



Publisher Wildlife Information Liaison Development Society www.wild.zooreach.org

Host **Zoo Outreach Organization** www.zooreach.org

43/2 Varadarajulu Nagar, 5th Street West, Ganapathy, Coimbatore, Tamil Nadu 641035, India Ph: +91 9385339863 | www.threatenedtaxa.org Email: sanjay@threatenedtaxa.org

EDITORS

Founder & Chief Editor

Dr. Sanjay Molur

Wildlife Information Liaison Development (WILD) Society & Zoo Outreach Organization (ZOO), 12 Thiruvannamalai Nagar, Saravanampatti, Coimbatore, Tamil Nadu 641035, India

Deputy Chief Editor Dr. Neelesh Dahanukar

Noida, Uttar Pradesh, India

Managing Editor

Mr. B. Ravichandran, WILD/ZOO, Coimbatore, India

Associate Editors

Dr. Mandar Paingankar, Government Science College Gadchiroli, Maharashtra 442605, India

Dr. Ulrike Streicher, Wildlife Veterinarian, Eugene, Oregon, USA
Ms. Privanka Iver. ZOO/WILD. Coimbatore. Tamil Nadu 641035. India

Dr. B.A. Daniel, ZOO/WILD, Coimbatore, Tamil Nadu 641035, India

Editorial Board

Dr. Russel Mittermeier

Executive Vice Chair, Conservation International, Arlington, Virginia 22202, USA

Prof. Mewa Singh Ph.D., FASc, FNA, FNASc, FNAPsy

Ramanna Fellow and Life-Long Distinguished Professor, Biopsychology Laboratory, and Institute of Excellence, University of Mysore, Mysuru, Karnataka 570006, India; Honorary Professor, Jawaharlal Nehru Centre for Advanced Scientific Research, Bangalore; and Adjunct Professor, National Institute of Advanced Studies, Bangalore

Stephen D. Nash

Scientific Illustrator, Conservation International, Dept. of Anatomical Sciences, Health Sciences Center. T-8. Room 045. Stony Brook University. Stony Brook. NY 11794-8081. USA

Dr. Fred Pluthero

Toronto, Canada

Dr. Priya Davidar

Sigur Nature Trust, Chadapatti, Mavinhalla PO, Nilgiris, Tamil Nadu 643223, India

Dr. Martin Fishe

Senior Associate Professor, Battcock Centre for Experimental Astrophysics, Cavendish Laboratory, JJ Thomson Avenue, Cambridge CB3 OHE, UK

Dr. John Fellowe

Honorary Assistant Professor, The Kadoorie Institute, 8/F, T.T. Tsui Building, The University of Hong Kong, Pokfulam Road, Hong Kong

Prof. Dr. Mirco Solé

Universidade Estadual de Santa Cruz, Departamento de Ciências Biológicas, Vice-coordenador do Programa de Pós-Graduação em Zoologia, Rodovia Ilhéus/Itabuna, Km 16 (45662-000) Salobrinho, Ilhéus - Bahia - Brasil

Dr. Rajeev Raghavan

Professor of Taxonomy, Kerala University of Fisheries & Ocean Studies, Kochi, Kerala, India

English Editors

Mrs. Mira Bhojwani, Pune, India Dr. Fred Pluthero, Toronto, Canada Mr. P. Ilangovan, Chennai, India

Web Development

Mrs. Latha G. Ravikumar, ZOO/WILD, Coimbatore, India

Typesetting

Mrs. Radhika, ZOO, Coimbatore, India Mrs. Geetha, ZOO, Coimbatore India **Fundraising/Communications**

Mrs. Payal B. Molur, Coimbatore, India

Subject Editors 2019–2021

Fungi

Dr. B. Shivaraju, Bengaluru, Karnataka, India

Dr. R.K. Verma, Tropical Forest Research Institute, Jabalpur, India

Dr. Vatsavaya S. Raju, Kakatiay University, Warangal, Andhra Pradesh, India

Dr. M. Krishnappa, Jnana Sahyadri, Kuvempu University, Shimoga, Karnataka, India

Dr. K.R. Sridhar, Mangalore University, Mangalagangotri, Mangalore, Karnataka, India

Dr. Gunjan Biswas, Vidyasagar University, Midnapore, West Bengal, India

Plants

Dr. G.P. Sinha, Botanical Survey of India, Allahabad, India

Dr. N.P. Balakrishnan, Ret. Joint Director, BSI, Coimbatore, India

Dr. Shonil Bhagwat, Open University and University of Oxford, UK

Prof. D.J. Bhat, Retd. Professor, Goa University, Goa, India

Dr. Ferdinando Boero, Università del Salento, Lecce, Italy

Dr. Dale R. Calder, Royal Ontaro Museum, Toronto, Ontario, Canada

Dr. Cleofas Cervancia, Univ. of Philippines Los Baños College Laguna, Philippines

Dr. F.B. Vincent Florens, University of Mauritius, Mauritius

Dr. Merlin Franco, Curtin University, Malaysia

Dr. V. Irudayaraj, St. Xavier's College, Palayamkottai, Tamil Nadu, India

Dr. B.S. Kholia, Botanical Survey of India, Gangtok, Sikkim, India

Dr. Pankaj Kumar, Kadoorie Farm and Botanic Garden Corporation, Hong Kong S.A.R., China

Dr. V. Sampath Kumar, Botanical Survey of India, Howrah, West Bengal, India

Dr. A.J. Solomon Raju, Andhra University, Visakhapatnam, India

Dr. Vijayasankar Raman, University of Mississippi, USA

Dr. B. Ravi Prasad Rao, Sri Krishnadevaraya University, Anantpur, India

Dr. K. Ravikumar, FRLHT, Bengaluru, Karnataka, India

Dr. Aparna Watve, Pune, Maharashtra, India

Dr. Qiang Liu, Xishuangbanna Tropical Botanical Garden, Yunnan, China

Dr. Noor Azhar Mohamed Shazili, Universiti Malaysia Terengganu, Kuala Terengganu, Malaysia

Dr. M.K. Vasudeva Rao, Shiv Ranjani Housing Society, Pune, Maharashtra, India

Prof. A.J. Solomon Raju, Andhra University, Visakhapatnam, India

Dr. Mandar Datar, Agharkar Research Institute, Pune, Maharashtra, India

Dr. M.K. Janarthanam, Goa University, Goa, India

Dr. K. Karthigeyan, Botanical Survey of India, India

Dr. Errol Vela, University of Montpellier, Montpellier, France

Dr. P. Lakshminarasimhan, Botanical Survey of India, Howrah, India

Dr. Larry R. Noblick, Montgomery Botanical Center, Miami, USA Dr. K. Haridasan, Pallavur, Palakkad District, Kerala, India

Dr. Analinda Manila-Fajard, University of the Philippines Los Banos, Laguna, Philippines

Dr. P.A. Sinu, Central University of Kerala, Kasaragod, Kerala, India Dr. Afroz Alam, Banasthali Vidyapith (accredited A grade by NAAC), Rajasthan, India

Dr. K.P. Rajesh, Zamorin's Guruvayurappan College, GA College PO, Kozhikode, Kerala, India

Dr. David E. Boufford, Harvard University Herbaria, Cambridge, MA 02138-2020, USA

Dr. Ritesh Kumar Choudhary, Agharkar Research Institute, Pune, Maharashtra, India

Dr. Navendu Page, Wildlife Institute of India, Chandrabani, Dehradun, Uttarakhand, India Dr. Kannan C.S. Warrier, Institute of Forest Genetics and Tree Breeding, Tamil Nadu, India

vertebrates

Dr. R.K. Avasthi, Rohtak University, Haryana, India

Dr. D.B. Bastawade, Maharashtra, India

Dr. Partha Pratim Bhattacharjee, Tripura University, Suryamaninagar, India

Dr. Kailash Chandra, Zoological Survey of India, Jabalpur, Madhya Pradesh, India

Dr. Ansie Dippenaar-Schoeman, University of Pretoria, Queenswood, South Africa Dr. Rory Dow, National Museum of natural History Naturalis, The Netherlands

Dr. Brian Fisher, California Academy of Sciences, USA

Dr. Richard Gallon, llandudno, North Wales, LL30 1UP

Dr. Hemant V. Ghate, Modern College, Pune, India

Dr. M. Monwar Hossain, Jahangirnagar University, Dhaka, Bangladesh

Mr. Jatishwor Singh Irungbam, Biology Centre CAS, Branišovská, Czech Republic.

Dr. Ian J. Kitching, Natural History Museum, Cromwell Road, UK

For Focus, Scope, Aims, and Policies, visit https://threatenedtaxa.org/index.php/JoTT/aims_scope
For Article Submission Guidelines, visit https://threatenedtaxa.org/index.php/JoTT/about/submissions
For Policies against Scientific Misconduct, visit https://threatenedtaxa.org/index.php/JoTT/policies_various

continued on the back inside cover

Cover: Common Silverline Spindasis vulcanus vulcanus in poster colours adapted from photograph by Kalpesh Tayade. © Pooja R. Patil.

Journal of Threatened Taxa | www.threatenedtaxa.org | 26 December 2022 | 14(12): 22277-22292

ISSN 0974-7907 (Online) | ISSN 0974-7893 (Print)

https://doi.org/10.11609/jott.8040.14.12.22277-22292

#8040 | Received 07 June 2022 | Final received 01 September 2022 | Finally accepted 02 December 2022





BELLEVILLE STATE OF THE STATE O

Systematics of the enigmatic and narrowly endemic toad genus *Bufoides*Pillai & Yazdani, 1973: rediscovery of *Bufoides kempi* (Boulenger, 1919) and expanded description of *Bufoides meghalayanus* (Yazdani & Chanda, 1971) (Amphibia: Anura: Bufonidae) with notes on natural history and distribution

R.S. Naveen ¹, S.R. Chandramouli ², Gautam Kadam ³, S. Babu ⁴, P.V. Karunakaran ⁵, H.N. Kumara ⁶, & N. Parthasarathy ⁷

1.3.4.5.6 Sálim Ali Centre for Ornithology and Natural History, Anaikatty, Coimbatore, Tamil Nadu 641108, India.
1.2.7 Department of Ecology and Environmental Sciences, School of Life Sciences, Pondicherry University, Puducherry 605014, India.
¹ naveen89240@gmail.com, ² findthesnakeman@gmail.com (corresponding author), ³ gautamkadam7wild@gmail.com,
⁴ sanbabs@gmail.com (corresponding author), ⁵ karunakaran.pv@gmail.com, ⁶ honnavallik@gmail.com, ⁿ nparthasarathypu@gmail.com

Abstract: *Bufoides kempi* (Boulenger, 1919) known only from the two historical syntype specimens until now was rediscovered after more than a century from near its type locality in the Garo Hills, Meghalaya, northeastern India. Analysis of mitochondrial 16S rRNA gene reveals congenericity between *B. kempi* and *B. meghalayanus* with an inter-specific genetic divergence of 4.67%. Description of *B. kempi* is expanded based on the six male and two female specimens collected during this study. We provide the first description of calls for this genus, notes on their breeding biology and larval morphology. Additional specimens of *B. meghalayanus* collected during this study are described to supplement its characterization.

