaheb Magar Mahavidyalaya, Maharashtra, India

##### Plants

Dr. G.P. Sinha, Botanical Survey of India, Allahabad, India

Dr. N.P. Balakrishnan, Ret. Joint Director, BSI, Coimbatore, India

Dr. Shonil Bhagwat, Open University and University of Oxford, UK

Prof. D.J. Bhat, Retd. Professor, Goa University, Goa, India

Dr. Ferdinando Boero, Università del Salento, Lecce, Italy

Dr. Dale R. Calder, Royal Ontario Museum, Toronto, Ontario, Canada

Dr. Cleofas Cervancia, Univ. of Philippines Los Baños College Laguna, Philippines

Dr. F.B. Vincent Florens, University of Mauritius, Mauritius

Dr. Merlin Franco, Curtin University, Malaysia

Dr. V. Irudayaraj, St. Xavier's College, Palayamkottai, Tamil Nadu, India

Dr. B.S. Kholia, Botanical Survey of India, Gangtok, Sikkim, India

Dr. Pankaj Kumar, Department of Plant and Soil Science, Texas Tech University, Lubbock, Texas, USA

Dr. V. Sampath Kumar, Botanical Survey of India, Howrah, West Bengal, India

Dr. A.J. Solomon Raju, Andhra University, Visakhapatnam, India

Dr. Vijayasankar Raman, University of Mississippi, USA

Dr. B. Ravi Prasad Rao, Sri Krishnadevaraya University, Anantpur, India

Dr. K. Ravikumar, FRLHT, Bengaluru, Karnataka, India

Dr. Aparna Watve, Pune, Maharashtra, India

Dr. Qiang Liu, Xishuangbanna Tropical Botanical Garden, Yunnan, China

Dr. Noor Azhar Mohamed Shazili, Universiti Malaysia Terengganu, Kuala Terengganu, Malaysia

Dr. M.K. Vasudeva Rao, Shiv Ranjani Housing Society, Pune, Maharashtra, India

Prof. A.J. Solomon Raju, Andhra University, Visakhapatnam, India

Dr. Mandar Datar, Agharkar Research Institute, Pune, Maharashtra, India

Dr. M.K. Janarthanam, Goa University, Goa, India

Dr. K. Karthigeeyan, Botanical Survey of India, India

Dr. Errol Vela, University of Montpellier, Montpellier, France

Dr. P. Lakshminarasimhan, Botanical Survey of India, Howrah, India

Dr. Larry R. Noblick, Montgomery Botanical Center, Miami, USA

Dr. K. Haridasan, Pallavur, Palakkad District, Kerala, India

Dr. Analinda Manila-Fajard, University of the Philippines Los Baños, Laguna, Philippines

Dr. P.A. Sinu, Central University of Kerala, Kasaragod, Kerala, India

Dr. Afroz Alam, Banasthali Vidyapith (accredited A grade by NAAC), Rajasthan, India

Dr. K.P. Rajesh, Zamorin's Guruvayurappan College, GA College PO, Kozhikode, Kerala, India

Dr. David E. Boufford, Harvard University Herbaria, Cambridge, MA 02138-2020, USA

Dr. Ritesh Kumar Choudhary, Agharkar Research Institute, Pune, Maharashtra, India

Dr. A.G. Pandurangan, Thiruvananthapuram, Kerala, India

Dr. Navendu Page, Wildlife Institute of India, Chandrabani, Dehradun, Uttarakhand, India

Dr. Kannan C.S. Warriar, Institute of Forest Genetics and Tree Breeding, Tamil Nadu, India

#### Invertebrates

Dr. R.K. Avasthi, Rohtak University, Haryana, India

Dr. D.B. Bastawade, Maharashtra, India

Dr. Partha Pratim Bhattacharjee, Tripura University, Suryamaninagar, India

Dr. Kailash Chandra, Zoological Survey of India, Jabalpur, Madhya Pradesh, India

Dr. Ansie Dippenaar-Schoeman, University of Pretoria, Queenswood, South Africa

Dr. Rory Dow, National Museum of Natural History Naturalis, The Netherlands

Dr. Brian Fisher, California Academy of Sciences, USA

Dr. Richard Gallon, Llandudno, North Wales, LL30 1UP

Dr. Hemant V. Ghate, Modern College, Pune, India

Dr. M. Monwar Hossain, Jahangirnagar University, Dhaka, Bangladesh

For Focus, Scope, Aims, and Policies, visit [https://threatenedtaxa.org/index.php/JoTT/aims\\_scope](https://threatenedtaxa.org/index.php/JoTT/aims_scope)

For Article Submission Guidelines, visit <https://threatenedtaxa.org/index.php/JoTT/about/submissions>

For Policies against Scientific Misconduct, visit [https://threatenedtaxa.org/index.php/JoTT/policies\\_various](https://threatenedtaxa.org/index.php/JoTT/policies_various)

continued on the back inside cover

Cover: Digital illustration of *Impatiens chamchumroonii* in Krita by Dupati Poojitha.



## Flower bud growth, mortality rate, and population structure of *Sapria himalayana* Griffith f. *albovinosa* Banziger & Hansen (Rafflesiaceae) in a subtropical forest, northeastern India

K. Shamran Maring<sup>1</sup> & Athokpam Pinokiyo<sup>2</sup>

<sup>1,2</sup>Department of Botany, Dhanamanjuri University, Imphal, Manipur 795001, India.

<sup>1</sup>puikans1996@gmail.com (corresponding author), <sup>2</sup>pinkithokpam@gmail.com

**Abstract:** *Sapria himalayana* Griff. f. *albovinosa* Banziger & Hansen (Rafflesiaceae) is rare and endemic to northeastern Thailand, Vietnam, and Manipur, with a short flowering season ranging from late October to late November, due to which its detailed phenology is poorly understood. To protect this valuable taxon from extinction threats, monitoring the growth of flower buds is crucial. The objectives of this study were to analyse the growth of flower buds at various developmental stages, the mortality rate, and the population structure of *Sapria himalayana* f. *albovinosa*. The buds were monitored, vertically photographed, and measurements of the plant buds and flowers were recorded for every observation. The present study resulted in the flower bud growth having six different developmental stages, namely the copule, copule-bract transition, bract, bract-perigone transition (BPT), perigone, and anthesis stage, with a diameter range of 0.3–2.3 cm, 2.4–4.0 cm, 4.1–5.5 cm, 5.6–6.1 cm, 6.2–8.1 cm, and 16–20 cm, respectively. The population was dominated by the copule stage in the initial observation, while it was dominated by the perigone and anthesis stages in the final observation, which indicated that the optimal flowering season was from late October to late November. A total of 58 flower buds were recorded, out of which 24.13% of flower buds were dead without reaching maturity due to fungal infections and were injured due to anthropogenic interventions. Hence, the information on the growth of buds, flower development stages and their mortality rate is vital for taxonomic studies, field monitoring, and conservation purposes.

**Keywords:** Anthesis, anthropogenic, bract, conservation, copule, crucial, endemic, extinction, perigone, rare.

**Maringa:** *Sapria himalayana* Griffith f. *albovinosa* Banziger & Hansen (Rafflesiaceae) tou ung paar a beimak taangni Northeastern Thailand, Vietnam and Manipur bi khyum heirou tomcha paar. Heirou paar ayei paar nei kheen heiri beimakni thum hoilei. Kheen heiri Kumthil tangla rou youbi Bilwa tangla bil tomcha paar. Ngam heini paar ayei maram ra thimi rapni langaidui muchangmak. Ayei paar a beimakni taangni pha ngam hoibini paar aya lhaipang aro mitmak nanung kalthung ngaakthung nei tloubam khat khoi. Heitounei ngamrou paar ayei kat um younei kheen rou youbini paar kim kham bil thi thung silthung nei tloubam heirou paar ayei khi nei changchap heirou paar ayei mashing (population) thi nei tloubam a namyerwa asiyei thilhak thi thut nei che aro shershing thutlei. Paar heiyei paarna bi heirou photo kou kanlhonni paar heiyei tang mandi (diameter in cm) namyerwa markhe bi langaidui thilhakur. Asiyei thilhak thi thut yei kheen heirou paar heiyei kat um yei kheen youbi kim kham bil heirou tangkak lailai paar heiyei tang malnamak thruktam mupha. Heirap heiri copule, copule-bract transition, bract, bract-perigone transition, perigone heirou anthesis stages with a diameter range of 0.3-2.3 cm, 2.4-4.0 cm, 4.1-5.5 cm, 5.6-6.1 cm, 6.2-8.1 cm and 16-20 cm, respectively. Paar heiyei mashing thithut youri nei tangla heirou ri copule stage ni duichi, heitoubini dongwai rou tangla heirou ri perigone heirou anthesis stages ya khintam ni duichi. Heiyei mukna ri paar heiri Kumthil tangla rou youbi Bilwa tangla bil matachani paar he paarnei kheen hoilei. Bi heirou ri paar mashing somnga-le-chot (58) paar muphalei, heiyei rilla saruk 24.13% paarshomakni hukur. Paar he mapung phamakni paarshomakni hukur heiyei ngamri rikwai dawurbi chulwurnei hoilei heirou lhaipang thimi ni paar leina bi heirou manshok nei ngam khat khi hoilei. Thakrei minlhangkur heirap namyer wani ngam hoibini beimak taangei pha paar awa lhangaidui thilhak ngakthung-silthung bini paar aya lhaipangpal bi aro mitheimak kim heimaknung beimak lungkhohi tloubam khat hoilei.

