10.11609/jott.2022.14.6.21127-21330 www.threatenedtaxa.org

> 26 June 2022 (Online 5 Print) 14 (6): 21127-21330 ISSN 0974-7907 (Online) ISSN 0974-7893 (Print)

> > Open Access

got conservation globally Journal or Threat





Publisher

Wildlife Information Liaison Development Society www.wild.zooreach.org

Host **Zoo Outreach Organization** www.zooreach.org

No. 12, Thiruvannamalai Nagar, Saravanampatti - Kalapatti Road, Saravanampatti, Coimbatore, Tamil Nadu 641035, India Ph: +91 9385339863 | www.threatenedtaxa.org

Email: sanjay@threatenedtaxa.org

EDITORS

Founder & Chief Editor

Dr. Sanjay Molur

Wildlife Information Liaison Development (WILD) Society & Zoo Outreach Organization (ZOO), 12 Thiruvannamalai Nagar, Saravanampatti, Coimbatore, Tamil Nadu 641035, India

Deputy Chief Editor

Dr. Neelesh Dahanukar Noida, Uttar Pradesh, India

Managing Editor

Mr. B. Ravichandran, WILD/ZOO, Coimbatore, India

Associate Editors

Dr. Mandar Paingankar, Government Science College Gadchiroli, Maharashtra 442605, India Dr. Ulrike Streicher, Wildlife Veterinarian, Eugene, Oregon, USA Ms. Privanka Iver. ZOO/WILD. Coimbatore. Tamil Nadu 641035. India Dr. B.A. Daniel, ZOO/WILD, Coimbatore, Tamil Nadu 641035, India

Editorial Board

Dr. Russel Mittermeier

Executive Vice Chair, Conservation International, Arlington, Virginia 22202, USA

Prof. Mewa Singh Ph.D., FASc, FNA, FNASc, FNAPsy

Ramanna Fellow and Life-Long Distinguished Professor, Biopsychology Laboratory, and Institute of Excellence, University of Mysore, Mysuru, Karnataka 570006, India; Honorary Professor, Jawaharlal Nehru Centre for Advanced Scientific Research, Bangalore; and Adjunct Professor, National Institute of Advanced Studies, Bangalore

Stephen D. Nash

Scientific Illustrator, Conservation International, Dept. of Anatomical Sciences, Health Sciences Center, T-8, Room 045, Stony Brook University, Stony Brook, NY 11794-8081, USA

Dr. Fred Pluthero

Toronto, Canada

Dr. Priya Davidar

Sigur Nature Trust, Chadapatti, Mavinhalla PO, Nilgiris, Tamil Nadu 643223, India

Dr. Martin Fisher

Senior Associate Professor, Battcock Centre for Experimental Astrophysics, Cavendish Laboratory, JJ Thomson Avenue, Cambridge CB3 OHE, UK

Dr. John Fellowes

Honorary Assistant Professor, The Kadoorie Institute, 8/F, T.T. Tsui Building, The University of Hong Kong, Pokfulam Road, Hong Kong

Prof. Dr. Mirco Solé

Universidade Estadual de Santa Cruz, Departamento de Ciências Biológicas, Vice-coordenador do Programa de Pós-Graduação em Zoologia, Rodovia Ilhéus/Itabuna, Km 16 (45662-000) Salobrinho. Ilhéus - Bahia - Brasil

Dr. Rajeev Raghavan

Professor of Taxonomy, Kerala University of Fisheries & Ocean Studies, Kochi, Kerala, India

English Editors Mrs. Mira Bhojwani, Pune, India

Dr. Fred Pluthero, Toronto, Canada Mr. P. Ilangovan, Chennai, India

Web Development

Mrs. Latha G. Ravikumar, ZOO/WILD, Coimbatore, India Typesetting

Mr. Arul Jagadish. ZOO, Coimbatore, India Mrs. Radhika, ZOO, Coimbatore, India Mrs. Geetha, ZOO, Coimbatore India

Fundraising/Communications Mrs. Payal B. Molur, Coimbatore, India

Subject Editors 2019-2021

Fungi

- Dr. B. Shivaraju, Bengaluru, Karnataka, India
- Dr. R.K. Verma, Tropical Forest Research Institute, Jabalpur, India
- Dr. Vatsavaya S. Raju, Kakatiay University, Warangal, Andhra Pradesh, India
- Dr. M. Krishnappa, Jnana Sahyadri, Kuvempu University, Shimoga, Karnataka, India
- Dr. K.R. Sridhar, Mangalore University, Mangalagangotri, Mangalore, Karnataka, India Dr. Gunjan Biswas, Vidyasagar University, Midnapore, West Bengal, India