Keywords: Amphibians, breeding biology, calls, Garo Hills, Khasi Hills, larval morphology, new records, syntype specimens.

Editor: Raju Vyas, Vadodara, Gujarat, India.

Date of publication: 26 December 2022 (online & print)

Citation: Naveen, R.S., S.R. Chandramouli, G. Kadam, S. Babu, P.V. Karunakaran, H.N. Kumara & N. Parthasarathy (2022). Systematics of the enigmatic and narrowly endemic toad genus *Bufoides* Pillai & Yazdani, 1973: rediscovery of *Bufoides kempi* (Boulenger, 1919) and expanded description of *Bufoides meghalayanus* (Yazdani & Chanda, 1971) (Amphibia: Anura: Bufonidae) with notes on natural history and distribution. *Journal of Threatened Taxa* 14(12): 22277–22292. https://doi.org/10.11609/jott.8040.14.12.22277-22292

Copyright: © Naveen et al. 2022. Creative Commons Attribution 4.0 International License. JoTT allows unrestricted use, reproduction, and distribution of this article in any medium by providing adequate credit to the author(s) and the source of publication.

Funding: National Mission on Himalayan Studies (GBPNI/NMHS-2017-18/MG 32, dated:28/3/2018).

Competing interests: The authors declare no competing interests.

Author details: R.S. NAVEEN works on herpetofauna & ichthyofauna and obtained his MSc in ecology from Pondicherry University. S.R. CHANDRAMOULI obtained his PhD in ecology and environmental sciences from Pondicherry University works on systematics, ecology, biogeography and conservation of herpetofauna of India, with special focus on the Andaman and Nicobar Islands. GAUTAM KADAM is interested in the study of spiders and herpetofauna and holds a MSc in biodiversity and wildlife conservation from Mumbai University. S. BABU is a senior scientist at SACON, Tamil Nadu and his research focuses on the conservation of birds, landscape ecology and remote sensing & GIS. HONNAVALLI N. KUMARA is a principal scientist in SACON. His research focuses on various aspects of conservation. DR. PV. KARUNAKARAN is a senior principal scientist at SACON, conducting biodiversity and natural resource based studies and strategic planning in different landscapes of India. PROF. N. PARTHASARATHY is a faculty at the department of Ecology and Environmental Sciences, engaging in teaching of ecology of terrestrial biome, forest botany, medicinal plant resources, functional ecology, and dynamics of tropical forests across the forests of Western and Eastern Ghats, Andamans and

Author contributions: SRC and SB conceived and designed the study, RSN & GK conducted field studies, SRC, RSN, & SB generated and analysed the data, SRC, RSN, & SB wrote the manuscript, SB, PVK, & HNK procured funds, SB, PVK, HNK, & NP supervised the study, reviewed, & edited the manuscript draft, and all authors approved the final draft.

Acknowledgements: This publication is an outcome of the project entitled 'Characterization of Community Reserves and Assessment of their Conservation Values in Meghalaya' funded by the National Mission on Himalayan Studies (GBPNI/NMHS-2017-18/MG 32, dated: 28.03.2018). Our sincere thanks are due to the additional principal chief conservator of forests (Wildlife) and chief wildlife warden (CWLW), Meghalaya, and officers of the Department of Forest and Environment, Government of Meghalaya for facilitating permission from the Community Reserve Management Committees to carry out the field study (No. FWC/ Research/15/603-04 dated 31st May, 2019). We are indebted to the management committees of each community reserves who have given us permission for this work. We thank the technology platform services, C-CAMP, National Centre for Biological Sciences for their expertise in sequencing a sample used in this study. We thank Soni R Marak for his assistance with fieldwork. We thank director, SACON for smooth execution of the project.









INTRODUCTION

The cosmopolitan anuran family Bufonidae Gray, 1825 is represented in India by nine genera comprising 33 species spread across several biogeographic regions (Frost 2022). One of the most poorly-known among them is the genus Bufoides Pillai & Yazdani (1973) which comprises two species namely B. meghalayanus (Yazdani & Chanda 1971) and B. kempi (Boulenger 1919) (after Chandramouli & Amarasinghe 2016). Among them, the type species, B. meghalayanus is fairly better-known in terms of its distribution, biology, ecology, and natural history (Yazdani & Chanda 1971; Pillai & Yazdani 1973; Das et al. 2009; Deuti et al. 2012). On the other hand, there has been no information on any of the above aspects for the previously-described species B. kempi. B. kempi was originally described as Nectophryne kempi from 'above Tura, 2,500 ft' based on two specimens, an adult and a subadult (ZSI 18481a,b), from which the species is known till date (Boulenger 1919). It was later transferred to the genus *Pedostibes* Günther, 1875 by Barbour (1938) and subsequently transferred to Bufoides by Chandramouli & Amarasinghe (2016) based on morphological characters. As a part of an on-going project documenting faunal diversity in community reserves of Meghalaya, we rediscovered B. kempi from near its type locality and located additional specimens of B. meghalayanus whose descriptions are expanded based on new data.

MATERIALS AND METHODS

The study was conducted in the northeastern Indian state of Meghalaya. Surveys were conducted in ten different locations spread across the Garo, Khasi, and Jaintia Hills of Meghalaya. Specifically, the type localities of the two known species viz. "above Tura, Garo Hills" (Boulenger 1919) for B. kempi and "Mawblang plateau" (Yazdani & Chanda 1971) for B. meghalayanus and the vicinity of these localities were surveyed intensively, in addition to the other sites to locate the target species. Additional sites within Meghalaya (Deuti et al. 2012), apart from these two localities from where Bufoides is known until now were also surveyed. Field sampling was carried out from March-May & October 2021. A total of seven specimens of B. kempi, comprising six adult males and one adult female along with a subadult female were collected from Eman Asakgre (25.37° N, 90.55° E, 100-250 m). Likewise, three adult specimens of Bufoides meghalayanaus were collected from a hill stream in the Khasi Hills (25.23° N, 91.73° E, 1,100–1,250 m). Other locations of *B. kempi* and *B. meghalayanus* were marked with a GPS. Encounter rates of these species, expressed as the number of individuals encountered over an hour's duration of field sampling, is presented as an index of their abundance.

The following measurements were recorded to the nearest 0.02 mm from the specimens with a dial caliper: snout-vent length (SVL, from the tip of the snout to the anterior margin of the cloaca), axilla-groin distance (AG, from the posterior margin of the forelimb at its insertion point on the body to the anterior margin of the hind limb at its insertion point on the body), head length (HL, from the posterior edge of the mandible to the tip of the snout), head width (HW, the maximum width of the head at the angle of the jaws), head depth (HD, the maximum depth of the head), body width (BW, the maximum width of the body at the trunk), eye diameter (ED, the greatest horizontal diameter of the orbit), eyenostril distance (EN, from the anterior border of the orbit to the middle of the nostril), eye-snout distance (ES, from the anterior border of the orbit to the tip of the snout), upper eyelid width (UEW, the maximum width of the upper eyelid), interorbital distance (IO, distance between the upper eyelids), internarial distance (IN, distance between the nostrils), upper arm length (UAL, from the axilla to elbow), lower arm length (LAL, from the posterior margin of the elbow to the base of the outer metacarpal tubercle), palm length (PAL, from the posterior border of the outer metacarpal tubercle to tip of the 3rd finger), femur length (FEL, from the cloaca to the knee), tibia length (TBL, from knee to heel), foot length (FOL, from inner metatarsal tubercle to the tip of the 4th toe). Webbing formulae follows Savage & Heyer (1997). A principal component analysis was conducted based on 18 morphometric measurements (standardised to their SVL; Table 1) of the two Bufoides species to examine their morphometric distinction from each other. Calls were recorded with the camera as videos and the audio (as .mp4 at an audio sampling rate of 48 kHz) was extracted and analysed with Adobe Audition 6 and Adobe Soundbooth CS3. Two specimens, SACON VA 157 (female) and VA 159 (male) were radiographed to study osteological characters of B. kempi. A brief description of its osteology is provided following the terminologies of Noble (1931).

Eggs of *B. kempi* observed in tree holes were collected and reared for 11 days, and the growth of the larvae was monitored with preservation of samples across various developmental stages. The following measurements of the tadpoles: HBL— head-body length; HBW— head-

body width; HBD— head-body depth; TOT— total length; TAIL— tail length; IO— inter-orbital distance; and TH— tail-fin height were recorded with a stereo microscope following Chandramouli & Kalaimani (2014). Staging of tadpoles follow Gosner (1960) and terminologies follow McDiarmid & Altig (1999). Labial tooth-row formula for the larvae follow Rödel (2000).

Total genomic DNA was extracted from one specimen of B. kempi (SACON VA 180) with a DNA extraction and purification kit, following the manufacturer's protocols. 16S rRNA gene was amplified using the primers 16sAR-L (5'-CGCCTGTTTATCAAAAACAT-3') and 16sB R-H (5'-CCGGTCTGAACTCAGATCACGT 3'), respectively (Kocher et al. 1989). Amplifications were performed in a Applied Bio Systems Veriti 96 well thermal cycler: 20 μl reactions with 4 μ l of 5X Phusion HF buffer, 0.4 μ l of 10 mM dNTP, 0.2 μl of Phusion DNA Polymerase, 0.1 μl each of forward and reverse primers, 2.0 µl of DNA template and 13.2 µl of nuclease free water with the following procedure: initial denaturation of DNA at 95 °C for 5 min, 35 cycles of: denaturation at 95 °C for 1 m, annealing at 55 °C for 1 min, extension at 72 °C for 1 m and at last, final extension at 72 °C for 10 min. The amplicon was checked by running it through an agarose gel electrophoresis for a clear band of the desired region in the amplified PCR product. The amplified PCR product was purified and sequenced commercially (National Centre for Biological Sciences, Bengaluru). The sequence thus obtained (NCBI voucher no: OP920605) was aligned along with ten other taxa from Bufonidae, comprising the genera Adenomus, Beduka, Blythophryne, Bufoides, Bufotes, Duttaphrynus, and *Pedostibes* with *Hyla arborea* as the outgroup taxon. The sequences were aligned with MUSCLE (Edgar 2004) in MEGA 6.0 (Tamura et al. 2013). This alignment of 491 bp was exported in FASTA and MEGA formats, and was then used to determine uncorrected pairwise genetic distances between the samples with MEGA 6. The FASTA alignment was converted to PHYLIP format in the Alignment Transformation Environment (ALTER) website (www.sing.ei.uvigo.es/ ALTER) and was subjected to a maximum likelihood (ML) analysis in RAxML GUI v. 1.3 (Stamatakis 2006) using the general time reversible model, GTR GAMMA, (as RAxML uses only the general time reversible (GTR) model of sequence evolution) with 500 bootstrap replicates. Likewise, for the Bayesian analysis, the FASTA alignment was converted to NEXUS format and analysed in MrBayes 3.1.2 (Ronguist & Huelsenbeck 2003) by running it for three million MCMC iterations initially until the standard deviation of the split frequencies reached a value of ≤ 0.001. Else, the analysis was continued for another 10000-100000 generations until the standard deviation of \leq 0.001 was obtained for the split frequencies. Initial 20% of the trees were discarded as 'burn-in'. The tree files generated were then visualized using Fig Tree v. 1.4.0.