**Editor:** Inocencio Buot Jr., University of the Philippines Los Banos, Laguna, Philippines.

**Date of publication:** 26 February 2026 (online & print)

**Citation:** Maring, K.S. & A. Pinokiyo (2026). Flower bud growth, mortality rate, and population structure of *Sapria himalayana* Griffith f. *albovinosa* Banziger & Hansen (Rafflesiaceae) in a subtropical forest, northeastern India. *Journal of Threatened Taxa* 18(2): 28287–28295. <https://doi.org/10.11609/jott.9964.18.2.28287-28295>

**Copyright:** © Maring & Pinokiyo 2026. Creative Commons Attribution 4.0 International License. JoTT allows unrestricted use, reproduction, and distribution of this article in any medium by providing adequate credit to the author(s) and the source of publication.

**Funding:** None.

**Competing interests:** The authors declare no competing interests.

**Author details, Author contributions, Acknowledgements & Ethics and field compliance statement:** See end of this article.



## INTRODUCTION

*Sapria himalayana* Griffith f. *albovinosa* Banziger & Hansen (Banziger et al. 2000), a new form of the Hermit's Spittoon, is rare and endemic to the northeastern Thailand and Vietnam Lang Biang plateau, but a new distribution is recorded from India at the evergreen sub-tropical forest of Machi Village, Tengenoupal District, Manipur (Maring & Pinokiyo 2024). It is differentiated from the typical form in having white dotted warts more or less evenly distributed on the entire wine-red perigone lobe instead of having sulphur-yellow dotted warts on blood-red perigone lobes (Banziger et al. 2000). All members of the genus *Sapria* are distinctive and narrowly host-specific plants belonging to the family Rafflesiaceae. The typical form of *Sapria himalayana* is rare throughout its range from northeastern India, southwestern China, Thailand, Myanmar, to Vietnam. Being a member of the holoparasitic plant family, Rafflesiaceae, and growing on various species of *Tetrastigma* (Vitaceae), it is a little-understood species (Wu & Raven 2003; Nikolov & Davis 2017; Tran et al. 2018; Tanaka et al. 2019; Syiemiong et al. 2022). In India, the typical species of *Sapria* was first reported from the tropical wet evergreen forests of the Mishmi Hills of Lohit District by William Griffith in 1847 and later from the Aka Hills in Kameng District by Norman Loftus Bor in 1938 (Andreas & Jis 2014; Borah & Ghosh 2018). This species is the largest root parasitic angiosperm, having a host-specific relationship with the plant genus *Tetrastigma* of the Vitaceae family.

*Tetrastigma bracteolatum* (Wallich) Planchon and *T. serrulatum* (Roxb.) Planchon have been reported as the host plant of *Sapria* in Namdapha Park (Arunachalam et al. 2004; Borah & Ghosh 2018). The members of the Rafflesiaceae have a reduced vegetative body among all angiosperms (Nikolov et al. 2014). *Sapria* is well distinguished from the other two genera of Rafflesiaceae, *Rafflesia* R.Br. ex Thomson and *Rhizanthus* Dumort by the presence of 10 perianth lobes in two whorls (Meijer 1997; Nikolov & Davis 2017; Tanaka et al. 2019).

At present, the genus *Sapria* consists of four species, viz., *S. himalayana* Griffith, which has the widest distribution range among them (Wu & Raven 2003; Bendiksby et al. 2010; Ahmad et al. 2020; Syiemiong et al. 2022), with one form, *S. himalayana* Griffith f. *albovinosa* Banziger & Hansen (Banziger et al. 2000), endemic to northeastern Thailand, *S. poilanei* Gagnep, endemic to Cambodia (Gagnepian 1941), *S. ram* H Banziger & B Hansen, endemic to Thailand (Banziger & Hansen 1997), and *S. myanmerensis* Nob. Tanaka, Nagam, Tagane & M.M. Aung, endemic to Myanmar

(Tanaka et al. 2019). *Sapria himalayana* has lost about 44% of conserved genes in Eurosids that are enriched for functions like photosynthesis, plastid organisation, defence, stress response, and nutrient assimilation. It has also gained some genes from its hosts through horizontal gene transfer, showing extreme genome remodelling under parasitism (Cai et al. 2021).

In Rafflesiaceae, previous studies on the bud development, growth rate, mortality, flower phenology, life history and autecology have been conducted for several species of *Rafflesia*, where only a few were found for *Sapria* and none for *Rhizanthus*. Hidayati et al. (2006) and Nais (2001) studied *Rafflesia patma*, *R. keithii*, and *R. pricei* for their similar rapid later-stage bud growth and low bloom success. Sofiyanti et al. (2007) reconstructed the life cycle of *Rafflesia hasseltii*, which was redrawn from Nais (2001). Galindon et al. (2016) and Tolod et al. (2020) reported a new species of *Rafflesia*, namely *R. consueloae* and studied its first flower and fruit development and life history. Susatya (2020) reported the growth of the flower bud, life history and population structure of *R. arnoldii*. Recent work by Rambey et al. (2023) reported on the population and ecology of the endangered *R. meijeri* in Indonesia. Also, Wee et al. (2024) reported the bud development, flower phenology and life history of *R. cantleyi*. The earlier works on ecology, status, and conservation of *Sapria himalayana* Griffith, was done by Elliot (1992). Also, Arunachalam et al. (2004) reported on the population and conservation of *S. himalayana* Griffith, in Namdapha National Park, Arunachal Pradesh, India.

Currently, there is little information regarding the studies on the flower bud growth of *S. himalayana* Griffith f. *albovinosa* Banziger & Hansen (2000), even though it is essential for conservation purposes. The main objectives of this study were to observe the flower bud growth at different developmental stages of *S. himalayana* f. *albovinosa* and the change of its population structure with respect to the growth of the flower bud.

## MATERIALS AND METHODS

### Study area

The study was conducted in the subtropical evergreen rainforest of Machi Village, situated under the jurisdiction of Tengenoupal District, Manipur, northeastern India. The site is located at a latitude of 24.504° N and a longitude of 94.143° E with an elevation of about 1,477 m. Machi Village is about 62 km away from the capital city, Imphal,

of the state of Manipur. The motorable road to the village is in poor condition, with several potholes and uneven surfaces and is also prone to landslides during the monsoon season, which could completely block the way. The present research site is located in a remote area of the village, also there is no motorable road to the site as shown in Image 1. The village is situated in the Indo-Myanmar biodiversity hotspot, which is a region rich in floral and faunal diversity, and harbours numerous interesting and endangered species. The vicinity of the village is inhabited by the Maring Tribe, one of the indigenous tribes of Manipur, which has rich ethnic cultures and traditions. The people of the village practice jhumming cultivation as their main source of income. The entire region of the village crossing the International Indo-Myanmar Road harbours the tropical rainforest to sub-tropical evergreen, and deciduous forests. It is also an ideal habitat for various rare and endangered carnivores and birds.

#### Field data collection

The present study was carried out from July to November 2024 at the study site. *Sapria himalayana* f. *albovinosa* were found at the community evergreen forest of Machi Village, Manipur, and were monitored,

and the measurements of the plant buds and flowers were recorded for every observation at the study site. The observation was made at two-week interval for five months. The flower buds of *Sapria himalayana* f. *albovinosa* are found only on the roots of the host plant, *Tetrastigma*, unlike many species of *Rafflesia* found on both roots and climbing stems of the hosts. They are covered by the litter of the forest floors, making it difficult to observe at first sight. All the buds were discovered through thorough searching, scrutinising, and by removing forest litter in search of other buds whenever a bud was first detected at the study site. As such, this may prove to be an inevitable study limitation and may underestimate the actual population. While there were cases in which some undetected below-ground buds escaped the initial observation, when they grew bigger and surfaced above ground later, they were eventually added to the monitoring system (Wee et al. 2024). The diameters of *Sapria* buds and flowers were measured by the widest diameter length (Elliot 1992). The observation of this study was limited to the visible structure of bud and flower developmental stages of *Sapria himalayana* f. *albovinosa* (Susatya 2020). Each bud and flower was vertically photographed, and its diameter was measured at every observation (Nais 2001; Kamal et al. 2022). The