Plants

- Dr. G.P. Sinha, Botanical Survey of India, Allahabad, India
- Dr. N.P. Balakrishnan, Ret, Joint Director, BSI, Coimbatore, India
- Dr. Shonil Bhagwat, Open University and University of Oxford, UK
- Prof. D.J. Bhat, Retd. Professor, Goa University, Goa, India
- Dr. Ferdinando Boero, Università del Salento, Lecce, Italy
- Dr. Dale R. Calder, Royal Ontaro Museum, Toronto, Ontario, Canada
- Dr. Cleofas Cervancia, Univ. of Philippines Los Baños College Laguna, Philippines
- Dr. F.B. Vincent Florens, University of Mauritius, Mauritius Dr. Merlin Franco, Curtin University, Malaysia
- Dr. V. Irudayaraj, St. Xavier's College, Palayamkottai, Tamil Nadu, India
- Dr. B.S. Kholia, Botanical Survey of India, Gangtok, Sikkim, India
- Dr. Pankaj Kumar, Kadoorie Farm and Botanic Garden Corporation, Hong Kong S.A.R., China
- Dr. V. Sampath Kumar, Botanical Survey of India, Howrah, West Bengal, India
- Dr. A.J. Solomon Raju, Andhra University, Visakhapatnam, India
- Dr. Vijayasankar Raman, University of Mississippi, USA
- Dr. B. Ravi Prasad Rao, Sri Krishnadevaraya University, Anantpur, India Dr. K. Ravikumar, FRLHT, Bengaluru, Karnataka, India
- Dr. Aparna Watve, Pune, Maharashtra, India
- Dr. Qiang Liu, Xishuangbanna Tropical Botanical Garden, Yunnan, China
- Dr. Noor Azhar Mohamed Shazili, Universiti Malaysia Terengganu, Kuala Terengganu, Malaysia Dr. M.K. Vasudeva Rao, Shiv Ranjani Housing Society, Pune, Maharashtra, India
- Prof. A.J. Solomon Raju, Andhra University, Visakhapatnam, India
- Dr. Mandar Datar, Agharkar Research Institute, Pune, Maharashtra, India
- Dr. M.K. Janarthanam. Goa University. Goa. India
- Dr. K. Karthigeyan, Botanical Survey of India, India
- Dr. Errol Vela, University of Montpellier, Montpellier, France
- Dr. P. Lakshminarasimhan, Botanical Survey of India, Howrah, India
- Dr. Larry R. Noblick, Montgomery Botanical Center, Miami, USA
- Dr. K. Haridasan, Pallavur, Palakkad District, Kerala, India
- Dr. Analinda Manila-Fajard, University of the Philippines Los Banos, Laguna, Philippines
- Dr. P.A. Sinu, Central University of Kerala, Kasaragod, Kerala, India
- Dr. Afroz Alam, Banasthali Vidyapith (accredited A grade by NAAC), Rajasthan, India
- Dr. K.P. Rajesh, Zamorin's Guruvayurappan College, GA College PO, Kozhikode, Kerala, India
- Dr. David E. Boufford, Harvard University Herbaria, Cambridge, MA 02138-2020, USA
- Dr. Ritesh Kumar Choudhary, Agharkar Research Institute, Pune, Maharashtra, India Dr. Navendu Page, Wildlife Institute of India, Chandrabani, Dehradun, Uttarakhand, India

Invertebrates

- Dr. R.K. Avasthi, Rohtak University, Haryana, India
- Dr. D.B. Bastawade, Maharashtra, India
- Dr. Partha Pratim Bhattacharjee, Tripura University, Suryamaninagar, India
- Dr. Kailash Chandra, Zoological Survey of India, Jabalpur, Madhya Pradesh, India
- Dr. Ansie Dippenaar-Schoeman, University of Pretoria, Queenswood, South Africa
- Dr. Rory Dow, National Museum of natural History Naturalis, The Netherlands
- Dr. Brian Fisher, California Academy of Sciences, USA Dr. Richard Gallon, llandudno, North Wales, LL30 1UP
- Dr. Hemant V. Ghate, Modern College, Pune, India
- Dr. M. Monwar Hossain, Jahangirnagar University, Dhaka, Bangladesh
- Mr. Jatishwor Singh Irungbam, Biology Centre CAS, Branišovská, Czech Republic.
- Dr. Ian J. Kitching, Natural History Museum, Cromwell Road, UK
- Dr. George Mathew, Kerala Forest Research Institute, Peechi, India

For Focus, Scope, Aims, and Policies, visit https://threatenedtaxa.org/index.php/JoTT/aims_scope For Article Submission Guidelines, visit https://threatenedtaxa.org/index.php/JoTT/about/submissions For Policies against Scientific Misconduct, visit https://threatenedtaxa.org/index.php/JoTT/policies_various

continued on the back inside cover

Cover: Euphaea pseudodispar shot at Kalindi River, Thirunelly, Wayanad district, Kerala. © Muneer P.K.

Journal of Threatened Taxa | www.threatenedtaxa.org | 26 June 2022 | 14(6): 21246-21265

ISSN 0974-7907 (Online) | ISSN 0974-7893 (Print) https://doi.org/10.11609/jott.7487.14.6.21246-21265

#7487 | Received 02 June 2021 | Final received 02 February 2022 | Finally accepted 20 May 2022

The study of algal diversity from fresh water bodies of Chimmony Wildlife Sanctuary, Kerala, India

Joel Jose¹ 💿 & Jobi Xavier² 💿

^{1,2} Department of Life Sciences, CHRIST (Deemed to be University), Dharmaram College P.O, Hosur Road, Bengaluru, Karnataka 560029, India. ¹ joeljose701@gmail.com, ² frjobi.xavier@christuniversity.in (corresponding author)

Abstract: The algal diversity of the freshwater ecosystem is very significant because they are the primary energy producers in the food web. The study for the algal diversity was conducted at Chimmony Wildlife Sanctuary, Thrissur, Kerala, India, from selected sampling sites (Pookoyil thodu, Kidakkapara thodu, Viraku thodu, Nellipara thodu, Anaporu thodu, Kodakallu thodu, Odan thodu, Mullapara thodu, Payampara thodu, Chimmony dam). The identified algal species belong to four different classes: Chlorophyceae, Euglenineae, Rhodophyceae, and Cyanophyceae. Sixty-one algal species were identified, represented by 37 genera, 22 families, and 14 orders. Among the four, Chlorophyceae was the dominant class.