RESULTS

Our analyses of molecular data (both maximum likelihood - ML and Bayesian - BI) recovered the two species allocated to the genus Bufoides to form a monophyletic group; with the two species B. meghalayanus and B. kempi showing a congeneric, sister relationship to each other with high support (87 & 1.0 in ML and BI, respectively). The ML and BI analyses recovered the genera Blythophryne Chandramouli et al., 2016 & Beduka Dubois et al., 2021 to be close to the genus Bufoides, as assessed earlier (Chandramouli et al. 2016) although with low support (36 & 0.63 in ML & BI, respectively). Pairwise genetic divergence between B. meghalayanus and B. kempi was found to be moderate (4.67 % at 16s rRNA) supporting their specific distinction from each other (Figures 1a,b). The PCA conducted based on 18 morphometric variables clearly separates the two species into two discrete clusters (Figure 2, Table 1).

SYSTEMATICS

Bufoides kempi (Boulenger, 1919)

Nectophrryne kempi Boulenger, 1919

Pedostibes kempi – Barbour, 1938

Bufoides kempi – Chandramouli & Amarasinghe, 2016

Syntypes: Two specimens; an adult (29.8 mm SVL) and a subadult (17.4 mm) (ZSI 18481 a&b, respectively)

Other material studied: SACON VA 157 (an adult female) and, VA 181(a subadult female), and SACON VA156; VA 158 –160; VA 164 & VA 180 six adult males collected from Eman Asakgre (25.37°N, 90.54°E, 200 m asl.), Garo Hills, Meghalaya (Image 1).

Diagnosis: (after Chandramouli & Amarasinghe 2016)

A semi-arboreal to rupicolous *Bufoides* from the Garo Hills diagnosed by: small to medium body size (SVL 24.1–32.36 mm); presence of irregular, non-keratinized cranial ridges (pre and post orbital); short, ovoid parotoid glands; absence of an externally visible tympanum; moderate degree of webbing between toes (two phalanges of toe IV free); partial webbing between fingers, and the presence of small, slightly dilated, rounded terminal digital discs at the tips of both fingers and toes. Dorsum black with mossy green shade along the flanks in males, females predominantly green with black reticulations; a



Table 1. Eigenvalues and the proportion of variance explained by each of the principal component.

	PC 1	PC 2	PC 3	PC 4	PC 5	PC 6	PC 7	PC 8
AG	0.13	-0.05	0.66	-0.25	-0.26	-0.16	0.19	0.13
BW	0.73	-0.38	-0.01	0.25	-0.25	0.11	-0.02	0.09
HL	0.24	0.32	-0.13	-0.46	-0.29	-0.10	0.05	-0.07
HW	0.51	0.20	-0.02	-0.02	0.31	0.01	0.04	0.10
HD	0.11	0.09	-0.19	0.08	0.02	-0.24	-0.15	-0.25
ED	0.16	0.08	0.06	0.10	-0.02	0.01	-0.36	0.23
EN	0.08	0.26	0.09	0.13	-0.16	0.22	-0.38	-0.49
ES	0.04	0.20	-0.22	0.16	-0.20	0.32	0.39	-0.18
UEW	0.04	0.08	-0.12	-0.09	0.16	0.13	-0.22	0.14
10	-0.06	0.22	-0.12	0.16	-0.05	0.19	-0.03	0.33
IN	-0.07	0.27	0.03	0.08	-0.17	0.34	-0.15	0.54
UAL	-0.06	0.18	0.53	0.54	0.05	-0.08	-0.14	-0.14
LAL	-0.01	0.14	0.01	0.22	-0.13	-0.29	0.48	0.12
PAL	0.11	0.06	-0.17	0.38	0.35	-0.13	0.26	0.01
FEL	0.14	0.54	0.01	-0.03	-0.15	-0.10	0.09	-0.17
TBL	-0.03	0.29	-0.01	0.04	0.09	-0.40	-0.10	0.31
TAR	0.06	0.14	0.33	-0.18	0.45	0.51	0.28	-0.07
FOL	0.19	0.07	0.05	-0.22	0.44	-0.21	-0.14	-0.04
Eigenvalue	0.01	0.01	0.00	0.00	0.00	0.00	0.00	0.00
% variance	46.40	24.50	11.91	6.90	5.37	2.64	1.47	0.81

pale white venter; eggs partially pigmented and laid in strings within water-filled tree holes (phytotelmata).

Description and variation: (based on the newly collected material) Table 2

Female (SVL 32.36 mm) slightly larger than males (mean SVL 26.38 mm ± 0.88, n = 6). Head flat, fairly large, and distinct, (HL:SVL 0.31), broader than long, slightly more wider in the female (HL:HW 0.82) than in males (HL:HW 0.91), with an obtusely pointed to rounded snout tip. Trunk short (AG:SVL 0.4) and slightly gracile in males (AG:BW1.45) than in females (AG:BW 1.81). Eyes fairly large (ED:HL 0.33) their diameter shorter than the snout length (ED:ES 0.71). Nostrils situated closer to the snout tip than to the eyes (EN:ES 0.74). Upper eyelids wide, (mean UEW 2.92± 0.12) rugose with keratinized pustules, narrower than the interorbital space (IO:UEW 1.79). Inter-orbital space broader than inter-narial space (IO:IN 1.59). Upper arms short (UAL:SVL 0.23), nearly as long as the lower arms (UAL:LAL 1.02); palm slightly shorter than the upper arms (UAL:PAL 0.88). Fingers partially webbed, webbing formula I_{0.1}II_{2.3}III_{3.2}IV; relative length of fingers III>IV>II>I. Outer metacarpal tubercle large and evident. Finger tips with slightly expanded rounded discs. Femur relatively short (FEL:SVL 0.38), tibia slightly longer than femur (FEL:TBL 0.91); foot about as long as the femur (FEL:FOL 0.99). Toes moderately webbed, webbing formula: $I_{0.0}II_{0.0.5}III_{0.5.2}IV_{2.1}V$, a relatively large inner and a slightly smaller ovoid metatarsal tubercle at the base of the foot. Toe tips with discs as broad as the toes; tarsal ridge not discernible. Vocal sac not discernibly distinct in males. Skin rugose in texture with keratinized granules.

Colouration in life

Males were generally dark grey in colour with traces of mossy green along the flanks and yellow patches near the axilla, belly and groin on the ventro-lateral region. Females are predominantly mossy green with an irregular black hour-glass pattern on the dorsum. Limbs visibly barred with black. Venter pale and much lighter than the dorsum (Image 2).

Osteology

Skull large and triangular, with an obtusely pointed snout tip. Pre and post-orbital ridges discernible. Frontoparietals fairly broad and hexagonal in shape. Nasal bones of the skull short, nearly as long as broad. Vertebral column with eight procoelous presacral vertebrae; the first four relatively larger than the following. Sacral diapophyses broad, flattened, and expanded laterally. Urostyle cylindrical, about half the length of the presacral



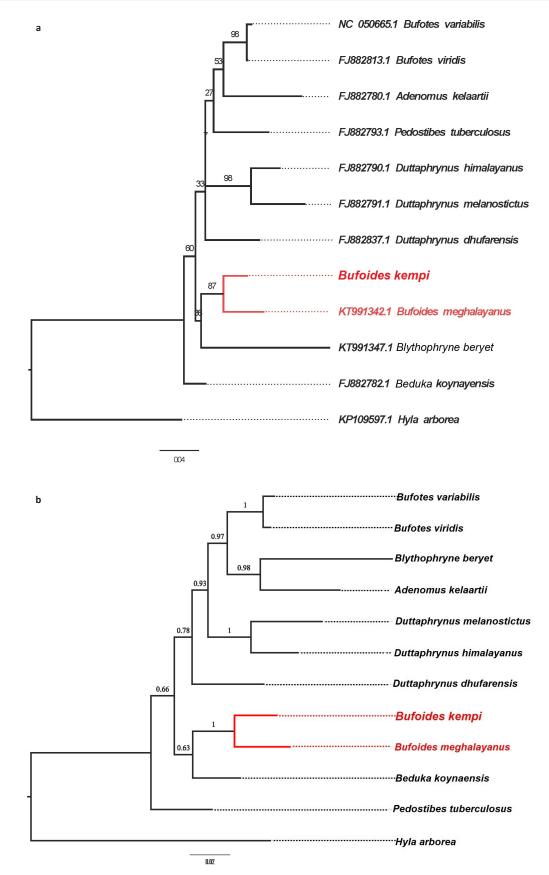


Figure 1. a— Maximum likelihood | b— Bayesian phylogenetic trees of Bufonids, showing the distinction of *Bufoides* from other genera and sister relationship between *B. meghalayanus* and *B. kempi*.



Table 2. Morphometric measurements of *Bufoides kempi* and *B. meghalayanus* at SACON.