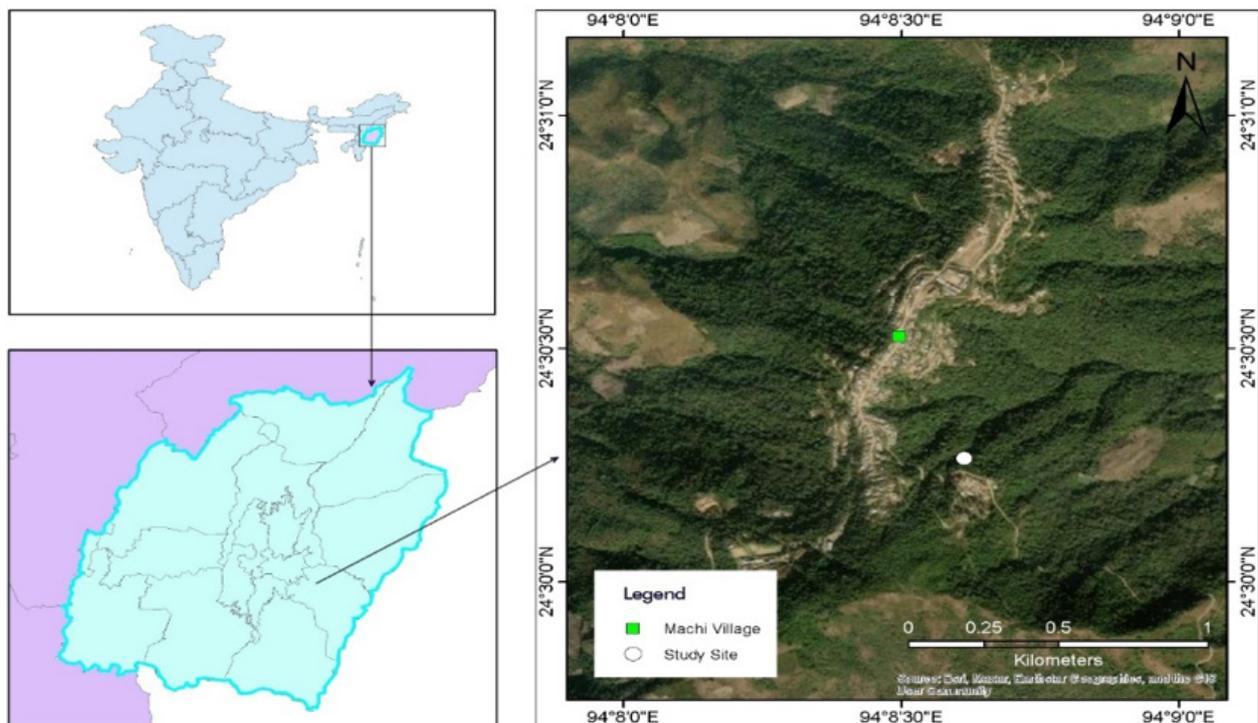


Image 1. A map showing the study area in the sub-tropical evergreen forest of Machi Village, Manipur, northeastern India. The white circle represents the location of *Sapria himalayana* f. *albovinosa*.

bud development was then categorized into different size classes for further analysis, following the methods by Nais (2001), Susatya (2020), and Tolod et al. (2020). The dead buds of all bud diameter sizes were recorded for every observation. Then the mortality rate of the *Sapria* buds was calculated by the formula given below (Nowak et al. 2004).

$$\text{Mortality rate} = \frac{\text{Total number of dead buds}}{\text{Total number of buds recorded}} \times 100$$

The abiotic parameters, such as air temperature, wind speed, humidity, soil temperature, soil moisture, soil pH, and light intensity, at the present study site were recorded using a thermometer and a 4-in-1 soil tester.

### Statistical analysis

A non-linear regression analysis was conducted to obtain the exponential growth model equation and coefficient of determination ( $R^2$ ) for the observed bud diameters to demonstrate a J-shaped bud growth curve. Also, correlation analysis was performed to examine the relationships between environmental factors such as ambient temperature, humidity, wind speed, soil pH, soil temperature, light intensity, and mortality of the buds for a five-month observation. A chi-square test was conducted to assess the bud stage distribution shifts across time of the observation. All statistical analyses were performed using SPSS version 26.

## RESULTS AND DISCUSSIONS

### Flower bud growth of *Sapria himalayana* f. *albovinosa*

The present study showed that the life cycle of *Sapria himalayana* f. *albovinosa* is complex, as most members of Rafflesiaceae have two parts – the invisible and visible parts (Hidayati et al. 2006; Nais 2001; Kamal et al. 2022). The invisible part includes the inoculation and germination of *Sapria*'s seed occurring inside its host plant roots, whereas the visible part is the emergence of the flower bud, mature bud, and anthesis. The visible part consists of several flower buds developing and is also the only plant structure that is exposed to external environmental factors. Therefore, flower buds at different sizes exhibited different growth rates and developmental stages. The observation limited to only the visible parts of the life cycle resulted in six different development stages of the flower, which were then categorized into the copule, copule-bract transition (CBT), bract, bract-perigone transition (BPT), perigone and anthesis stages (Image 2). Copule, bract

and perigone stages were defined by 80–100% of the images of vertically photographed bud respectively covered by copule, bract, and perigone structures. A bud was categorized into CBT, if it grew between copule and bract stages, and the coverage of the images of the photographed bud by the bract reached 40% to 80%. Meanwhile, a bud was grouped into BPT, if it grew between the bract and the perigone stages, with coverage of the images of the photographed bud by the perigone reaching 40% to 80%. Any bud with less than 40% of the coverage by either bract or perigone was also categorised into either copule or bract stages (Susatya 2020). The anthesis stage occurred once during the observation at the bud's diameter range of 16–20 cm. The observation resulted that the diameter range of copule, CBT, bract, BPT, and perigone stages respectively were 0.3–2.3 cm, 2.4–4.0 cm, 4.1–5.5 cm, 5.6–6.1 cm, and 6.2–8.1 cm (Table 1). The growth development of *Sapria*'s bud was not in a discrete pattern, where one stage was replaced completely by the next stage. The same growth development was also observed in the typical species of *Sapria* (Elliot 1992). It consisted of a series of overlapping development stages, where before one stage was complete, the following stage had already developed. It was a basic reason why transition stages were introduced in this research. The first visible structure was copule, which was basically the bark of the host plant root covering the actual *Sapria* structure. The first visible structure of *S. himalayana* f. *albovinosa* at Machi had the diameter range of 0.3–2.3 cm (Table. 1). The start of the development of the inner structures of *Sapria* was still unknown though it is needed to be studied whether all the inner structures had been developed in the copule stage or not. The inner structures in the species of *Rafflesia* had already developed in the copule stage while observing its dead bud of 6 cm diameter (Susatya 2020). As the bud grew, the upper copule started to crack to allow the first true structure of the *Sapria* or bract to be visible. Bract was originally pastel

**Table 1. The range of diameters of buds and flowers according to its stages.**

Name of bud stages	Range of bud diameter (cm)
Copule	0.3–2.3 cm
Copule-bract transition (CBT)	2.4–4.0 cm
Bract	4.1–5.5 cm
Bract perigone transition (BPT)	5.6–6.1 cm
Perigone	6.2–8.1 cm
Anthesis	16–20 cm

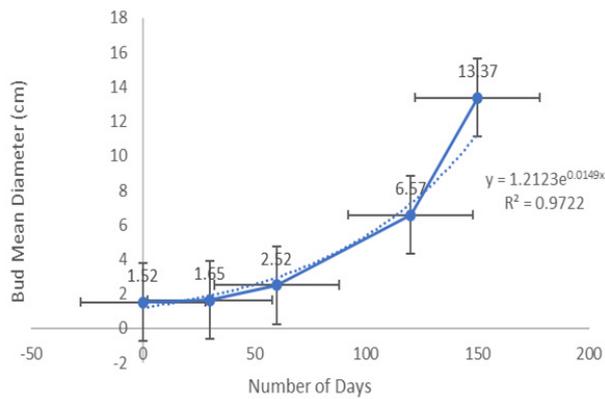


Figure 1. Growth curve of *Sapria himalayana* f. *albovinosa* during the five-month observation.

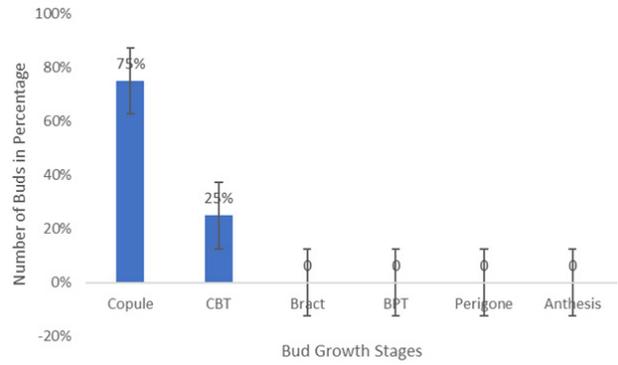


Figure 2. The population structure of *Sapria himalayana* f. *albovinosa* in August.