Keywords: Biodiversity, Chlorophyceae, conservation, Cyanophyceae, Euglenineae, freshwater, Rhodophyceae, taxonomy, Thrissur.

Editor: Anonymity requested.

Date of publication: 26 June 2022 (online & print)

OPEN

ACCESS

()

Citation: Jose, J. & J. Xavier (2022). The study of algal diversity from fresh water bodies of Chimmony Wildlife Sanctuary, Kerala, India. Journal of Threatened Taxa 14(6): 21246–21265. https://doi.org/10.11609/jott.7487.14.6.21246-21265

Copyright: © Jose & Xavier 2022. Creative Commons Attribution 4.0 International License. JoTT allows unrestricted use, reproduction, and distribution of this article in any medium by providing adequate credit to the author(s) and the source of publication.

Funding: None.

Competing interests: The authors declare no competing interests.

Author details: JOEL JOSE (JJ) has completed post-graduation in Botany from CHRIST (Deemed to be University), Bengaluru. He is currently a research scholar in the Department of Life Sciences at CHRIST (Deemed to be University). He is passionate about the algae taxonomy, stress physiology, plant taxonomy, and environmental studies. JOBI XAVIER (JX) has completed post-graduation and doctoral degree in botany. He is working as Associate Professor in the Department of Life Sciences at CHRIST (Deemed to be University). His area of interest is phytochemistry, plant taxonomy, plant ecology, and plant physiology. He has authored a book on Trees of CHRIST University.

Author contributions: JJ did the experimental design, field data collection and the data analysis. JX has supervised the work and preparation of final manuscript.

Acknowledgements: We greatly thank the Kerala Forest Department and other forest officials for the permission for fieldwork in Chimmony Wildlife Sanctuary. We also thank for the co-operation and support by management and staff of the CHRIST (Deemed to be University), Bangalore.

INTRODUCTION

Algae are the most abundant aquatic organisms present in the freshwater ecosystem. Algae were responsible for the beginning of multicellular life on our planet and could be the key to our future survival. They are an essential source for producing fine chemicals, natural pigments, vitamins, polysaccharides, bioflocculants, and growth promoters. Algae are also a significant producers of oxygen than plants (Rai et al. 2000).

The freshwater ecosystems are mainly categorized into two types: lotic and lentic. The rivers, streams, waterfalls, canals fall into the lotic type, and the stagnant waters like pools, lakes, reservoirs and paddy fields fall into the lentic type. The freshwater algal diversity varies from unicellular phytoplankton to colonial and much larger multicellular algae. The algal biodiversity depends upon the physicochemical parameters of the water bodies. In the food chain of aquatic ecosystems, algae are the primary producers, making them very important. So the conservation and knowledge about algal biodiversity are necessary for maintaining a healthy aquatic ecosystem.

The information regarding species diversity is an essential component to realize life in its fullness and conserve it for future generations (Pandey 1995). Therefore, there is a strong demand for research on biodiversity in developing countries (Briji 2005; Tessy &

Sreekumar 2017). Generally, the taxonomy is considered an outdated science that cannot keep up with the present biodiversity crisis (De Clerck et al. 2013). But for the future development in biodiversity research, systematics and taxonomy are important (Koen & Segers 2005).

The study of biodiversity as the present one opens new opportunities to understand the different algal forms in their respective natural habitat. The large algal species in the freshwater ecosystem depict its diversity. In the current scenario, hardly a few genera are used in the industry, giving a broad scope for other potential obtainable algae. Even though plenty of literature is available on fresh water algal diversity of Kerala, there is no published record available on the algal diversity of Chimmony Wildlife Sanctuary. Hence the study.

MATERIALS AND METHODS

Study area

Collection Sites

The study was conducted in Chimmony Wildlife Sanctuary (CWS; Figure 1), which is situated in the Thrissur District of Kerala state. It belongs to Mukundapuram taluk and within geographical limits of 10.40° & 10.48° E and 76.41° & 76.56° N. CWS has an area extent of 85.067 km² and water spread area of 10.1 km². The sanctuary consists of more than 250 streams, which drains into the Chimmony Reservoir (George 2012; Velayudhan et al.



2021). In this study, 10 different streams were selected to study the algal flora (Table 1).

Sampling

The algal samples were collected from 10 different stations using forceps, scalpel, and blade. The collections were made from the surface level, the underside of rocks, mucilage masses attached to dripping rocks, and tree trunks. 4% formalin solution was used for preservation. The collected specimens were observed under a microscope by preparing wet mounts within 48 hours. The algal specimens were identified using standard literature, monographs and research papers (Ralfs 1848; Turner 1892; Desikachary 1959; Randhawa 1959; Prescott 1961; Pal & Kundu 1962; Ramanathan 1964; Philipose 1967; Hindak 1977; Hirose et al. 1977; Hindak 1984; West & West 1904; Kouwets & Coesel 1984; Prasad & Misra 1992; Wolowski 1998; Wotowski & Hindak 2005).