Species					Bufoides kemp	i			
Voucher no:	VA 164	VA 159	VA 158	VA 160	VA 180	VA 156	MEAN	±SD	VA 157
Sex	М	М	М	М	М	М			F
SVL	25.3	29.3	26.8	24.1	32.7	26.3	27.4	3.1	32.4
AG	8.5	11.3	10.9	7.8	13.9	10.6	10.5	2.2	13.8
BW	6.2	9.5	7.3	5.3	12.5	5.5	7.7	2.8	7.6
HL	7.7	9.5	8.06	8.7	10.4	8.4	8.8	1.0	9.9
HW	8.8	10.6	8.8	9.2	13.9	9.2	10.1	2.0	12.1
HD	4.1	4.0	4	4.2	5.1	3.6	4.2	0.5	5.5
ED	2.8	3.1	2.9	2.3	3.8	2.5	2.9	0.5	3.3
EN	2.6	2.9	3.4	3.1	3.3	2.2	2.9	0.4	3.5
ES	4.1	4.7	3.7	4.6	4.9	2.9	4.1	0.8	4.5
UEW	2.6	2.9	2.2	2.7	2.8	2.7	2.7	0.3	3.2
10	4.9	4.9	4.3	4.7	4.9	4.0	4.6	0.4	5.9
IN	2.9	2.9	2.4	2.6	2.2	2.2	2.5	0.3	4.1
UAL	6.5	6.2	7.7	5.4	8.9	6.2	6.8	1.3	7.2
LAL	6.3	6.2	6.1	5.8	8.1	5.6	6.4	0.9	7.4
PAL	7.2	6.9	6.3	6.7	9.4	6.2	7.1	1.2	8.9
FEL	9.6	10.1	9.7	10.2	12.2	9.4	10.2	1.0	12.3
TBL	11.2	11.1	10.9	10.6	13.3	11.2	11.4	1.0	13.1
TAR	5.4	7.6	6.2	6.2	8.9	7.3	6.9	1.3	7.5
FOL	8.8	10.2	9.3	9	12.7	10.4	10.1	1.4	13.0
F1	1.7	2.3	1.2	1.4	2.3	1.6	1.8	0.5	2.5
F2	2.8	2.6	2.4	1.7	3.1	2.4	2.5	0.5	4.9
F3	5.4	4.4	4.2	3.6	5.9	3.5	4.5	1.0	7.1
F4	3.7	4.0	2.9	2.6	4.9	2.8	3.5	0.9	5.5
T1	1.3	1.4	1.2	0.9	1.5	1.3	1.3	0.2	2.1
T2	2.5	4.1	1.8	2.7	2.7	2.1	2.7	0.8	2.9
T3	4.9	3.8	2.6	4.1	4.0	3.3	3.8	0.8	3.9
T4	7.4	6.0	4.7	5.9	6.6	4.8	5.9	1.0	7.7
T5	4.0	2.4	2.7	4.2	3.9	2.9	3.3	0.8	4.9

Species		Buf	oides meghalaya	nus	
Voucher no:	VA 215	VA 251	VA 252	MEAN	±SD
Sex	М	М	М		
SVL	31.3	33.5	31.2	31.9	1.3
AG	10.7	13.2	12.1	12.3	1.4
BW	12.0	14.1	10.9	12.3	1.6
HL	9.7	12.9	12.7	11.8	1.8
HW	12.0	16.8	12.9	13.9	2.5
HD	5.4	6.2	5.4	5.7	0.5
ED	3.2	5.3	3.8	4.1	1.1
EN	2.1	4.9	3.6	3.5	1.4
ES	4.2	5.4	5.1	4.9	0.6
UEW	3.2	4.1	3.1	3.5	0.6



Species		Bufoides meghalayanus VA 215 VA 251 VA 252 MEAN ±SD 4.3 5.7 5.1 5.0 0.7 1.1 3.2 2.9 2.4 1.2 5.3 8.2 6.4 6.6 1.5									
Voucher no:	VA 215	VA 251	VA 252	MEAN	±SD						
Ю	4.3	5.7	5.1	5.0	0.7						
IN	1.1	3.2	2.9	2.4	1.2						
UAL	5.3	8.2	6.4	6.6	1.5						
LAL	6.5	7.4	7.6	7.1	0.6						
PAL	8.6	9.7	7.5	8.6	1.1						
FEL	9.6	14.4	13.2	12.4	2.5						
TBL	11.9	14.4	13.4	13.2	1.3						
TAR	6.4	9.3	6.9	7.6	1.5						
FOL	11.9	14.3	11.8	12.7	1.4						
F1	2.1	2.0	1.8	1.9	0.2						
F2	3.2	3.2	3.1	3.2	0.1						
F3	5.4	4.7	4.4	4.8	0.5						
F4	4.3	4.4	3.9	4.2	0.3						
T1	2.1	2.4	1.6	2.0	0.4						
T2	3.2	2.9	3.2	3.1	0.2						
Т3	5.4	4.5	3.7	4.5	0.8						
T4	9.7	6.6	5.9	7.4	2.0						
T5	6.5	5.0	3.8	5.1	1.3						

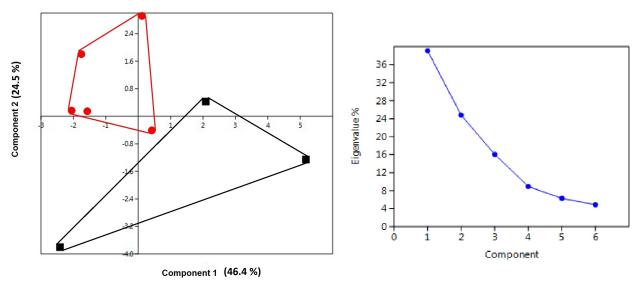


Figure 2. Left— Plot of PCA showing morphometric distinction between *B. meghalayanus* (black squares) and *B. kempi* (red dots) | Right— Scree plot showing the eigenvalues of each principal components.

vertebral column, lacking lateral expansions. Ilia curved laterally, as long as the urostyle. Ischium protruding posteriorly. Pectoral girdle arciferal. Humerus longer than radio-ulna. Phalangeal formula of the fingers: 2-2-3-3. Femur long, nearly as long as the tibiofibula; tarsus about 3/4th the length of tibiofibula. Phalangeal formula of the toes 2-2-3-4-3 (Image 3).

Breeding biology and natural history

A total of 17 individuals were seen in the following precise locations surrounding Eman Asakgre community reserve, South Garo Hills, the details of which are mentioned below (Image 4).



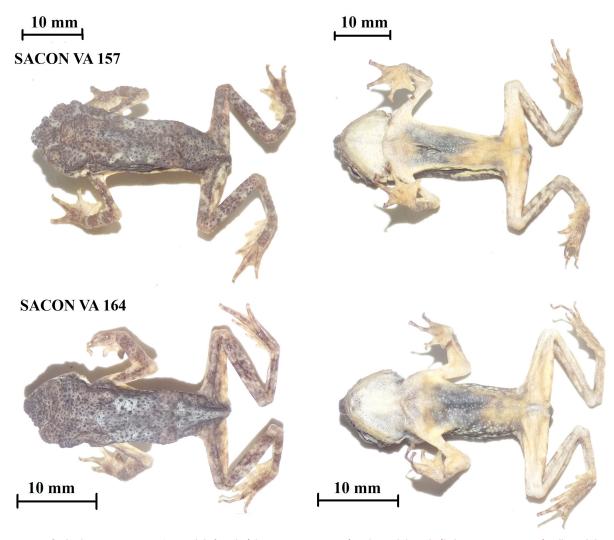


Image 1. Bufoides kempi in preservation—adult female (above: SACON VA 157) and an adult male (below: SACON VA 164) collected during this study.

Chibanda cave (25.36° N, 90.53° E, 122 m)

The cave was about 6 m below the ground level, surrounded by moist evergreen forest. A small creek was flowing into the cave and the surface of the rocks and boulders in the area was covered with moss and were wet. The canopy cover provided about 70% shade to the ground. The first individual, a subadult male was found under a boulder near the mouth of the cave. Second individual was seen inside the cave in a deep narrow Horizontal crevice of a limestone rock. Odorrana chloronota, Amolops assamensis, Limnonectes khasianus, and Ingerana borealis were some of the anuran species that were observed in sympatry with B. kempi at this location. During the night surveys in the subsequent months, seven more individuals were spotted in total. Some individuals were observed on leaves of shrubs without exhibiting any specific

behaviour between 1900–2200 h. Later in October, an adult male was sighted in a tree hole filled with rain water at a height of about 2 m above the ground and a subadult female was recorded on low lying shrubs at about one foot above the ground.

Dhangit cave (25.36° N, 90.52° E, 220 m)

This cave, surveyed in May was at a depth of about 12 m, surrounded by moist evergreen forest; the terrain was rocky and filled with boulders covered with moss. No individuals were found inside the cave, however two males were observed near the cave about 150 m away which were calling actively from a cavity filled with rain water (5 cm deep, water temperature 21° C) measuring 10 cm in diameter at about 1.2 m off the ground at around 2200 h possibly trying to attract the attention of a receptive female nearby. Two males were observed









Image 2. Bufoides kempi (male—left & middle and female—right) in life from Eman Asakge, Garo Hills. © left and middle: S.R. Chandramouli, right: R.S. Naveen

Table 3. Larval measurements of Bufoides kempi.

Stage	20	20	20	Mean	±SD	21	30	31	31	Mean
HBL	5.4	6.14	5.8	5.77	0.37	5.3	5.34	5.28	5.5	5.39
тот	10.1	9.56	9.58	9.75	0.31	10.6	11.64	12.2	14.36	13.28
TAIL	4.7	3.42	3.78	3.98	0.66	5.3	6.3	6.92	8.86	7.89
HBW	1.7	2.3	1.74	1.91	0.34	2.76	2.36	2.72	2.8	2.76
TH	1.6	2.44	1.7	1.91	0.46	2.34	1.88	2.44	2.2	2.32
10	0.3	0.3	0.3	0.00	0.00	0.4	1.46	2.64	2.6	2.62
нвн	2.82	2.6	1.9	2.44	0.48	2.0	2.24	2.24	2.42	2.33

to show aggression by kicking each other while calling and one of them was seen kicking the other with its hind feet repeatedly. Upon further search in the region, a tree cavity with about 30–40 eggs laid in strings was found. Eggs from this cavity were collected and maintained in a plastic jar with water from the same cavity for the next 11 days.

Cehise Stream (25.34° N, 90.51° E, 250 m)

The area surrounding a small stream flowing near the village of Eman Dura Banda was surveyed in May. This area had a rocky terrain and was covered with moist evergreen forest, with a tall canopy. An adult male was found resting under a boulder.

Eman Asakgre Community Reserve (25.37° N, 90.54° E, 108 m)

Congregation of four males was seen along with a female in a buttress root cavity at a height of 0.6 m above the ground, measuring about 15 cm in diameter, filled with 10 cm of rain water. The water temperature here was 23.5° C, the humidity of the location was 80%. The group was first spotted at about 1730 h in amplexus with four males and a female. One of the four males was seen mounted ventrally while the other three

males were mounted dorsally and laterally. Amplexus was axillary. The episode lasted till about 2100 h by the time the female laid egg strings with 30–40 partially pigmented eggs that measured about 2 mm diameter. Once the eggs were laid both males and the female started leaving the cavity and no further attendance was observed.