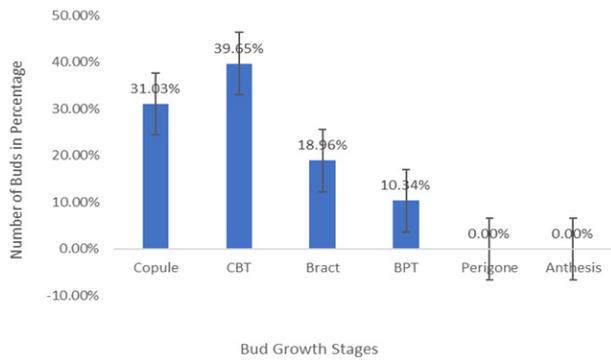


Figure 3. The population structure of *Sapria himalayana* f. *albovinosa* in September.

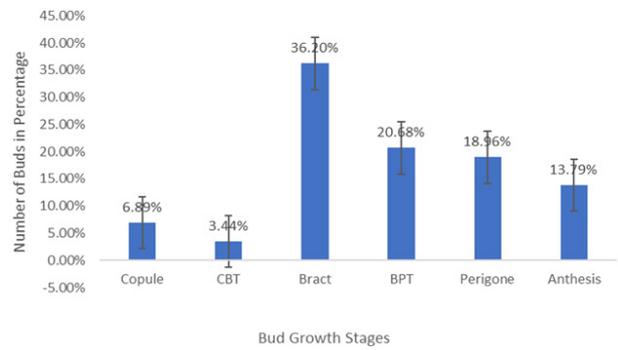


Figure 4. The population structure of *Sapria himalayana* f. *albovinosa* in October.

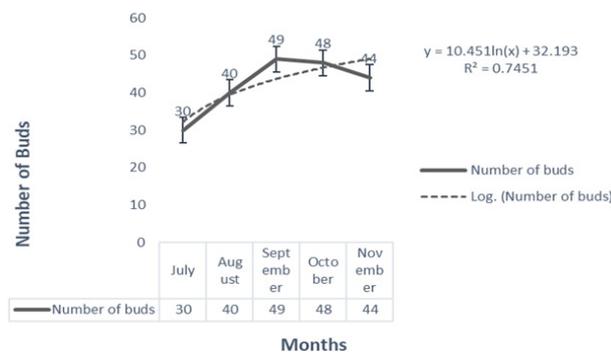


Figure 5. Population growth curve showing number of buds with respect to the month of observation.

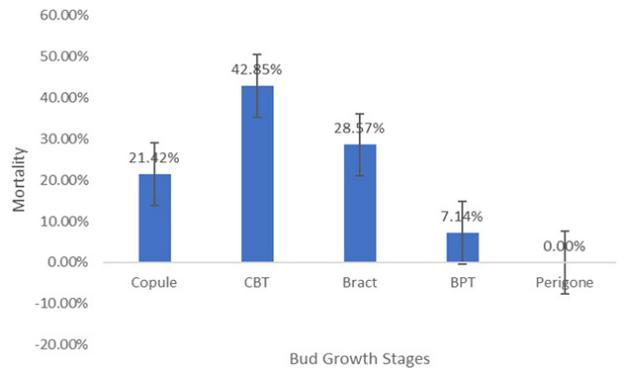


Figure 6. The bud mortality rate of *Sapria himalayana* f. *albovinosa* according to its growth development stage.

pink and white colour, but eventually turned black as it grew older. The bract was gradually replaced by a pale pink perigone stage consisting of buds with a diameter range of 6.2–8.1 cm (Image 2). The bract consisted of two series of five imbricate and whorled scales (Elliot 1992). The pinkish perigone lobes of the bud indicated anthesis to occur within 3–4 days. The field observation

showed that when the upper layer of the perigone lobe was slightly raised, then the anthesis would take place within 2–3 days, and lasted between 4–5 days. All flower structures decomposed within a month after flowering. The column was the only female structure that did not decompose and further developed into mature fruit (Elliot 1992). As the flowers opened, they emitted an



**Image 2.** The flower bud development stages of *Sapria himalayana* f. *albovinosa*: A—copule stage | B—copule-bract transition stage (CBT), copule (cp) gradually replaced by bracts (br) | C—bract stage, a stage where a visible bud is a bract, a similar structure to a sepal | D—bract-perigone transition stage (BPT), a stage where bracts are still largely recognized (br) and gradually replaced by perigone (pr) | E—perigone stage, a visible bud at this stage is all covered by the perigone, a similar structure to a petal | F—Anthesis stage, a flower fully bloomed. © K. Shamran Maring.

odour similar to that of rotting meat, which lasted for 2–3 days. Within 3–4 days after opening, flowers started to darken and eventually turned black. As the flowers turned black, all the plant structures shrank. The base of male flowers and their attachment to the host shriveled rapidly and eventually detached from their host roots. In female flowers, the perigone tube and lobes, diaphragm and disk shriveled as in males, but the column, ovary and surrounding tissues at the base of the column remained alive (Elliot 1992). The base of the perigone tube swelled and remained white externally for about two months after flower opening. This structure constituted the fruit of *S. himalayana* f. *albovinosa*, though a detailed study on the fruits and seeds dispersal is required. The growth rate of flower buds at earlier stages was found to be very slow, while the buds at older stages showed higher growth rates. Therefore, the growth curve of the flower bud showed a typical J-shaped growth curve or an exponential growth curve (Figure 1).

The non-linear regression equation  $y = 1.2123 e^{0.0149x}$  and  $R^2 = 0.9722$  was obtained by plotting the

bud mean diameter across the number of observed days. The bud diameter grows by about 1.49% of its current size in every additional day. This means the bud diameter doubles roughly every 46–47 days under the observed growth pattern. Therefore, on the initial day of observation, the bud starts at about 1.5 cm in diameter, then, by about 150 days, they grow to above 13 cm in diameter, matching the measured data points (Figure 1).

The present study showed that *S. himalayana* Griffith f. *albovinosa* found at Machi comparatively has crateriform and bilobed or multilobed rammenta apices in male and female flowers, respectively, while the Vietnamese taxon is also bilobed or multilobed (female) or crateriform (male) (Maring & Pinokiyo 2024). The diameter of the crest disk and aperture of the diaphragm for the Machi individuals are 3.4–3.5 cm and 1.9–2.0 cm, respectively. The *S. himalayana* Griffith f. *albovinosa* from Machi Village has a comparatively larger floral span (13–20 cm diameter) than the *S. himalayana* Griffith f. *albovinosa* (11–16 cm diameter) (Banziger et al. 2000), *S. myanmerensis* (10 cm) (Tanaka et al. 2019), *S. poilanei*

**Table 2. The abiotic factors of *Sapria himalayana* f. *albovinosa* in Machi evergreen rainforest, Manipur.**

Abiotic factors	July	August	September	October	November
Mean air temperature (°C)	23	23	22	20	19
Mean humidity (%)	80	84	86	72	67
Mean wind speed (m/s)	1.60	1.66	1.66	1.66	1.66
Mean soil temperature (°C)	19	24	21	20	18
Mean soil pH	6.5	6.5	6.0	5.5	5.5
Mean soil moisture	Wet+	Wet	Wet	Normal	Dry
Mean light intensity (lux)	90	90	100	100	100

(6.5–12 cm), and *S. ram* (5.5–11 cm) (Banziger & Hansen 1997).

#### Population structure of *Sapria himalayana* f. *albovinosa*

The population status of *S. himalayana* f. *albovinosa* is very small when compared to the other higher plants. Also, the population of this endemic infraspecific taxon is much smaller than that of the typical species of *Sapria* (Elliot 1992). The initial observation in the month of July showed that the total population is 100% dominated by the copula stage. At the second observation in August, the population structure was 75% copule and 25% copule bract transition (CBT) Stage (Figure 2). Meanwhile, buds at CBT were fewer due to the changes in the flower bud development stage. Larger stages, such as bract and BPT, were observed respectively with 18.96% and 10.34% in September (Figure 3). Within three months, the population structure was significantly changed due to the mortality, new recruitment, and growth of buds from one stage to the next growth development stages. The population structure of this period was shifted toward bract and BPT. During the next observation, in the month of October, the population structure interestingly exhibited all six bud stages, where the bract stage dominated it with 36.20% (Figure 4). The pattern of population structure appeared to be opposite to the initial one, where the perigone and anthesis stages were exhibited with 18.96% and 13.79%, respectively. In the last month's observations, the population structure was dominated by full bloom/anthesis and after-blooming flowers. The flower buds were found at their full bloom stage in October to December, but the optimal flowering season is from late October to late November. During the initial period of research, the number of flower buds increased gradually due to the new recruitment of the buds, while the population size decreased in the later months of observation. This condition could be due to insufficient nutrients for the population of the flower bud to maintain its viability and complete its life cycle

(Figure 5). A chi-square test revealed a significant shift in bud stage distribution across time ( $X^2 = 202.997$ ,  $df = 20$ ,  $p = 2.87 \times 10^{-32}$ ), this indicates that the proportion of buds at different developmental stages varied markedly with time and did not remain constant over time.