RESULTS

In the study conducted in CWS, 61 algal species were recorded, which belongs to four different classes (Chlorophyceae, Euglenineae, Rhodophyceae, and Cyanophyceae). These species are represented by 37 genera, 22 families, and 14 orders (Table 2). The class Chlorophyceae represents 33 taxa under 22 genera, the class Euglenineae represents seven taxa under four genera, the class Rhodophyceae represents one taxa under one genera, and the class Cyanophyceae represents 20 taxa under 10 genera.

Class: Chlorophyceae Order: Volvocales Family: Chlamydomonadaceae Genus: *Chlamydomonas* Ehrenberg

1. Chlamydomonas globosa Snow (Image 1) Prescott, 1961, p.71, pl.1, figs. 8,9

The cells are globose, enclosed in a hyaline, gelatinous sheath. The cell is $3-5 \mu m$ in diameter and $5-10 \mu m$ long. The cell consists of a parietal cup-shaped chloroplast with basal pyrenoid and a contractile vacuole at the anterior end. The cell is covered with a smooth membrane and two flagella at the anterior end. The pigment spot is small and inconspicuous.

Family: Volvocaceae Genus: Gonium Mueller

2. Gonium pectorale Mueller (Image 2)

Prescott, 1961, p. 75, pl.1, fig. 22

The colony consists of 16 ellipsoid to subspherical cells arranged in a flat quadrangular plate. This quadrangular plate consists of four inner cells covered by 12 marginal cells. The anterior ends of marginal cells were projecting outwards. Each cell is enclosed in an individual sheath and the cells are 5–20 μ m in diameter.

Order: Tetrasporales Family: Tetrasporaceae

Genus: Tetraspora Link

3. *Tetraspora gelatinosa* (Vauch.) Desvaux (Image 3) Prescott, 1961, p. 88, pl.5, figs. 3,4

The thallus is a macroscopic attached floating cylindrical sac where each cell are irregularly arranged. The thallus is globular and bullate, in which spherical cells are arranged in a tetrad manner. The thallus is covered in a thick mucilaginous sheath, and the cells are $6-10 \mu m$ in diameter.

Order: Chlorococcales Family: Chlorococcaceae Genus: *Chlorococcum* Fries

4. *Chlorococcum humicola* (Naeg.) Rabenhorst (Image 4,5)

Prescott, 1961, p. 212, pl.45, fig. 1

The colony is unicellular, non-motile, with spherical cells in various small clumps. Each cells consist of a completely filled spherical chloroplast with a single pyrenoid. The cell is $7-10 \mu m$ in diameter.

Family: Selenastraceae

Genus: *Monoraphidium* Komarkova - Legnerova 5. *Monoraphidium griffithii* (Berkeley) Komarekova -Legnerova (Image 6)

Table 1. Latitude and Longitude of sampling sites

	Sampling sites	Latitude (E) and Longitude (N)
1	Pookoyil thodu	10.4600, 76.4744
2	Kidakkapara thodu	10.4641, 76.4658
3	Viraku thodu	10.4497, 76.4444
4	Nellipara thodu	10.4458, 76.4638
5	Anaporu thodu	10.4300, 76.5069
6	Kodakallu thodu	10.4388, 76.5141
7	Odan thodu	10.4522, 76.5047
8	Mullapara thodu	10.4558, 76.4983
9	Payampara thodu	10.4544, 76.4913
10	Chimmony dam	10.4605, 76.4722

Table 2. Algal species identified from Chimmony Wildlife Sanctuary.