Description of calls

The call of *B. kempi* described here was composed of syllables of 'treek... treek...' that lasted for a duration of 2.1 s and was composed of three distinct notes, each of which comprise seven—ten pulses. The mean duration of each note was 6.33 ms, with a mean interval of 1.05 s in between. Maximum amplitude of the call was -1 dB with a dominant frequency of 2.5 kHz (Image 5).

Larval description: (Table 3)

On day three, 39 larvae emerged which were reared subsequently for the next eight days during which their development was documented. Stage-wise descriptions of the larvae are presented in detail below.

Stage 20: (n = 3)

The larvae reached stage 20 on day two after



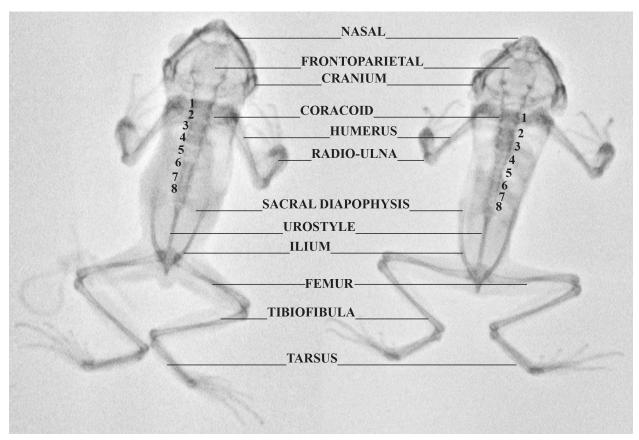


Image 3. Osteology of B. kempi based on an adult male and female vouchers SACON VA 157 and VA 159, respectively.

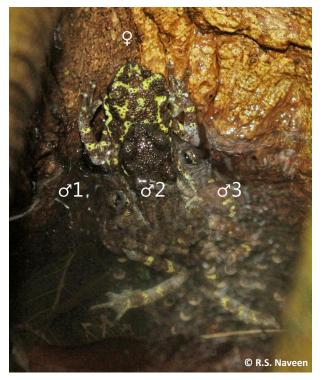


Image 4. Axillary amplexus between one female and three males of *B. kempi* observed in a phytotelmata at Eman Asakgre, Garo Hills.

emerging from the eggs. At this stage, they measured 9.75 mm \pm 0.31 in total length, with a head-body length of 5.77 mm \pm 0.37; streamlined and narrow in form (HBW 1.91 mm \pm 0.3); slightly higher than broad (HBH 2.44 mm \pm 0.48); tail a little shorter than the head-body (3.98 mm \pm 0.66); and with an average tail height of 1.91 mm \pm 0.46. Eyes and mouthparts not discernible at this stage.

Stage 22: (n = 1)

The larvae reached stage 22 on day four after emergence from the eggs. At this stage, the body & tail elongated a little more, with the larva measuring 10.6 mm in total length, with a head-body length of 5.3 mm and a relatively longer tail (5.3 mm) which equalled the HBL. Head-body oval & narrow (HBW 2.76 mm; HBH 2.0 mm). Tail-fin relatively well developed than in the earlier stage, longer and broader, with a height of 2.34 mm. Eyes dorsal in position and traces of gills discernible at this stage.

Stage 30: (n = 2)

At this stage, the larvae grew a little longer, measuring



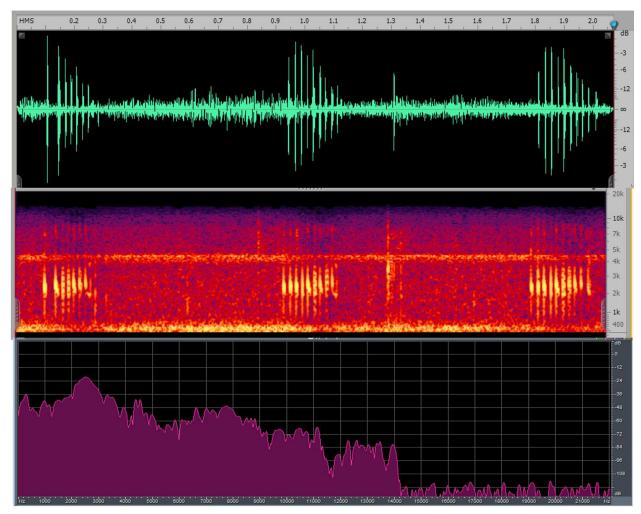


Image 5. Oscillogram, spectrogram and power spectrum of the call of B. kempi.

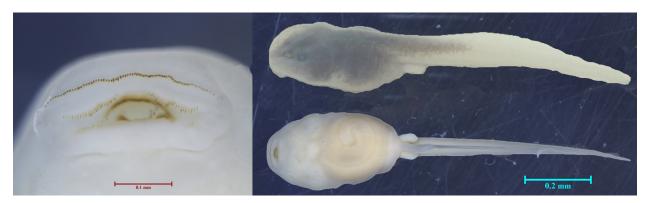


Image 6. Stage 30 larva of *B. kempi*: Mouthpart and ventral view.

13.28 mm in length, with a 7.9 mm long tail and 5.39 mm long head-body. Head-body ovoid, broader (HBW 2.76 mm) than deep (HBH 2.33 mm). Oral disc discernible at this stage with keratodont and keratinized jaw sheaths; oral tooth-row formula 1/1+1/1/1. Tail fins transparent

and high 2.32 mm. Rudimentary hindlimb buds visible at the posterior end of the head-body. Eyes well developed than in the earlier stage and in lateral position, with an inter-orbital space of 2.62 mm (Image 6).



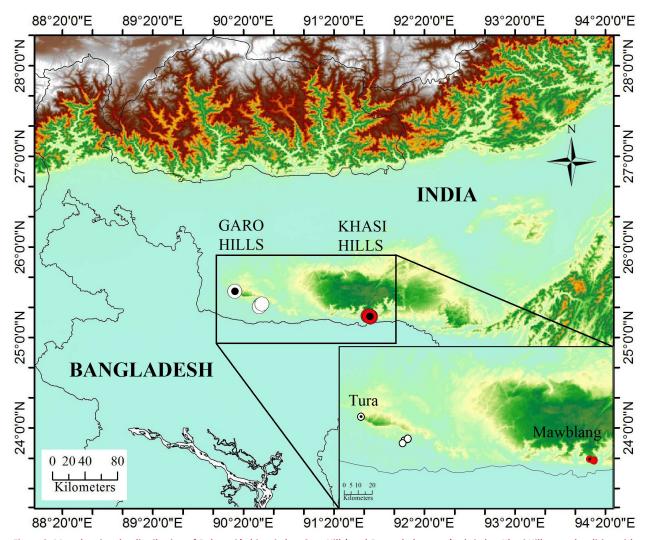


Figure 3. Map showing the distribution of *B. kempi* (white circles; Garo Hills) and *B. meghalayanus* (red circles; Khasi Hills; type localities with a dot in the middle).

Distribution

During the present study, *B. kempi* was recorded from the above four locations, of which, two are quite close-by and the type locality, Tura, lies at about 64 km northwest of the present study sites. However, our surveys at locations north of the Garo ridge at a higher elevation such as Mandalgre (25.50 °N, 90.37 °E, 1,019 m) and Daribokgre (25.48 °N, 90.31 °E, 1,123 m) could not locate this species. Further surveys at the vicinity of the current study sites and locations to the north of the Garo ridge in the lower reaches are necessary to determine whether the species occurs in those areas as well (Figure 3).

Abundance

The rate of encounter of *B. kempi* was 0.53/hour, or two hours of effort to locate one individual in this region.

The encounter rate across the survey duration of 57 hours ranged from 0.17–2.0 (Table 4).

Bufoides meghalayanus (Yazdani & Chanda, 1971)

Ansonia meghalayana Yazdani & Chanda, 1971

Holotype: ZSIC A 6969, an adult from Mawblang, Cherrapunji, Khasi Hills, Meghalaya.

Material studied: SACON VA 215, SACON VA 251 and SACON VA 252 three adult males from a hill stream in the Khasi Hills, Meghalaya (Image 7).

Diagnosis and comparison: A semi-arboreal to rupicolous *Bufoides* from the Khasi Hills diagnosed by: small–medium body size (SVL 31. –33.5 mm); presence of irregular, non-keratinized cranial ridges (pre and post orbital); short, elongated parotoid glands (vs. ovoid in *B. kempi*); absence of an externally visible tympanum; well-developed webbing between toes, with only half a



phalange free of webbing on toe IV (vs. relatively poor, with two phalanges free of webbing on toe IV in *B. kempi*); partial, but better developed webbing between fingers than *B. kempi* and the presence of small, slightly dilated, rounded terminal digital discs at the tips of both fingers and toes. Dorsum black with mossy green along the flanks in males, females predominantly green with black reticulations; a dark grey venter with small white spots (vs. pale white in *B. kempi*).

Description and Variation: (based on the newly collected material) Table 2

Body small (31.99 mm ± 0.76), trunk relatively short (AG:SVL 0.38) and stout (AG:BW 1.0). Head large (HL:SVL 0.37); slightly broader than long (HL:HW 0.85); and half as deep as long (HL:HD 2.08); snout tip obtusely pointed in dorsal view. Eyes large (ED:HL 0.35); snout slightly longer than eye (ED:ES 0.84); interorbital space about one and half times the width of the upper eyelid (IO:UEW 1.46) and nearly twice the internarial distance (IO:IN 2.09). Tympanum absent. Upper arm short (UAL:SVL 0.21); lower arm slightly longer than upper arm (UAL:LAL 0.93); palm a little longer (UAL:PAL 0.77). Fingers partially webbed, webbing formula I₀₋₁II₀₋₂III₁₋₁IV; relative length of fingers III>IV>II>I. Outer metacarpal tubercle large & evident. Thigh relatively short (FEL:SVL 0.39); tibia a little longer than the thigh (FEL:TBL 0.94); foot nearly as long as the thigh (FEL:FOL 0.98). Toes partially webbed, webbing formula: $I_{0-0}II_{0-0}III_{0-0.5}V_{0.5-0}V$, a relatively large inner and a slightly smaller ovoid metatarsal tubercles at the base of the foot. Toe tips with discs as broad as the toes; tarsal ridge not discernible. Finger and toe-tips bearing slightly expanded terminal discs lacking circummarginal grooves. Dorsal colouration uniform black, with irregular feeble yellow markings; venter grey with fine white spots.