In the five-month observation, a total of 58 flower buds were recorded, out of which 28 buds were recruits and 14 buds died without reaching maturity. The causes of the bud mortality were fungal infections and injuries from anthropogenic interventions, where the injured parts of flower buds were immediately followed by a rotting process that led to the bud mortality. All losses occurred at buds belonging to the copule, CBT, bract, and BPT with copule-bract transition (CBT) stage showing the highest mortality rate of 42.85% (Figure 6). The population structure of *S. himalayana* f. *albovinosa* showed a mortality rate of 24.13%, which is much lower when compared to the mortality rate 40% of the typical species (Elliot 1992). Although the mortality rate is much lower than that of the typical species of *Sapria*, the total population status of this endemic taxon is very small, indicating an alarming signal concerning the future population of *S. himalayana* f. *albovinosa*.

#### Abiotic factors of *Sapria himalayana* f. *albovinosa* habitat

The ambient temperature of the study site was found to decrease from 23°C (July) to 19°C (November), which is due to the onset of the winter season. The air humidity ranges 67–86 %, and the light intensity was found to be very low, ranging 90–100 lux, probably be caused by the dense canopy cover (Table 2). The high air humidity and low light intensity play a vital role in the existence of *S. himalayana* f. *albovinosa* because the plant preferably grows on the understory of forest floors. Also, in the present study, the flower buds showed higher mortality rates as the light intensity increased. There is a moderately strong positive correlation between light intensity and bud mortality, with a correlation value

( $r$ ) = 0.6123, indicating that higher mortality rates are generally associated with higher light intensity. Whereas, the soil pH showed a moderate negative correlation ( $r$  = -0.326) with the bud mortality. In Table 2, the soil pH ranges 5.5–6.5, and it is classified as acidic, while the soil temperature ranges 18–24 °C. The soil moisture showed very wet conditions in July, which could be due to the monsoon season, while the soil was dry in November, which could be due to the winter season. Therefore, the abiotic parameters of the environment play an important role in the growth of flower buds as well as in the population dynamics of this rare endemic taxon.

## CONCLUSION

The flower bud growth of *Sapria himalayana* f. *albovinosa* has six developmental stages consisting of the copule, CBT, bract, BPT, perigone, and anthesis stages. More detailed studies on the growth rate, mortality rate and population structure of this rare parasitic plant are required to determine the fate of the young flower buds and to estimate its complete life cycle. The findings from this study are useful in intensifying the knowledge of this rare parasitic plant that is becoming vulnerable and is on the brink of extinction. Due to its rarity, ongoing habitat loss, vandalism of existing colonies, and high degree of host specificity, various conservation actions are required to protect this taxon. Hence, the information on the growth of buds, different flower bud developmental stages, and population status is vital for taxonomic studies, field monitoring, and conservation purposes.

## REFERENCES

- Ahmad, A., A. Kumar, G.S. Rawat & G.V. Gopi (2020). Recent record of a threatened holoparasitic plant *Sapria himalayana* Griff. In Mehao Wildlife Sanctuary, Arunachal Pradesh, India. *Journal of Threatened Taxa* 12(10): 16399–16404. <https://doi.org/10.11609/jott.5168.12.10.16399-16401>
- Andreas, H. & S. Jis (2014). *Sapria himalayana* Griffith, an endangered species from the Mishmi Hills, Dilbang Valley, Arunachal Pradesh. *Indian Forester* 140(4): 433–434.
- Arunachalam, A., D. Adhikari, R. Sarmah, M. Majumder & M.L. Khan (2004). Population and conservation of *Sapria himalayana* Griffith. in Namdapha National Park, Arunachal Pradesh, India. *Biodiversity and Conservation* 13: 2391–2397. <https://doi.org/10.1023/B:BOIC.0000048488.94151.f8>
- Bänziger, H. & B. Hansen (1997). Unmasking the real identity of *Sapria poilanei* Gagnepain emend., and description of *Sapria ram* sp. n. (Rafflesiaceae). *Natural History Bulletin of the Siam Society*. 45: 149–170.
- Bänziger, H., B. Hansen & K. Kreetiyutanont (2000). A new form of the hermit's spittoon, *Sapria himalayana* Griffith f. *albovinosa* Bänziger and Hansen f. nov. (Rafflesiaceae), with notes on its ecology. *Natural History Bulletin Siam Society* 48: 213–219.
- Bendiksby, M., T. Schumacher, G. Gussarova, J. Nais, M. Kamarudin, S. Nery, D. Madulid, S.A. Smith & T.J. Barkman (2010). Elucidating the evolutionary history of the southeast Asian, holoparasitic, giant-flowered Rafflesiaceae: Pliocene vicariance, morphological convergence and character displacement. *Molecular Phylogenetic and Evolution* 57(2): 620–633. <https://doi.org/10.1016/j.ympev.2010.08.005>
- Borah, D. & D. Ghosh (2018). *Sapria himalayana*: The Indian cousin of the world's largest flower. *Resonance* 23(4): 479–489. <https://doi.org/10.1007/s12045-018-0637-8>
- Cai, L., B.J. Arnold, Z. Xi, D.E. Khost, N. Patel, C.B. Hartmann, S. Manickam, S. Sasirat, L.A. Nikolov, S. Mathews, T.B. Sackton & C.C. Davis (2021). Deeply altered genome architecture in the endoparasitic flowering plant *Sapria himalayana* Griff. (Rafflesiaceae). *Current Biology* 31(5): 1002–1011. <https://doi.org/10.1016/j.cub.2020.12.045>
- Elliot, S. (1992). Status, ecology and conservation of *Sapria himalayana* Griffith. (Rafflesiaceae) in Thailand. *Journal of Wildlife Thailand* 2(1): 44–52.
- Galindon, J.M.M., P.S. Ong & E.S. Fernando (2016). *Rafflesia consueloae* (Rafflesiaceae), the smallest among giants; a new species from Luzon Island, Philippines. *PhytoKeys* (61): 37–46. <https://doi.org/10.3897/phytokeys.61.7295>
- Gagnepain, F. (1941). U neespece nouvelle d'un genre monotype: *Sapria*. *Notulae systematicae* (Paris) 9: 144–145. <https://biostor.org/reference/266807>
- Hidayati, S.N., W. Meijer, J.M. Baskin & J.L. Walck (2006). A contribution to the life history of the rare Indonesian holoparasite *Rafflesia patma* (Rafflesiaceae). *Biotropica* 32(3): 408–414. <https://doi.org/10.1111/j.1744-7429.2000.tb00487.x>
- Kamal, S.H.S., M.N. Suratman, S. Khamis, A.N.N. Hassan & M.S. Mohammad (2022). Growth rate, mortality rate and life cycle of *Rafflesia azlanii* and *R. cantleyi* in Belum-Temenggor Forest Complex, Perak, Malaysia. *Sains Malaysiana* 51(4): 943–957. <https://doi.org/10.17576/jsm-2022-5104-01>
- Maring, K.S. & A. Pinokoyo (2024). A taxonomic note on *Sapria himalayana* f. *albovinosa* Bänziger and B. Hansen from India. *Indian Forester* 150(3): 292–295. <https://doi.org/10.36808/if/2024/v150i3/169401>
- Meijer, W. (1997). Rafflesiaceae. In: Kalkman, C., D.W. Kirkup, H.P. Nootboom, P.F. Stevens & W.J.J.O. de Wilde (eds.). *Flora Malesiana*, Series I: Spermatophyta, 13: 1–42. Rijksherbarium/HortusBotanicus, Leiden, The Netherlands.
- Nais, J. (2001). *Rafflesia* of the world. Sabah Park in association with Natural History Publications (Borneo) Sdn. Bhd., Kota Kinabalu Smith RL. 1986. *Elements of Ecology*. Harper & Row Publishers. New York, 243 pp.
- Nikolov, L.A. & C.C. Davis (2017). The big, the bad, and the beautiful: Biology of the world's largest flowers. *Journal of Systematics and Evolution* 55(6): 516–524. <https://doi.org/10.1111/jse.12260>
- Nikolov, L.A., P.B. Tomlinson, S. Manickam, P.K. Endress, E.M. Kramer & C.C. Davis (2014). Holoparasitic Rafflesiaceae possess the most reduced endophytes and yet give rise to the world's largest flowers. *Annals of Botany* 114(2): 233–242. <https://doi.org/10.1093/aob/mcu114>
- Nowak, D.J., M. Kuroda & D.E. Crane (2004). Tree mortality rates and tree population projections in Baltimore, Maryland, USA. *Urban Forestry & Urban Greening* 2(3): 139–147. <https://doi.org/10.1078/1618-8667-00030>
- Rambey, R., N. Saputra, I.F. Rambe, B. Nopandry, S. Zunaidi, E.L. Christy, T. Setiawan, Y. Affuddin & A. Hartanto (2023). Population and autecology of the endangered *Rafflesia meijeri* in Batang Gadis National Park, Indonesia. *Biodiversitas* 24(3): 1845–1852. <https://doi.org/10.13057/biodiv/d240360>
- Sofiyanti, N., K. Mat-Salleh, P. Puruwanto & E. Syahputra (2007). The Note on Morphology of *Rafflesia hasseltii* Surigar from Bukit Tiga Puluh National Park, Riau. *Biodiversitas* 8(4): 257–261. <https://doi.org/10.13057/biodiv/d080402>