	Class	Order	Family	Genus	Species
1			Chlamydomonadaceae	Chlamydomonas	globosa Snow
2		Volvocales	Volvocaceae	Gonium	pectorale Mueller
3		Tetrasporales	Tetrasporaceae	Tetraspora	<i>gelatinosα</i> (Vauch.) Desvaux
4			Chlorococcaceae	Chlorococcum	humicola (Naeg.) Rabenhorst
5		Chlorococcales	Selenastraceae	Monoraphidium	<i>griffithii</i> (Berkeley) Komarekova - Legnerova
6					indicum Hindak
7			Scenedesmaceae	Scenedesmus	<i>quadricauda</i> var. <i>maximus</i> West & West
8		Ulotrichales	Ulothrichaceae	Ulothrix	aequalis Kuetzing
9		Cladophorales	Cladophoraceae	Pithophora	oedogonia (Mont.) Wittrock
10		Chaetophorales	Trentepohliaceae	Trentepohlia	aurea (L.) Martius
11		Oedogoniales	Orderenierer		areschougii Wittrock
12		Oedogoniales	Oedogoniaceae	Oedogonium	croasdaleae Jao
13				Mougeotia	scalaris Hassall
14				Zygnema	<i>carinatum</i> Taft
15					acanthophora (Skuja) Czurda
16					condensata (Vauch.) Kuetzing
17	Chlorophyceae		Zygnemataceae		decimina (Mueller) Kuetzing
18	Chiorophyceae			Spirogyra	<i>fuellebornei</i> Schmidle
19					micropunctata Transeau
20					novaeangliae Transeau
21		Zygnematales			rhizobrachialis Jao
22				Cylindrocystis	brebissonii (Ralfs) De Bary
23			Mesotaeniaceae	Netrium	digitus (Ehrbg.) Itzigs. & Rothe
24			Desmediaceae	Actinotaenium	silvae-nigrae (Rabanus) Kouwets & Coesel
25				Closterium	ehrenbergii meneghinii var. ehrenbergii
26					moniliferum Ehrenberg ex Ralfs
27					tumidulum Gay
28				Cosmarium	botrytis Meneg
29					subtumidum Nordst
30				Micrasterias	radians Turn var. bogoriensis (Breb) G.S West
31				Pleurotaenium	trabecula (Ehrbg) Nag
32				Staurastrum	zonatum Borges var. majus Presc.
33		Charales	Characeae	Nitella	<i>furcata</i> (Roxburgh <i>apud</i> Bruzelius) Agardh
34		Euglenales	Astasiaceae	Euglena	elastica Prescott
35	Euglenineae				minuta Prescott
36				Phacus	<i>curvicauda</i> Swirenko
37					obolus Pochmann
38					orbicularis var. caudatus Skzortzow
39				Lepocinclis	acus (Muller) marin and Melkonian
40				Trachelomonas	hispida var. papillata Skvortzow
41	Rhodophyceae	Batrachospermales	Batrachospermaceae	Sheathia	boryana (Sirodot) Salomaki & M.L.Vis

	Class	Order	Family	Genus	Species
42		Chroococcales	Chroococcaceae	Aphanocapsa	<i>pulchra</i> (Kutz) Rabenh
43				Microcystis	<i>aeruginosa</i> Kutz.
44			Microchaetaceae	Microchaete	uberrima Carter, N
45					<i>limosa</i> Agardh ex Gomont
46				Oscillatoria	subbrevis Schmidle
47		Nostocales	Oscillatoriaceae		vizagapattensis Rao, C. B
48				Phormidium	abronema Skuja
49	Cyanophyceae				hansgirgi Schmidle
50					microtomum Skuja
51					molle (Kutz.) Gomont
52					<i>retzii</i> (Ag.) Gomont
53					truncicola Ghose
54					<i>usterii</i> Schmidle
55			Nostocaceae	Anabaena	anomala Fritsch
56					sphaerica Bornet et Flahault
57				Cylindrospermum	stagnale (Kutz.) Born.et Flah
58			Rivulariaceae	Gloeotrichia	echinulata (J. E. Smith) P. Richter
59				Scytonema	ocellatum Lyngbye ex Born. et Flah
60					<i>rivulare</i> Borzi ex Born. et Flah
61		Stigonematales	Nostochopsidaceae	Nostochopsis	lobatus Wood em. Geitler

Hindak, 1984, p. 219, pl. 79, figs. 5,8

The cell is straight and fusiform, having a tapering from the centre towards the pointed ends. The cell is $45-50 \ \mu m \log and 2-3 \ \mu m broad$.

Monoraphidium indicum Hindak (Image 7) Hindak, 1977, p.105, pl.44

The cells are very thin and are accurately curved. The cell has a tapering towards the end and it is pointed. The cell is 40–45 μ m long and 1.5–2 μ m broad.

Family: Scenedesmaceae Genus: Scenedesmus Meyen

7. Scenedesmus quadricauda var. maximus West & West (Image 8)

M. T Philipose, 1967, p. 283, fig. 187 g

The colonies are usually four-celled with much larger cells. The cell is 25–30 μ m long and 10–11 μ m in diameter. The spines are 25–35 μ m long.

Order: Ulotrichales Family: Ulothrichaceae

Genus: Ulothrix Kuetzing

Ulothrix aequalis Kuetzing (Image 9)
 K.R Ramanathan, 1964, p.36, pl.9 I-L
 The thallus is non-branching, filamentous with

cylindrical cells. The cells are 12–14 μ m broad and 24–28 μ m long. The cells consist of a striated cell wall, girdle shaped broad chloroplast covering half of the wall surface with one or more pyrenoids.

Order: Cladophorales

Family: Cladophoraceae

Genus: Pithophora Wittrock

9. *Pithophora oedogonia* (Mont.) Wittrock (Image 10, 11; Image 12, 13)

Prescott, 1961, p.140, pl.22, figs. 7-10

The filaments are slender 50–60 μ m in diameter with solitary branching. Each cell are cylindrical and long. The akinetes are cylindrical and slightly swollen and acuminate at the terminal. Akinetes are 55–140 μ m in diameter and 90–350 μ m long.