Description of calls

The call of *B. meghalayanus* recorded during this study comprised of a series of high pitched syllables of 'ti-tuk' that lasted for a duration of 20 s and was composed of eight separated notes, each of which comprise two pulses. The mean duration of each note was 0.28 ms, with a mean interval of 2.45 s in between. Maximum amplitude of the call was -7 dB with a dominant frequency of 1.0 kHz (Image 8).

Distribution: (Table 4)

During this study, *B. meghalayanus* was recorded from a few locations in the Khasi hills within an altitudinal range of 1060–1240 m that are mapped in Figure 3. Additional localities were provided by Deuti et al. (2012).

Abundance

The rate of encounter of *B. meghalayanus* was 0.875 / hour, or about an hour of effort to locate one individual in this region. The encounter rate across the survey duration of 38 h ranged from 0.67–5.33 (Table 4).

DISCUSSION

B. kempi was described by Boulenger in 1919 based on the two specimens collected by S.W. Kemp, and presented to him by Nelson Annandale from 'above Tura, 2,500 ft' in the Garo Hills. Since its description, no further records or observations of this species have been made until now. Studies conducted in this region have uncovered several new and noteworthy species but B. kempi remained elusive to scientists until now (Datta-Roy et al. 2013; Deuti et al. 2012; Biju et al. 2016; Giri et al. 2019). Although Das et al. (2009) mentioned a specimen (MFA 10134) of Bufoides collected from Tura, Garo Hills, no taxonomic assessment of this specimen has been made until now, which still remains unidentified. Therefore, with the results of the present study, we announce the authentic rediscovery of B. kempi after a period of more than a century (1919-2022) from near the type locality, Garo Hills. First ever field observations on its ecology, behaviour, breeding biology, and natural history have been presented here. Observation on their breeding in phytotelmata and multiple males participating in amplexus with a single female have been made for the first time. Das & Dutta (2007) noted the absence of any larval descriptions for B. kempi, which has now been provided for the first time, across three developmental stages. Based on our field observations, the distribution of B. kempi mapped here shows that it is restricted to the lower reaches of a small hillock in the western part of Meghalaya, south of the Garo Hills, across an elevation range of 100-250 m. above which B. kempi was not detected despite intensive surveys. In comparison, B. meghalayanus was found to be a strictly montane species occurring only on the hilltops between elevations of 1,000-1,240 m. Further surveys north of the Garo ridge could possibly uncover additional locations characterized by lowland evergreen forests with perennial streams and rock boulders, that could potentially be occupied by B. kempi and we recommend additional surveys in such localities in the future.

Unlike *B. kempi*, for *B. meghalayanus*, the original descriptions of the species as well as the genus were comprehensive in terms of both morphology & natural history (Yazdani & Chanda 1971; Pillai & Yazdani 1973).





Image 7. Adult male *Bufoides meghalayanus* in life (dorsal and ventral views). © S.R. Chandramouli.

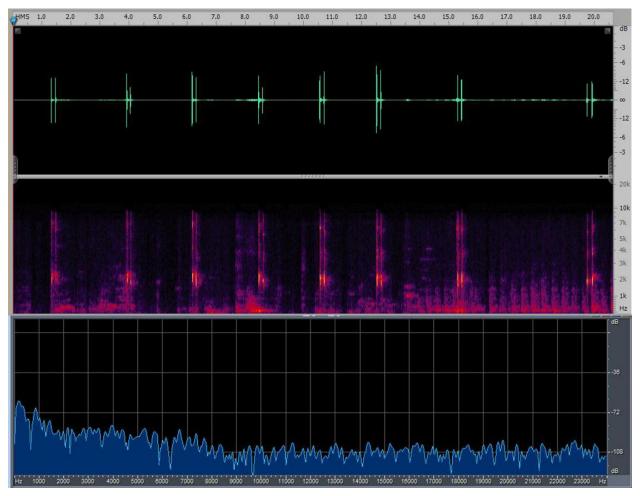


Image 8. Oscillogram, spectrogram and power spectrum of the call of *B. meghalayanus*.



Table 4. Abundance estimates of *B. kempi* and *B. meghalayanus*.

					В	ufoides ker	npi					
Site	Lat. (⁰N)	Long. (°E)	Elevation (m asl.)	time	Duration (h)	No. of ind.	ER	Microhabitat	Forest type	Habitat	Canopy cover	Ambient tempe- rature (°C)
Eman Asakgre	25.40	90.54	225	day	3	1	0.33	Under boulder	Evergreen	Dry stream	90	26
Eman Asakgre	25.36	90.53	122	day	6	1	0.17	Under boulder	Evergreen	Stream	90	27
Eman Asakgre	25.37	90.54	202	day	6	5	0.83	Tree hole	Evergreen	Forest	90	26
Eman Asakgre	25.36	90.53	122	night	6	2	0.33	Tree hole	Evergreen	Forest	90	23
Eman Asakgre	25.36	90.53	122	night	5	1	0.20	on leaf	Evergreen	Forest	90	23
Eman Asakgre	25.36	90.53	220	night	6	2	0.33	Tree hole	Evergreen	Forest	90	23
Eman Asakgre	25.36	90.53	220	night	6	2	0.33	on leaf	Evergreen	Forest	90	23
Eman Asakgre	25.36	90.53	220	night	6	4	0.67	on leaf/under boulder	Evergreen	Forest	90	23
Eman Asakgre	25.34	90.51	250	night	6	1	0.17	Under boulder	Evergreen	Forest	90	23
Eman Asakgre	25.37	90.54	202	night	6	3	0.50	on leaf	Evergreen	Forest	90	23
Eman Asakgre	25.36	90.53	122	night	1	2	2.00	Tree hole/on leaf	Evergreen	Forest	90	23
					57	24	0.53					
					Bufoi	des megha	layanus	T	ı			1
Stream behind Mawsmi cave	25.25	91.72	1200	Day/ Night	1	0	0		Montane	Stream	70	21
Stream behind Mawsmi cave	25.25	91.72	1200	Day/ Night	1	0	0		Montane	Stream	70	21
Stream 1 behind Mablang village	25.24	91.74	1200	Day/ Night	1	0	0		Montane	Stream	80	21
Stream 1 behind Mablang village	25.23	91.74	1200	Day/ Night	2	1	0.67	Rock Crevice	Montane	Stream	80	21
Stream behind Mawsmi cave	25.25	91.72	1200	Night	1	0	0		Montane	Stream	70	20
Stream 1 behind Mablang village	25.23	91.74	1200	Day/ Night	4	0	0		Montane	Stream	80	20
Stream behind Mawsmi cave	25.25	91.72	1200	Day/ Night	5	0	0		Montane	Stream	70	20
Stream 2 behind Mablang village	25.23	91.74	1200	Day/ Night	4	0	0		Montane	Stream	80	20
Stream 2 behind Mablang village	25.23	91.74	1200	Day	4	0	0		Montane	Stream	80	20
Stream 1 on-route to Thangkarank park	25.239	91.73	1200	Day	6	4	1.33	Rock Crevice	Montane	Dry stream	90	18
Stream 2 on-route to Thangkarank park	25.23	91.74	1200	Day/ Night	6	32	10.67	Rock Crevice / Pandanus tree	Montane	Dry stream	95	21
Stream 1 on-route to Thangkarank park	25.23	91.73	1200	Day	6	19	3.17	Rock Crevice	Montane	Dry stream	90	22
					38	56	0.875					



Subsequent studies have supplemented information on its morphology (Das et al. 2009), osteology (Chandramouli & Amarasinghe 2016); ecology and distribution (Deuti et al. 2012). Das et al. (2009) after examining the specimen reported by Pawar & Birand (2001) from Mizoram, opined that it is not conspecific with *B. meghalyanus*. Hence, we do not include that record within the range of *B. meghalyanus*. Therefore, the Mizoram population of *Bufoides* reported by Das et al. (2009) still needs a proper taxonomic assessment.

Availability of Data

Specimens collected and studied are deposited in the collections of SACON. DNA sequence generated in this study has been deposited in the genbank under the NCBI voucher number OP920605.

REFERENCES

- Barbour, T. (1938). Notes on Nectophryne. Proceedings of the Biological Society of Washington 51: 191–196.
- Biju, S.D., G. Senevirathne, S. Garg, S. Mahony, R.G. Kamei, A. Thomas, Y.S. Shouche, C.J. Raxworthy, M. Meegaskumbura & I.V. Bocxlaer (2016). Frankixalus, a New Rhacophorid Genus of Tree hole Breeding Frogs with Oophagous Tadpoles. PLoS ONE 11(1): e0145727. https://doi.org/10.1371/journal.pone.0145727
- Boulenger, G.A. (1919). Descriptions of three new batrachians from the Garo Hills, Assam. *Records of the Indian Museum* 16: 207–208. https://doi.org/10.5962/bhl.part.25921
- Chandramouli, S. R., K. Vasudevan, S. Harikrishnan, S. K. Dutta, S. J. Janani, R. Sharma, I. Das & R. K. Aggarwal (2016). A new genus and species of arboreal toad with phytotelmonous larvae, from the Andaman Islands, India (Lissamphibia, Anura, Bufonidae). *ZooKeys* 555: 57–90.
- Chandramouli, S.R. & A.A.T. Amarasinghe (2016). Taxonomic reassessment of the arboreal toad genus *Pedostibes* Günther 1876 (Anura: Bufonidae) and some allied oriental bufonid genera. *Herpetologica* 7: 137–147. https://doi.org/10.1655/HERPETOLOGICA-D-15-00053
- Chandramouli, S.R. & A. Kalaimani (2014). Description of the larvae of Günther's toad *Duttaphrynus hololius* (Günther, 1876) (Anura: Bufonidae) with notes on development and oral ultra-structure. *Alytes*, 31: 3–12.
- Das, I. & S.K. Dutta (2007). Sources of larval identities for amphibians of India. *Hamadryad* 31: 152–181.
- Das, I., D. Rangad, R.K.L. Tron, K. Deuti & R.N.K. Hooroo (2009).
 Rediscovery of the endangered Khasi Hills rock toad, *Bufoides meghalayana* in Meghalaya, Northeastern India. *Froglog* 92: 1–4.
- Datta-Roy, A., I. Das, A.M. Bauer, R.K.L. Tron & P. Karanth (2013).