- Susatya, A. (2020).** The growth of flower bud, life history, and population structure of *Rafflesia arnoldii* (Rafflesiaceae) in Bengkulu, Sumatra, Indonesia. *Biodiversitas* 21(2): 792–798. <https://doi.org/10.13057/biodiv/d210247>
- Syiemiong, P., S.S. Chaturvedi, T. Arbenz & T. Tamaş (2022).** A note on *Sapria himalayana* (Griffith 1844) (Rafflesiaceae) from Jaintia Hills (Meghalaya, India). *Biodiversity Journal* 13(1): 73–78. <https://doi.org/10.31396/Biodiv.Jour.2022.13.1.73.78>
- Tanaka, N., H. Nagamasu, S. Tagane, M.M. Aung, A.K. Win & P.P. Hnin (2019).** Contributions to the flora of Myanmar IV: A new species and a newly recorded taxon of the genus *Sapria* (Rafflesiaceae). *Taiwania* 64(4): 357–362. <https://doi.org/10.6165/tai.2019.64.357>
- Tolod, J.R., J.M.M. Galindon, R.R. Atienza, M.V. Duya, E.S. Fernando & P.S. Ong (2020).** Flower and Fruit Development and Life History of *Rafflesia consueloae* (Rafflesiaceae). *Philippine Journal of Science* (150): 321–334. <https://doi.org/10.56899/150.sl.23>
- Trần, H.D., H.T. Lu’u, Q.D. Nguyen, H.C. Nguyen, P. Athen & K.M. Wong (2018).** Identification, sexual dimorphism and aspects of the natural history of *Sapria himalayana* (Rafflesiaceae) on Vietnam’s Lang Biang Plateau. *Botanical Studies* 59: 29. <https://doi.org/10.1186/s40529-018-0243-9>
- Wee, S.K., S.B. Tan, S.H. Tan & B.K.B. Lee (2024).** Bud development, flower phenology and life history of holoparasitic *Rafflesia cantleyi*. *Journal of Plant Research* (137): 423–443. <https://doi.org/10.1007/s10265-024-01522-7>
- Wu, Z. & P.H. Raven (2003).** *Flora of China, No.5 Science Press*. Beijing and Missouri Botanical Garden Press, St. Louis, 505 pp.
- Author details:** K. SHAMRAN MARING is currently pursuing PhD degree at Dhanamanjuri University, Manipur, working on the angiosperm flora of Tengnoupal District, Manipur. Her research focuses on plant taxonomy, primarily concentrating on floristic, ecological studies, and biodiversity conservation. PROF. ATHOKRAM PINOKIYO is currently working as the Head, Department of Botany, Dhanamanjuri University, Imphal, Manipur, and she has got a teaching experience for about 15 years. Her keen work is on taxonomy, diversity, ecology, and conservation of lichens (including Angiosperms). She has got a research experience of more than 24 years in the field of taxonomy (angiosperms and lichens).
- Author contributions:** KSM: conceptualised the study design, carried out the field surveys, data collection, data handling, data curation, photography, visualisation, statistical analysis, writing, review and editing of the manuscript. AP: supervised the research work and revision of the manuscript. Both authors read and agreed to the final manuscript.
- Ethics and field compliance statement:** The field research was conducted with prior permission from the relevant forest and community authorities. Prior informed consent was obtained from the Machi Village Authority, a Maring community, inhabiting the study area, with approval letter no. MCI-VA/85/2-2025 dated 03 October 2025. All field activities were conducted in accordance with the local customs, cultural values, community protocols, traditional knowledge and the ethical standards for research in the Machi Village community forest. No endangered or protected species were harmed or removed from their natural habitats during the course of this research. All observations and data collection were conducted in a non-destructive and minimally invasive manner consistent with best ecological research practices.
- Acknowledgements:** The authors are grateful to the Department of Botany, Dhanamanjuri University, Imphal, Manipur, for helpful advice and permission given for this study. K SHAMRAN MARING express gratitude to the National Fellowship for Higher Education for Schedule Tribe Students (NFST), bearing Award No. 202223-NFST-MAN-02118, Ministry of Tribal Affairs, Government of India, for the financial assistance provided. Also, thanks are due to the local people of Machi Village who gave a helpful hand in the field survey of the present study.





Mr. Jatishwor Singh Irungbam, Biology Centre CAS, Branišovská, Czech Republic.  
Dr. Ian J. Kitching, Natural History Museum, Cromwell Road, UK  
Dr. George Mathew, Kerala Forest Research Institute, Peechi, India  
Dr. John Noyes, Natural History Museum, London, UK  
Dr. Albert G. Orr, Griffith University, Nathan, Australia  
Dr. Sameer Padhye, Katholieke Universiteit Leuven, Belgium  
Dr. Nancy van der Poorten, Toronto, Canada  
Dr. Kareen Schnabel, NIWA, Wellington, New Zealand  
Dr. R.M. Sharma, (Retd.) Scientist, Zoological Survey of India, Pune, India  
Dr. Manju Siliwal, WILD, Coimbatore, Tamil Nadu, India  
Dr. G.P. Sinha, Botanical Survey of India, Allahabad, India  
Dr. K.A. Subramanian, Zoological Survey of India, New Alipore, Kolkata, India  
Dr. P.M. Sureshan, Zoological Survey of India, Kozhikode, Kerala, India  
Dr. R. Varatharajan, Manipur University, Imphal, Manipur, India  
Dr. Eduard Vives, Museu de Ciències Naturals de Barcelona, Terrassa, Spain  
Dr. James Young, Hong Kong Lepidopterists' Society, Hong Kong  
Dr. R. Sundararaj, Institute of Wood Science & Technology, Bengaluru, India  
Dr. M. Nithyanandan, Environmental Department, La Ala Al Kuwait Real Estate. Co. K.S.C., Kuwait  
Dr. Himender Bharti, Punjabi University, Punjab, India  
Mr. Purnendu Roy, London, UK  
Mr. Saito Motoki, The Butterfly Society of Japan, Tokyo, Japan  
Dr. Sanjay Sondhi, TITLI TRUST, Kalpavriksh, Dehradun, India  
Dr. Nguyen Thi Phuong Lien, Vietnam Academy of Science and Technology, Hanoi, Vietnam  
Dr. Nitin Kulkarni, Tropical Research Institute, Jabalpur, India  
Dr. Robin Wen Jiang Ngiam, National Parks Board, Singapore  
Dr. Lionel Monod, Natural History Museum of Geneva, Genève, Switzerland.  
Dr. Asheesh Shivam, Nehru Gram Bharti University, Allahabad, India  
Dr. Rosana Moreira da Rocha, Universidade Federal do Paraná, Curitiba, Brasil  
Dr. Kurt R. Arnold, North Dakota State University, Saxony, Germany  
Dr. James M. Carpenter, American Museum of Natural History, New York, USA  
Dr. David M. Claborn, Missouri State University, Springfield, USA  
Dr. Kareen Schnabel, Marine Biologist, Wellington, New Zealand  
Dr. Amazonas Chagas Júnior, Universidade Federal de Mato Grosso, Cuiabá, Brasil  
Mr. Monsoon Jyoti Gogoi, Assam University, Silchar, Assam, India  
Dr. Heo Chong Chin, Universiti Teknologi MARA (UiTM), Selangor, Malaysia  
Dr. R.J. Shiel, University of Adelaide, SA 5005, Australia  
Dr. Siddharth Kulkarni, The George Washington University, Washington, USA  
Dr. Priyadarsanan Dharma Rajan, ATREE, Bengaluru, India  
Dr. Phil Alderslade, CSIRO Marine And Atmospheric Research, Hobart, Australia  
Dr. John E.N. Veron, Coral Reef Research, Townsville, Australia  
Dr. Daniel Whitmore, State Museum of Natural History Stuttgart, Rosenstein, Germany.  
Dr. Yu-Feng Hsu, National Taiwan Normal University, Taipei City, Taiwan  
Dr. Keith V. Wolfe, Antioch, California, USA  
Dr. Siddharth Kulkarni, The Hormiga Lab, The George Washington University, Washington, D.C., USA  
Dr. Tomas Ditrich, Faculty of Education, University of South Bohemia in Ceske Budejovice, Czech Republic  
Dr. Mihaly Foldvari, Natural History Museum, University of Oslo, Norway  
Dr. V.P. Uniyal, Wildlife Institute of India, Dehradun, Uttarakhand 248001, India  
Dr. John T.D. Caleb, Zoological Survey of India, Kolkata, West Bengal, India  
Dr. Priyadarsanan Dharma Rajan, Ashoka Trust for Research in Ecology and the Environment (ATREE), Royal Enclave, Bangalore, Karnataka, India