Order: Chaetophorales

Family: Trentepohliaceae

Genus: Trentepohlia Martius

10. *Trentepohlia aurea* **(L.)** Martius (Image 14,15,16) Prescott, 1961, p.133, pl.67, figs. 6–9

The cells are rusty-brown in colour sometimes the thallus shows yellow colour in shaded regions. The cells are slightly swollen but slightly reduced in diameter towards apices. The cell has a smooth wall, and it is 4–10

Jose & Xavier 🖉 🚮

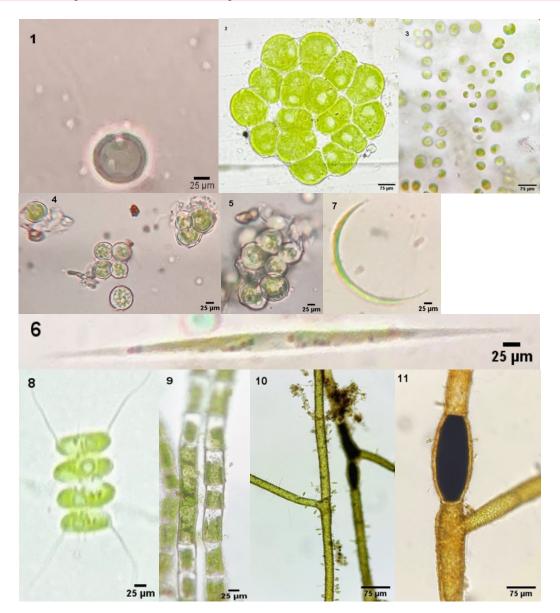


Image 1–11. 1—Chlamydomonas globosa | 2—Gonium pectorale | 3—Tetraspora gelatinosa | 4, 5—Chlorococcum humicola | 6— Monoraphidium griffithii | 7—Monoraphidium indicum | 8—Scenedesmus quadricauda var. maximus | 9—Ulothrix aequalis | 10, 11— Pithophora oedogonia. (© Joel Jose)

 μ m in diameter. The sporangia are generally terminal on curved cells with 15–20 μ m in diameter. The gametangia are not frequently observed, and they will be the same size as the sporangia.

Order: Oedogoniales Family: Oedogoniaceae Genus: *Oedogonium* Link

11. Oedogonium areschougii Wittrock (Image 17)

Prescott, 1961, p. 204

The filament is nannandrous & gynandrosporous. The filaments are cylindrical in shape with a 10–12 μm

diameter and 35–28 μ m long. The oogonia is pyriform globose shaped and operculate with 30-35 μ m diameter and 36–40 μ m long. The smooth-walled oospore is not completely filled inside the oogonia. The diameter of the oospore is 23–25 μ m. The dwarf males are unicellular attached near or on the oogonia with 6–7 μ m diameter and 13–15 μ m long.

12. Oedogonium croasdaleae Jao (Image 18, 19)

Prescott, 1961, p.204, pl.41, fig. 11

The filament is nannandrous and gynandrosporous. The vegetative cells are cylindrical 25–30 μm in diameter

and 150–200 μ m long. The oogonia are two in a series, 60–70 μ m in diameter and 80–113 μ m long. The dwarf males are 9–17 μ m in diameter and 48–55 μ m long.

Order: Zygnematales Family: Zygnemataceae Genus: *Mougeotia* C.A. Agardh

13. Mougeotia scalaris Hassall (Image 20)

Prescott, 1961, p. 304, pl.71, figs. 6,7

The filaments are $14-20 \ \mu\text{m}$ in diameter and $34-182 \ \mu\text{m}$ long. The chloroplast consists of 4-6 pyrenoids. The zygospores are globose to ovate with smooth walls and formed in the tube due to scalariform conjugation. The zygospore measures up to $30-35 \ \mu\text{m}$ in length and $26-30 \ \mu\text{m}$ in diameter.

Genus: Zygnema Agardh

14. Zygnema carinatum Taft (Image 21)

Randhawa 1959, p.225, fig. 160

The filaments are greenish and unbranched. The cells are rectangular to square in shape. Presence of two star-shaped chloroplasts. The cell is 11–15 μ m long and 10 μ m broad. The scalariform conjugation results in the formation of globose shaped zygospore in the tube. The globose zygospore is formed at the right angle of the tube, and it measures 13–16 μ m in length and 15–20 μ m in breadth.

Genus: Spirogyra Link

15. Spirogyra acanthophora (Skuja) Czurda (Image 22) Randhawa, 1959, p.376, fig. 413

The filaments are 300–328 μ m long and 60–65 μ m wide. The zygospores are 37–42 μ m in diameter and 50–62 μ m in length.

16. Spirogyra condensata (Vauch.) Kuetzing (Image 3: 23)

Prescott, 1961, p. 312, pl.72, figs. 5,6

The filaments are 111–153 μ m long and 40–53 μ m wide. Smooth walled zygospores were formed due to conjugation, and it measures up to 35–37 μ m in diameter and 52–60 μ m in length.

17. Spirogyra decimina (Mueller) Kuetzing (Image 24, 25)

Prescott, 1961, p. 313

The filaments are 130–133 μ m long and 20–24 μ m wide. Presence of two chloroplasts. The zygospores are cylindrical to ovate with a smooth wall that measures up to 32–38 μ m in diameter to 30–35 μ m in length.

Spirogyra fuellebornei Schmidle (Image 26) Randhawa. 1959. P. 316, fig. 291

The filaments are long and cylindrical having 238– 376 μ m long and 26–31 μ m broad. Presence of two chloroplast, having 3–4 turns in a cell. The zygospores are 30–39 μ m in diameter and 58–65 μ m in length.