 Lizard Wears Shades. A Spectacled Sphenomorphus (Squamata:

- Scincidae), from the Sacred Forests of Mawphlang, Meghalaya, North-east India. *Zootaxa*, 3701 (2): 257–276. https://doi.org/10.11646/zootaxa.3701.2.7
- Deuti, K., S. Rey & S.K. Dey (2012). Status survey of the Khasi Hills rock toad (*Bufoides meghalayana*) at Cherrapunjee, Meghalaya. *Records of Zoological Survey of India* III: 21–25. https://doi.org/10.26515/rzsi/v111/i3/2011/158851
- Edgar, R. C. (2004). MUSCLE: multiple sequence alignment with high accuracy and high throughput. *Nucleic acids research* 32(5): 1792–1797. https://doi.org/10.1093/nar/gkh340
- Frost, D.R. (2022). Amphibian Species of the World: an Online Reference. Version 6.1 (Date of access 10 Mar 2022). Electronic Database accessible at https://amphibiansoftheworld.amnh.org/index.php. American Museum of Natural History, New York, USA. https://doi.org/10.5531/db.vz.0001
- Giri, V.B., R. Chaitanya, S. Mahony, S. Lalrounga, C. Lalrinchhana, A. Das, V. Sarkar, P. Karanth & V. Deepak (2019). On the systematic status of the genus *Oriocalotes* Günther, 1864 (Squamata: Agamidae: Draconinae) with the description of a new species from Mizoram state, Northeast India. *Zootaxa* 4638 (4): 451–484. https://doi.org/10.11646/zootaxa.4638.4.1
- **Gosner, K.L. (1960).** A simplified table for staging anuran embryos and larvae with notes on identification. *Herpetologica*, 16: 183–190.
- Kocher, T.D., W.K. Thomas, A. Meyer, S.V. Edwards, S. Pääbo, F.X. Villablanca & A.C. Wilson (1989). Dynamics of mitochondrial DNA evolution in animals: amplification and sequencing with conserved primers. *Proceedings of the National Academy of Sciences of the United States of America*, 86: 6192–6200. https://doi.org/10.1073/pnas.86.16.6196
- McDiarmid, R.W. & R. Altig (1999). *Tadpoles: the biology of anuran larvae*. University of Chicago Press, Chicago, 476pp.
- **Noble, G.K. (1931).** *The Biology of the Amphibia.* McGraw Hill Book Company, London: 577pp.
- Pillai, R.S. & G.M. Yazdani (1973). Bufoides, a new genus for the rock-toad, Ansonia meghalayana Yazdani & Chanda, with notes on its ecology and breeding habits. Journal of the Zoological Society of India 25: 65–70.
- Ronquist, F. & J.P. Huelsenbeck (2003). MrBayes 3: Bayesian phylogenetic inference under mixed models. *Bioinformatics* 19: 1572–1574. https://doi.org/10.1093/bioinformatics/btg180
- **Rödel, M.-O. (2000).** Herpetofauna of West Africa. Volume I: Amphibians of the West African Savanna. Frankfurt am Main (Edition Chimaira): 332 pp.
- Savage, J.M. & W.R. Heyer (1997). Digital webbing formulae for anurans: a refinement. *Herpetological Review* 28: 131.
- Stamatakis, A. (2006). RAXML-VI-HPC: Maximum likelihood-based phylogenetic analyses with thousands of taxa and mixed models. *Bioinformatics* 22: 2688–2690. https://doi.org/10.1093/bioinformatics/btl446
- Tamura, K., G. Stecher, D. Peterson, A. Filipski, & S. Kumar (2013).
 MEGA6: Molecular Evolutionary Genetics Analysis version 6.0.
 Molecular Biology and Evolution 30 (12): 2725–2729. https://doi.org/10.1093/molbey/mst197
- Yazdani, G.M. & S.K. Chanda (1971). A new toad, Ansonia meghalayana (family Bufonidae) from Meghalaya (Assam), India, with observations on its breeding on Pandanus furcatus Roxb. (Pandanales: Pandanaceae). Journal of the Assam Science Society 14: 76–80.

ZOOREACH

- Dr. George Mathew, Kerala Forest Research Institute, Peechi, India
- Dr. John Noyes, Natural History Museum, London, UK
- Dr. Albert G. Orr, Griffith University, Nathan, Australia
- Dr. Sameer Padhye, Katholieke Universiteit Leuven, Belgium
- Dr. Nancy van der Poorten, Toronto, Canada Dr. Kareen Schnabel, NIWA, Wellington, New Zealand
- Dr. R.M. Sharma, (Retd.) Scientist, Zoological Survey of India, Pune, India
- Dr. Manju Siliwal, WILD, Coimbatore, Tamil Nadu, India
- Dr. G.P. Sinha, Botanical Survey of India, Allahabad, India
- Dr. K.A. Subramanian, Zoological Survey of India, New Alipore, Kolkata, India
- Dr. P.M. Sureshan, Zoological Survey of India, Kozhikode, Kerala, India
- Dr. R. Varatharajan, Manipur University, Imphal, Manipur, India Dr. Eduard Vives, Museu de Ciències Naturals de Barcelona, Terrassa, Spain
- Dr. James Young, Hong Kong Lepidopterists' Society, Hong Kong
- Dr. R. Sundararaj, Institute of Wood Science & Technology, Bengaluru, India
- Dr. M. Nithyanandan, Environmental Department, La Ala Al Kuwait Real Estate. Co. K.S.C.,
- Dr. Himender Bharti, Punjabi University, Punjab, India
- Mr. Purnendu Roy, London, UK
- Dr. Saito Motoki, The Butterfly Society of Japan, Tokyo, Japan Dr. Sanjay Sondhi, TITLI TRUST, Kalpavriksh, Dehradun, India
- Dr. Nguyen Thi Phuong Lien, Vietnam Academy of Science and Technology, Hanoi, Vietnam
- Dr. Nitin Kulkarni, Tropical Research Institute, Jabalpur, India
- Dr. Robin Wen Jiang Ngiam, National Parks Board, Singapore
- Dr. Lional Monod, Natural History Museum of Geneva, Genève, Switzerland.
- Dr. Asheesh Shivam, Nehru Gram Bharti University, Allahabad, India
- Dr. Rosana Moreira da Rocha, Universidade Federal do Paraná, Curitiba, Brasil
- Dr. Kurt R. Arnold, North Dakota State University, Saxony, Germany
- Dr. James M. Carpenter, American Museum of Natural History, New York, USA
- Dr. David M. Claborn, Missouri State University, Springfield, USA
- Dr. Kareen Schnabel, Marine Biologist, Wellington, New Zealand
- Dr. Amazonas Chagas Júnior, Universidade Federal de Mato Grosso, Cuiabá, Brasil
- Mr. Monsoon Jyoti Gogoi, Assam University, Silchar, Assam, India
- Dr. Heo Chong Chin, Universiti Teknologi MARA (UiTM), Selangor, Malaysia
- Dr. R.J. Shiel, University of Adelaide, SA 5005, Australia
- Dr. Siddharth Kulkarni, The George Washington University, Washington, USA
- Dr. Priyadarsanan Dharma Rajan, ATREE, Bengaluru, India
- Dr. Phil Alderslade, CSIRO Marine And Atmospheric Research, Hobart, Australia
- Dr. John E.N. Veron, Coral Reef Research, Townsville, Australia
- Dr. Daniel Whitmore, State Museum of Natural History Stuttgart, Rosenstein, Germany.
- Dr. Yu-Feng Hsu, National Taiwan Normal University, Taipei City, Taiwan
- Dr. Keith V. Wolfe, Antioch, California, USA
- Dr. Siddharth Kulkarni, The Hormiga Lab, The George Washington University, Washington,
- Dr. Tomas Ditrich, Faculty of Education, University of South Bohemia in Ceske
- Budeiovice, Czech Republic
- Dr. Mihaly Foldvari, Natural History Museum, University of Oslo, Norway
- Dr. V.P. Uniyal, Wildlife Institute of India, Dehradun, Uttarakhand 248001, India
- Dr. John T.D. Caleb, Zoological Survey of India, Kolkata, West Bengal, India
- Dr. Priyadarsanan Dharma Rajan, Ashoka Trust for Research in Ecology and the Environment (ATREE), Royal Enclave, Bangalore, Karnataka, India

Fishes

- Dr. Neelesh Dahanukar, IISER, Pune, Maharashtra, India
- Dr. Topiltzin Contreras MacBeath, Universidad Autónoma del estado de Morelos, México
- Dr. Heok Hee Ng, National University of Singapore, Science Drive, Singapore
- Dr. Rajeev Raghavan, St. Albert's College, Kochi, Kerala, India
- Dr. Robert D. Sluka, Chiltern Gateway Project, A Rocha UK, Southall, Middlesex, UK
- Dr. E. Vivekanandan, Central Marine Fisheries Research Institute, Chennai, India
- Dr. Davor Zanella, University of Zagreb, Zagreb, Croatia
- Dr. A. Biju Kumar, University of Kerala, Thiruvananthapuram, Kerala, India
- Dr. Akhilesh K.V., ICAR-Central Marine Fisheries Research Institute, Mumbai Research
- Centre, Mumbai, Maharashtra, India
- Dr. J.A. Johnson, Wildlife Institute of India, Dehradun, Uttarakhand, India
- Dr. R. Ravinesh, Gujarat Institute of Desert Ecology, Gujarat, India

Amphibians

- Dr. Sushil K. Dutta, Indian Institute of Science, Bengaluru, Karnataka, India
- Dr. Annemarie Ohler, Muséum national d'Histoire naturelle, Paris, France

Reptiles

- Dr. Gernot Vogel, Heidelberg, Germany
- Dr. Raju Vyas, Vadodara, Gujarat, India
- Dr. Pritpal S. Soorae, Environment Agency, Abu Dubai, UAE.
- Prof. Dr. Wayne J. Fuller, Near East University, Mersin, Turkey
- Prof. Chandrashekher U. Rivonker, Goa University, Taleigao Plateau, Goa. India
- Dr. S.R. Ganesh, Chennai Snake Park, Chennai, Tamil Nadu, India
- Dr. Himansu Sekhar Das, Terrestrial & Marine Biodiversity, Abu Dhabi, UAE

Journal of Threatened Taxa is indexed/abstracted in Bibliography of Systematic Mycology, Biological Abstracts, BIOSIS Previews, CAB Abstracts, EBSCO, Google Scholar, Index Copernicus, Index Fungorum, JournalSeek, National Academy of Agricultural Sciences, NewJour, OCLC WorldCat, SCOPUS, Stanford University Libraries, Virtual Library of Biology, Zoological Records.