#### Fishes

Dr. Topiltzin Contreras MacBeath, Universidad Autónoma del estado de Morelos, México  
Dr. Heok Hee Ng, National University of Singapore, Science Drive, Singapore  
Dr. Rajeesh Raghavan, St. Albert's College, Kochi, Kerala, India  
Dr. Robert D. Sluka, Chiltern Gateway Project, A Rocha UK, Southall, Middlesex, UK  
Dr. E. Vivekanandan, Central Marine Fisheries Research Institute, Chennai, India  
Dr. Davor Zanella, University of Zagreb, Zagreb, Croatia  
Dr. A. Biju Kumar, University of Kerala, Thiruvananthapuram, Kerala, India  
Dr. Akhilesh K.V., ICAR-Central Marine Fisheries Research Institute, Mumbai Research Centre, Mumbai, Maharashtra, India  
Dr. J.A. Johnson, Wildlife Institute of India, Dehradun, Uttarakhand, India  
Dr. R. Ravinesh, Gujarat Institute of Desert Ecology, Gujarat, India

#### Amphibians

Dr. Sushil K. Dutta, Indian Institute of Science, Bengaluru, Karnataka, India  
Dr. Annemarie Ohler, Muséum national d'Histoire naturelle, Paris, France

#### Reptiles

Dr. Gernot Vogel, Heidelberg, Germany  
Dr. Raju Vyasa, Vadodara, Gujarat, India  
Dr. Pritpal S. Soorae, Environment Agency, Abu Dhabi, UAE.  
Prof. Dr. Wayne J. Fuller, Near East University, Mersin, Turkey  
Prof. Chandrashekhar U. Rivonker, Goa University, Taleigao Plateau, Goa, India  
Dr. S.R. Ganesh, Kalinga Foundation, Agumbe, India.  
Dr. Himansu Sekhar Das, Terrestrial & Marine Biodiversity, Abu Dhabi, UAE

**Journal of Threatened Taxa** is indexed/abstracted in Bibliography of Systematic Mycology, Biological Abstracts, BIOSIS Previews, CAB Abstracts, EBSCO, Google Scholar, Index Copernicus, Index Fungorum, JournalSeek, National Academy of Agricultural Sciences, NewJour, OCLC WorldCat, SCOPUS, Stanford University Libraries, Virtual Library of Biology, Zoological Records.

NAAS rating (India) 5.64

#### Birds

Dr. Hem Sagar Baral, Charles Sturt University, NSW Australia  
Mr. H. Byju, Coimbatore, Tamil Nadu, India  
Dr. Chris Bowden, Royal Society for the Protection of Birds, Sandy, UK  
Dr. Priya Davidar, Pondicherry University, Kalapet, Puducherry, India  
Dr. J.W. Duckworth, IUCN SSC, Bath, UK  
Dr. Rajah Jayapal, SACON, Coimbatore, Tamil Nadu, India  
Dr. Rajiv S. Kalsi, M.L.N. College, Yamuna Nagar, Haryana, India  
Dr. V. Santharam, Rishi Valley Education Centre, Chittoor Dt., Andhra Pradesh, India  
Dr. S. Balachandran, Bombay Natural History Society, Mumbai, India  
Mr. J. Praveen, Bengaluru, India  
Dr. C. Srinivasulu, Osmania University, Hyderabad, India  
Dr. K.S. Gopi Sundar, International Crane Foundation, Baraboo, USA  
Dr. Gombobaatar Sundev, Professor of Ornithology, Ulaanbaatar, Mongolia  
Prof. Reuven Yosef, International Birding & Research Centre, Eilat, Israel  
Dr. Taej Mundkur, Wetlands International, Wageningen, The Netherlands  
Dr. Carol Inskipp, Bishop Auckland Co., Durham, UK  
Dr. Tim Inskipp, Bishop Auckland Co., Durham, UK  
Dr. V. Gokula, National College, Tiruchirappalli, Tamil Nadu, India  
Dr. Arkady Lelej, Russian Academy of Sciences, Vladivostok, Russia  
Dr. Simon Dowell, Science Director, Chester Zoo, UK  
Dr. Mário Gabriel Santiago dos Santos, Universidade de Trás-os-Montes e Alto Douro, Quinta de Prados, Vila Real, Portugal  
Dr. Grant Connette, Smithsonian Institution, Royal, VA, USA  
Dr. P.A. Azeez, Coimbatore, Tamil Nadu, India

#### Mammals

Dr. Giovanni Amori, CNR - Institute of Ecosystem Studies, Rome, Italy  
Dr. Anwaruddin Chowdhury, Guwahati, India  
Dr. David Mallon, Zoological Society of London, UK  
Dr. Shomita Mukherjee, SACON, Coimbatore, Tamil Nadu, India  
Dr. Angie Appel, Wild Cat Network, Germany  
Dr. P.O. Nameer, Kerala Agricultural University, Thrissur, Kerala, India  
Dr. Ian Redmond, UNEP Convention on Migratory Species, Lansdown, UK  
Dr. Heidi S. Riddle, Riddle's Elephant and Wildlife Sanctuary, Arkansas, USA  
Dr. Karin Schwartz, George Mason University, Fairfax, Virginia.  
Dr. Lala A.K. Singh, Bhubaneswar, Orissa, India  
Dr. Mewa Singh, Mysore University, Mysore, India  
Dr. Paul Racey, University of Exeter, Devon, UK  
Dr. Honnavalli N. Kumara, SACON, Anaikatty P.O., Coimbatore, Tamil Nadu, India  
Dr. Nishith Dharaiya, HNG University, Patan, Gujarat, India  
Dr. Spartaco Gippoliti, Socio Onorario Società Italiana per la Storia della Fauna "Giuseppe Altobello", Rome, Italy  
Dr. Justus Joshua, Green Future Foundation, Tiruchirappalli, Tamil Nadu, India  
Dr. H. Raghuram, Sri S. Ramasamy Naidu Memorial College, Virudhunagar, Tamil Nadu, India  
Dr. Paul Bates, Harison Institute, Kent, UK  
Dr. Jim Sanderson, Small Wild Cat Conservation Foundation, Hartford, USA  
Dr. Dan Challender, University of Kent, Canterbury, UK  
Dr. David Mallon, Manchester Metropolitan University, Derbyshire, UK  
Dr. Brian L. Cypher, California State University-Stanislaus, Bakersfield, CA  
Dr. S.S. Talmale, Zoological Survey of India, Pune, Maharashtra, India  
Prof. Karan Bahadur Shah, Budhanilakantha Municipality, Kathmandu, Nepal  
Dr. Susan Cheyne, Borneo Nature Foundation International, Palangkaraja, Indonesia  
Dr. Hemanta Kafley, Wildlife Sciences, Tarleton State University, Texas, USA

#### Other Disciplines

Dr. Aniruddha Belsare, Columbia MO 65203, USA (Veterinary)  
Dr. Mandar S. Paingankar, University of Pune, Pune, Maharashtra, India (Molecular)  
Dr. Jack Tordoff, Critical Ecosystem Partnership Fund, Arlington, USA (Communities)  
Dr. Ulrike Streicher, University of Oregon, Eugene, USA (Veterinary)  
Dr. Hari Balasubramanian, EcoAdvisors, Nova Scotia, Canada (Communities)  
Dr. Rayanna Hellem Santos Bezerra, Universidade Federal de Sergipe, São Cristóvão, Brazil  
Dr. Jamie R. Wood, Landcare Research, Canterbury, New Zealand  
Dr. Wendy Collinson-Jonker, Endangered Wildlife Trust, Gauteng, South Africa  
Dr. Rajeshkumar G. Jani, Anand Agricultural University, Anand, Gujarat, India  
Dr. O.N. Tiwari, Senior Scientist, ICAR-Indian Agricultural Research Institute (IARI), New Delhi, India  
Dr. L.D. Singla, Guru Angad Dev Veterinary and Animal Sciences University, Ludhiana, India  
Dr. Rupika S. Rajakaruna, University of Peradeniya, Peradeniya, Sri Lanka  
Dr. Bahar Baviskar, Wild-CER, Nagpur, Maharashtra 440013, India

#### Reviewers 2021–2023

Due to paucity of space, the list of reviewers for 2021–2023 is available online.

The opinions expressed by the authors do not reflect the views of the Journal of Threatened Taxa, Wildlife Information Liaison Development Society, Zoo Outreach Organization, or any of the partners. The journal, the publisher, the host, and the partners are not responsible for the accuracy of the political boundaries shown in the maps by the authors.