19. Spirogyra micropunctata Transeau (Image 27)

Prescott, 1961, p. 317, pl.73, fig. 9

The filaments are 243–300 μ m long and 29–35 μ m wide. The scalariform conjugation produces an ellipsoidal zygospore, which measures up to 35–40 μ m in diameter and 60–72 μ m long.

20. Spirogyra novaeangliae Transeau (Image 28)

Prescott, 1961, p. 318, pl.75, figs. 1-3

The filaments are 200–230 μ m long and 58 μ m wide. The zygospore is ovate to ellipsoidal. The zygospore exhibits a brown colour which measures up to 50–60 μ m in diameter and 85–90 μ m in length.

21. *Spirogyra rhizobrachialis* Jao (Image 3: 29) Prescott, 1961, p. 320, pl.76, figs. 1, 2

The filaments are 43–50 μ m in diameter and 120–211 μ m long. Presence of two crenate and deeply toothed chloroplast. The fertile cylindrical cells form zygospores through conjugation. The zygospore is ellipsoidal brown, which measures up to 40–50 μ m in diameter and 111 μ m in length.

Family: Mesotaeniaceae

Genus: Cylindrocystis De Bary

22. Cylindrocystis brebissonii (Ralfs) De Bary (Image 30, 31)

W. West & G.S. West, 1904, pl. 4, figs. 23–32, pl.5, fig. 10

The cells are cylindrical with round apices. The chloroplast consists of a few large radiating prolongations. The cell body is $35-40 \ \mu m$ long and $22-28 \ \mu m$ in broad.

Genus: *Netrium* (Nageli) Itzigsohn & Rothe in Rabenhorst

23. *Netrium digitus* (Ehrbg.) Itzigs. & Rothe (Image 32) W. West & G. S. West, 1904, pl. 6, fig. 14–16

The cell is generally large and elliptic to oblong in shape. The cell is gradually attenuated from the centre towards the apices, which is rounded and truncated. The chloroplast is axile with deeply notched free margins. The cell body is 150–160 μ m long and 40–45 μ m in diameter.

Jose & Xavier

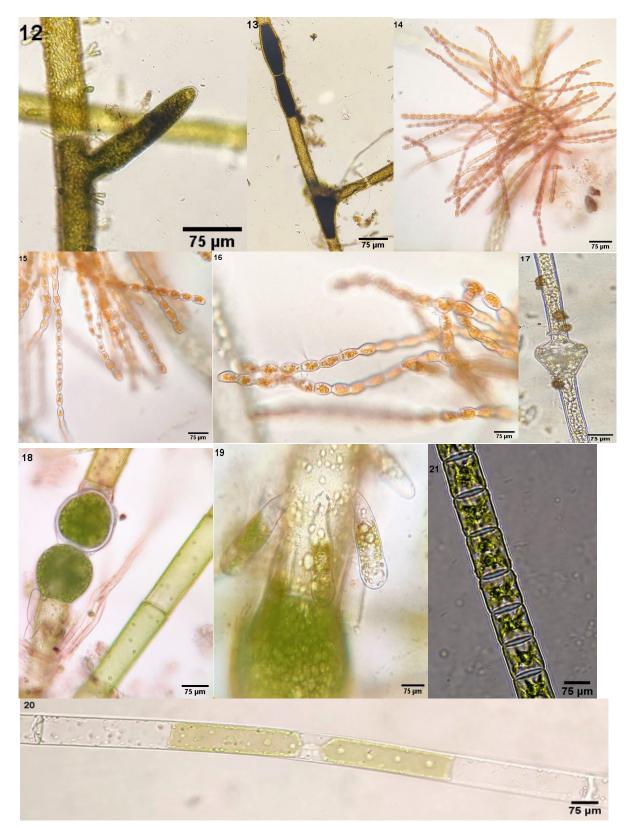


Image 12–21. 12, 13–Pithophora oedogonia | 14, 15, 16–Trentepohlia aurea | 17–Oedogonium areschougii | 18, 19–Oedogonium croasdaleae | 20–Mougeotia scalaris | 21–Zygnema carinatum. (© Joel Jose)



Image 22–29. 22–*Spirogyra acanthophora* | 23–*Spirogyra condensata* | 24, 25–*Spirogyra decimina* | 26–*Spirogyra fuellebornei* | 27– Spirogyra micropunctata | 29–*Spirogyra rhizobrachialis*. (© Joel Jose)

Family: Desmediaceae

Genus: Actinotaenium (Nageli) Teiling 24. Actinotaenium silvae-nigrae (Rabanus) Kouwets &

Coesel (Image 33)

Kouwets & Coesel, 1984, p. 555-562, fig. 23

The cell is cylindrical with broadly rounded ends with a smooth cell wall. The cell is 60–65 μm long and 20–25 μm wide.

Genus: Closterium Nitzsch ex Ralfs

25. *Closterium ehrenbergii Meneghinii* var. *Ehrenbergii* (Image 34)

Hirose, H, et al., 1977

The cell body is large and bulged at the centre with a smooth cell wall. The chloroplasts consist of 4–7 laminae with many scattered pyrenoids. The cell body is 250–890 μ m long and 50–165 μ m wide.