NAAS rating (India) 5.64

Birds

- Dr. Hem Sagar Baral, Charles Sturt University, NSW Australia
- Mr. H. Byju, Coimbatore, Tamil Nadu, India
- Dr. Chris Bowden, Royal Society for the Protection of Birds, Sandy, UK
- Dr. Priya Davidar, Pondicherry University, Kalapet, Puducherry, India
- Dr. J.W. Duckworth, IUCN SSC, Bath, UK
- Dr. Rajah Jayapal, SACON, Coimbatore, Tamil Nadu, India
- Dr. Rajiv S. Kalsi, M.L.N. College, Yamuna Nagar, Haryana, India
- Dr. V. Santharam, Rishi Valley Education Centre, Chittoor Dt., Andhra Pradesh, India
- Dr. S. Balachandran, Bombay Natural History Society, Mumbai, India
- Mr. J. Praveen, Bengaluru, India
- Dr. C. Srinivasulu, Osmania University, Hyderabad, India
- Dr. K.S. Gopi Sundar, International Crane Foundation, Baraboo, USA
- Dr. Gombobaatar Sundev, Professor of Ornithology, Ulaanbaatar, Mongolia
- Prof. Reuven Yosef, International Birding & Research Centre, Eilat, Israel
- Dr. Taej Mundkur, Wetlands International, Wageningen, The Netherlands
- Dr. Carol Inskipp, Bishop Auckland Co., Durham, UK
- Dr. Tim Inskipp, Bishop Auckland Co., Durham, UK Dr. V. Gokula, National College, Tiruchirappalli, Tamil Nadu, India
- Dr. Arkady Lelej, Russian Academy of Sciences, Vladivostok, Russia
- Dr. Simon Dowell, Science Director, Chester Zoo, UK
- Dr. Mário Gabriel Santiago dos Santos, Universidade de Trás-os-Montes e Alto Douro,
- Quinta de Prados, Vila Real, Portugal
- Dr. Grant Connette, Smithsonian Institution, Royal, VA, USA
- Dr. M. Zafar-ul Islam, Prince Saud Al Faisal Wildlife Research Center, Taif, Saudi Arabia

- Dr. Giovanni Amori, CNR Institute of Ecosystem Studies, Rome, Italy
- Dr. Anwaruddin Chowdhury, Guwahati, India
- Dr. David Mallon, Zoological Society of London, UK
- Dr. Shomita Mukherjee, SACON, Coimbatore, Tamil Nadu, India
- Dr. Angie Appel, Wild Cat Network, Germany
- Dr. P.O. Nameer, Kerala Agricultural University, Thrissur, Kerala, India
- Dr. Ian Redmond, UNEP Convention on Migratory Species, Lansdown, UK
- Dr. Heidi S. Riddle, Riddle's Elephant and Wildlife Sanctuary, Arkansas, USA
- Dr. Karin Schwartz, George Mason University, Fairfax, Virginia.
- Dr. Lala A.K. Singh, Bhubaneswar, Orissa, India
- Dr. Mewa Singh, Mysore University, Mysore, India
- Dr. Paul Racey, University of Exeter, Devon, UK
- Dr. Honnavalli N. Kumara, SACON, Anaikatty P.O., Coimbatore, Tamil Nadu, India
- Dr. Nishith Dharaiya, HNG University, Patan, Gujarat, India
- Dr. Spartaco Gippoliti, Socio Onorario Società Italiana per la Storia della Fauna "Giuseppe Altobello", Rome, Italy
- Dr. Justus Joshua, Green Future Foundation, Tiruchirapalli, Tamil Nadu, India
- Dr. H. Raghuram, The American College, Madurai, Tamil Nadu, India
- Dr. Paul Bates, Harison Institute, Kent, UK
- Dr. Jim Sanderson, Small Wild Cat Conservation Foundation, Hartford, USA Dr. Dan Challender, University of Kent, Canterbury, UK
- Dr. David Mallon, Manchester Metropolitan University, Derbyshire, UK
- $\hbox{Dr. Brian L. Cypher, California State University-Stanislaus, Bakersfield, CA}$
- Dr. S.S. Talmale, Zoological Survey of India, Pune, Maharashtra, India Prof. Karan Bahadur Shah, Budhanilakantha Municipality, Kathmandu, Nepal
- Dr. Susan Cheyne, Borneo Nature Foundation International, Palangkaraja, Indonesia Dr. Hemanta Kafley, Wildlife Sciences, Tarleton State University, Texas, USA

Other Disciplines

- Dr. Aniruddha Belsare, Columbia MO 65203, USA (Veterinary)
- Dr. Mandar S. Paingankar, University of Pune, Pune, Maharashtra, India (Molecular) Dr. Jack Tordoff, Critical Ecosystem Partnership Fund, Arlington, USA (Communities)
- Dr. Ulrike Streicher, University of Oregon, Eugene, USA (Veterinary)
- Dr. Hari Balasubramanian, EcoAdvisors, Nova Scotia, Canada (Communities) Dr. Rayanna Hellem Santos Bezerra, Universidade Federal de Sergipe, São Cristóvão, Brazil
- Dr. Jamie R. Wood, Landcare Research, Canterbury, New Zealand Dr. Wendy Collinson-Jonker, Endangered Wildlife Trust, Gauteng, South Africa
- Dr. Rajeshkumar G. Jani, Anand Agricultural University, Anand, Gujarat, India Dr. O.N. Tiwari, Senior Scientist, ICAR-Indian Agricultural Research Institute (IARI), New
- Dr. L.D. Singla, Guru Angad Dev Veterinary and Animal Sciences University, Ludhiana, India
- Dr. Rupika S. Rajakaruna, University of Peradeniya, Peradeniya, Sri Lanka Dr. Bahar Baviskar, Wild-CER, Nagpur, Maharashtra 440013, India

Reviewers 2019-2021 Due to pausity of space, the list of reviewers for 2018–2020 is available online.

The opinions expressed by the authors do not reflect the views of the Journal of Threatened Taxa, Wildlife Information Liaison Development Society, Zoo Outreach Organization, or any of the partners. The journal, the publisher, the host, and the partners are not responsible for the accuracy of the political boundaries shown in the maps by the authors.

Print copies of the Journal are available at cost. Write to:

- The Managing Editor, JoTT,
- c/o Wildlife Information Liaison Development Society,
- 43/2 Varadarajulu Nagar, 5th Street West, Ganapathy, Coimbatore,
- Tamil Nadu 641035, India
- ravi@threatenedtaxa.org





The Journal of Threatened Taxa (JoTT) is dedicated to building evidence for conservation globally by publishing peer-reviewed articles online every month at a reasonably rapid rate at www.threatenedtaxa.org. All articles published in JoTT are registered under Creative Commons Attribution 4.0 International License unless otherwise mentioned. JoTT allows allows unrestricted use, reproduction, and distribution of articles in any medium by providing adequate credit to the author(s) and the source of publication.

ISSN 0974-7907 (Online) | ISSN 0974-7893 (Print)

December 2022 | Vol. 14 | No. 12 | Pages: 22207-22354

Date of Publication: 26 December 2022 (Online & Print)

DOI: 10.11609/jott.2022.14.12.22207-22354

Communications

A preliminary survey of moss flora of Chail Wildlife Sanctuary, Himachal Pradesh, India

- Meenal Sharma, Anju Rao & S.S. Kumar, Pp. 22207-22214

New distribution record and DNA barcoding of *Sapria himalayana* Griff. (Rafflesiaceae), a rare and endangered holoparasitic plant from Mizoram, India

Laldinfeli Ralte, Hmingremhlua Sailo, Sagolshem Priyokumar
 Singh, Laldinliana Khiangte & Y. Tunginba Singh, Pp. 22215–
 22220

Species distribution modeling of a cucurbit *Herpetospermum darjeelingense* in Darjeeling Himalaya, India

- Debasruti Boral & Saurav Moktan, Pp. 22221-22231

An updated catalogue of true flies (Insecta: Diptera) from northern Pakistan

- Noor Fatima & Ding Yang, Pp. 22232-22259

Desert Carabidae (Insecta: Coleoptera) of India

- S.V. Akhil, Sabu K. Thomas & Sanjeev Kumar, Pp. 22260-22269

Photographic evidence of fish assemblage in artificial reef site of Palk Bay - an implication for marine resource management

Koushik Sadhukhan, T. Shanmugaraj, Ramesh Chatragadda &
 M.V. Ramana Murthy, Pp. 22270–22276

Systematics of the enigmatic and narrowly endemic toad genus *Bufoides* Pillai & Yazdani, 1973: rediscovery of *Bufoides kempi* (Boulenger, 1919) and expanded description of *Bufoides meghalayanus* (Yazdani & Chanda, 1971) (Amphibia: Anura: Bufonidae) with notes on natural history and distribution

R.S. Naveen, S.R. Chandramouli, Gautam Kadam, S. Babu,
 P.V. Karunakaran, H.N. Kumara & N. Parthasarathy, Pp. 22277–22292

Avifaunal diversity in Indian Institute of Technology Guwahati Campus, Assam, India

- Umang H. Rathod & Rupam Bhaduri, Pp. 22293-22308

Reviews

Threatened flora of Uttarakhand: an update

– D.S. Rawat, Satish Chandra & Preeti Chaturvedi, Pp. 22309– 22328

A systematic review on the feeding ecology of Sloth Bear Melursus ursinus Shaw, 1791 in its distribution range in the Indian subcontinent

- Vasantkumar Rabari & Nishith Dharaiya, Pp. 22329-22336

Short Communications

Mercury in tuna from the western equatorial Atlantic Ocean and health risk assessment

 Ana Paula Madeira Di Beneditto, Inácio Abreu Pestana, Igor
 David da Costa, Marcelo Gomes de Almeida, Braulio Cherene Vaz de Oliveira & Carlos Eduardo de Rezende, Pp. 22337–22340

First photographic record of Spotted Deer Axis axis (Erxleben, 1777) (Artiodactyla: Cervidae) in Great Indian Bustard Sanctuary, Maharashtra, India

– Shaheer Khan, S. Ramesh Kumar & Bilal Habib, Pp. 22341–22345

Comparative study of morphology and keratin levels in hair from deer and goat

– Sangeeta Patle, Divya Bagchi & K.P. Singh, Pp. 22346–22350

Response & Reply

Is trade the reason for the unusual colour morph of Cobra from Goa? Response to Sawant et al.

– Raju Vyas & Harshil Patel, Pp. 22351–22353

Corrections to 'An unusual morph of *Naja naja* (Linnaeus, 1758) from Goa, India (Serpentes: Squamata)'

 Nitin Sawant, Amrut Singh, Shubham Rane, Sagar Naik & Mayur Gawas, P. 22354

Publisher & Host