Print copies of the Journal are available at cost. Write to:  
The Managing Editor, JoTT,  
c/o Wildlife Information Liaison Development Society,  
3A2 Varadarajulu Nagar, FCI Road, Ganapathy, Coimbatore,  
Tamil Nadu 641006, India  
ravi@threatenedtaxa.org & ravi@zooreach.org



OPEN ACCESS



The Journal of Threatened Taxa (JoTT) is dedicated to building evidence for conservation globally by publishing peer-reviewed articles online every month at a reasonably rapid rate at [www.threatenedtaxa.org](http://www.threatenedtaxa.org). All articles published in JoTT are registered under [Creative Commons Attribution 4.0 International License](https://creativecommons.org/licenses/by/4.0/) unless otherwise mentioned. JoTT allows unrestricted use, reproduction, and distribution of articles in any medium by providing adequate credit to the author(s) and the source of publication.

ISSN 0974-7907 (Online) | ISSN 0974-7893 (Print)

February 2026 | Vol. 18 | No. 2 | Pages: 28262–28454

Date of Publication: 26 February 2026 (Online &amp; Print)

DOI: 10.11609/jott.2026.18.2.28262-28454

[www.threatenedtaxa.org](http://www.threatenedtaxa.org)

## Articles

**Floristic composition and conservation significance of vascular plants in Kalatop-Khajjiar Wildlife Sanctuary, Himachal Pradesh, India**

– Sumit, Gulshan Kumar, Sumit Singh, Kanwaljeet Singh, Taslima Sheikh, P. Vishal Ahuja & Arvind Kumar, Pp. 28263–28274

**Assessing the tree diversity along the Dudhganga River in Kolhapur District of Maharashtra, India**

– Sachin Chavan & Rajaram Gurav, Pp. 28275–28286

**Flower bud growth, mortality rate, and population structure of *Sapria himalayana* Griffith f. *albavinosa* Banziger & Hansen (Rafflesiaceae) in a subtropical forest, northeastern India**

– K. Shamran Maring & Athokpam Pinokiyo, Pp. 28287–28295

**Comparing three sampling techniques for surveying and monitoring arthropods in Moroccan agroecosystems**

– Hanae El Harche, Pp. 28296–28306

**Community structure of Lepidoptera in Nantu-Bolihuto Wildlife Reserve, Sulawesi, Indonesia**

– Chairunnisah J. Lamangantjo, Marini Susanti Hamidun, Sasmianti & Dewi Wahyuni K. Baderan, Pp. 28307–28316

**Foraging niche segregation among woodpeckers in the oak-pine forest of Kumaon Himalaya, Uttarakhand, India**

– Rafat Jahan, Satish Kumar & Kaleem Ahmed, Pp. 28317–28328

**Local knowledge, attitudes, and perceptions of ecosystem services and disservices provided by the Painted Stork *Mycteria leucocephala* Pennant, 1769 (Aves: Ciconiidae) in northern India: insights for conservation**

– Yashmita-Ulman & Manoj Singh, Pp. 28329–28342

## Communications

**Analysis revealed minuscule DNA sequence data availability for Indian marine macroalgal diversity**

– Digvijay Singh Yadav, Aswin Alichen & Vaibhav A. Mantri, Pp. 28343–28349

**Checklist of rust fungi of the Nuratau Nature Reserve, Uzbekistan**

– I.M. Mustafae, M.M. Iminova, I.Z. Ortiqov, S.A. Teshaboyeva & N.Q. Iskanov, Pp. 28350–28357

**Checklist of moths (Lepidoptera: Heterocera) from the campus of University of North Bengal, Siliguri, India**

– Abhirup Saha, Ratnadeep Sarkar, Rujas Yonle, Subhajit Das, Prapti Das & Dhiraj Saha, Pp. 28358–28369

**Vulture diversity and long-term trends in the Ranikhet region, Kumaon Himalaya, Uttarakhand, India**

– Mirza Altaf Baig, Nazneen Zehra & Jamal Ahmad Khan, Pp. 28370–28377

**Nesting dynamics of Red-wattled Lapwing *Vanellus indicus* Boddaert, 1783 in urban and rural regions of Indore, India**

– Kratika Patidar & Vipul Keerti Sharma, Pp. 28378–28386

**Assessing avian diversity and conservation status in Dhamapur Lake World Heritage Irrigation Structure, Sindhudurg, Maharashtra, India**

– Yogesh Koli, Pravin Sawant & Mayuri Chavan, Pp. 28387–28398

**Population status and habitat use of Indian Grey Wolf *Canis lupus pallipes* in Pench Tiger Reserve, Madhya Pradesh, India**

– Iqra Rabbani & Sharad Kumar, Pp. 28399–28405

**Activity budgets of a zoo-housed Mishmi Takin *Budorcas taxicolor taxicolor* (Mammalia: Artiodactyla: Bovidae) herd**

– Nabanita Ghosh, Pranita Gupta, Joy Dey & Basavaraj S. Holeyachi, Pp. 28406–28412

**Extended distribution of *Nymphoides peltata* (S.G.Gmel.) Kuntze (Menyanthaceae) in Manipur, India**

– Aahen Chanu Waikhom & Bimolkumar Singh Sadokpam, Pp. 28413–28418

## Short Communications

***Impatiens chamchumroonii* (Balsaminaceae), a new record for the flora of Vietnam**

– Cuong Huu Nguyen, Diep Quang Dinh, Dinh Duc Nguyen & Keoudone Souvannakhoumane, Pp. 28419–28423

**Occurrence of the wood fern *Arachniodes sledgei* Fraser-Jenk. (Pteridophyta: Dryopteridaceae) in the northern Western Ghats, India**

– Sachin Patil & Jagannath Patil, Pp. 28424–28427

## Notes

**A note on the Petal-less Caper *Maerua apetala* (B. Heyne ex Roth) Jacobs (Capparaceae)**

– Shamsudheen Abdul Kader & Bagavathy Parthipan, Pp. 28428–28429

**Record of *Euploea mulciber* (Cramer, [1777]) (Lepidoptera: Nymphalidae) in Delhi, India: evidence of range extension in a restored urban ecosystem**

– Aisha Sultana, Mohammad Shah Hussain & Balwinder Kaur, Pp. 28430–28432

**Hump-nosed Pit Viper *Hypnale hypnale* feeding on an Allapalli Skink *Eutropis allapallensis* in Karwar, India**

– Nonita Rana, Karthy Shivapushanam, S.J.D. Frank & Govindan Veeraswami Gopi, Pp. 28433–28435

**Sighting of vagrant Red-backed Shrike *Lanius collurio* in the coastal areas of Thoothukudi, Tamil Nadu, India**

– Kishore Muthu, Anand Shibu & Santhanakrishnan Babu, Pp. 28436–28437

**First record of the Diamond Dove *Geopelia cuneata*, an Australian endemic, in Sikhna Jwhlwao National Park, Assam, India**

– Bibhash Sarkar, Bijay Basfore, Leons Mathew Abraham & Anjana Singha Naorem, Pp. 28438–28440

**First photographic record of the Rusty-spotted Cat *Prionailurus rubiginosus* (I. Geoffroy Saint-Hilaire, 1831) (Mammalia: Carnivora: Felidae) in Kuldiha Wildlife Sanctuary, Odisha, India**

– Tarun Singh, Harshvardhan Singh Rathore, N. Abhin, Subhalaxmi Muduli, Yash Deshpande, Vivek Sarkar, Diganta Sovan Chand, Samrat Gowda, Prakash C. Gogineni, Manoj V. Nair, Bivash Pandav & Samrat Mondol, Pp. 28441–28443

**First photographic evidence of the Rusty-spotted Cat *Prionailurus rubiginosus* (I. Geoffroy Saint-Hilaire, 1831) (Mammalia: Carnivora: Felidae) in Kapilash Wildlife Sanctuary, Odisha, India**

– Alok Kumar Naik, Sumit Kumar Kar, Shyama Bharati, Ashit Chakraborty & Ashis Kumar Das, Pp. 28444–28446

**Record of a Tiger *Panthera tigris* (Linnaeus, 1758) (Mammalia: Carnivora: Felidae) in Saptari District of eastern Nepal: implications for conservation and habitat connectivity**

– Gobinda Prasad Pokharel, Chiranjibi Prasad Pokharel, Ashish Gurung, Bishnu Singh Thakuri, Ambika Prasad Khatiwada, Aastha Joshi, Birendra Gautam, Mithilesh Mahato, Naresh Subedi & Madhu Chetri, Pp. 28447–28450

## Book Review

**At the Point of No Return? – Reading Pankaj Sekhsaria's Island on Edge: The Great Nicobar Crisis**

– Himangshu Kalita, Pp. 28451–28454

Publisher &amp; Host



Threatened Taxa