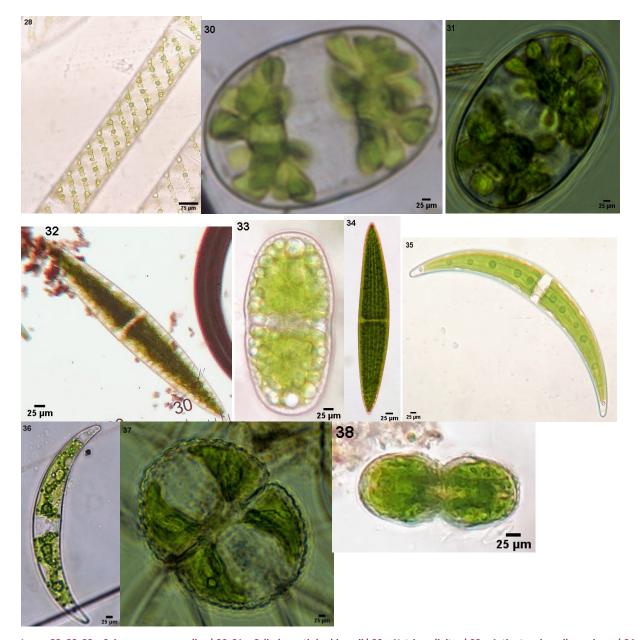


Image 28–38. 28–Spirogyra novaeangliae | 30, 31–Cylindrocystis brebissonii | 32–Netrium digitus | 33–Actinotaenium silvae-nigrae | 34– Closterium ehrenbergii | 35–Closterium moniliferum | 36–Closterium tumidulum | 37–Cosmarium botrytis | 38–Cosmarium subtumidum. (© Joel Jose)

26. *Closterium moniliferum* Ehrenberg ex Ralfs (Image 35)

Prasad & Misra, 1992, p. 113, pl. 12, fig. 4.

The cell is curved with rounded apices. The chloroplast consists of 7–10 pyrenoids arranged in a median series. The cell is 140–155 μ m long and 7–20 μ m broad.

27. Closterium tumidulum Gay (Image 36)

Turner, 1892, p.19, pl.1, fig. 20

The cell is small and curved with an acute tip. The cell

is 90–100 μm long and 10–15 μm broad.

Genus: Cosmarium Ralfs

28. Cosmarium botrytis Meneg (Image 37)

Ralfs, 1848, p.99, pl. 16, fig. 1

The cell has denticulate margins with a deeply constricted linear notch at the centre. The cell is 54.1–77.6 μ m long and 40.6–60.6 μ m broad.

29. *Cosmarium subtumidum* Nordst (Image 38) Prescott, 1961, p. 70, pl. 29, figs. 12, 13

Jose & Xavier

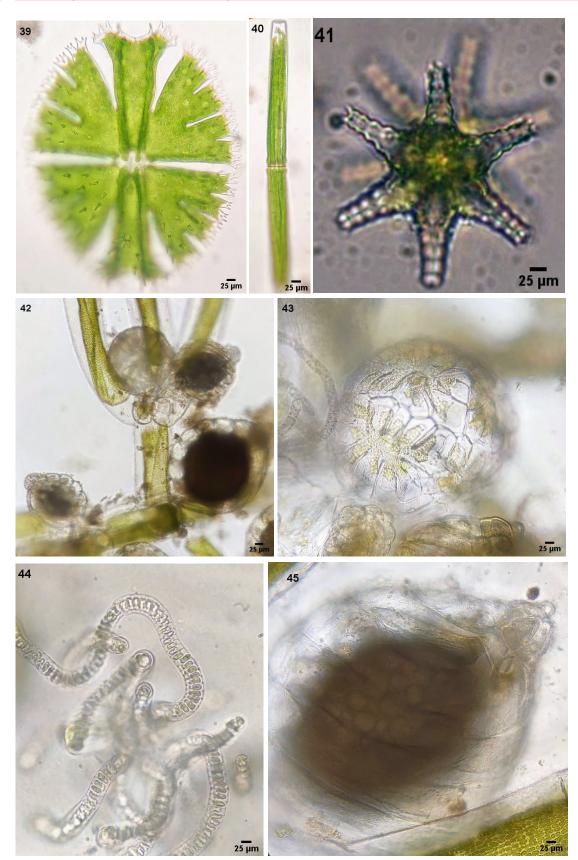


Image 39–45. 39–*Micrasterias radians* var. *bogoriensis* | 40–*Pleurotaenium trabecula* | 41–*Staurastrum zonatum* var. *majus* | 42–45– *Nitella furcata*. (© Joel Jose)

The cell body is 30–43 μ m long, 14–19 μ m wide and isthmus is 12–14 μ m.

Genus: Micrasterias C. Agardh

30. *Micrasterias radians* Turn var. *bogoriensis* (Breb) G.S. West (Image 39)

Prescott, 1961, p.51, pl.23, figs. 2, 3

The cell body is 121–206 μm long, 126–170 μm wide and the isthmus is 14–17 μm wide

Genus: Pleurotaenium Nageli

31. *Pleurotaenium trabecula* (Ehrbg) Nag (Image 40) Prescott, 1961, p. 18, pl. 3, fig. 4

The cylindrical cell body is 400–434 μ m long and 30– 40 μ m in diameter. The cell is constricted at the centre, with a slight bulge at the base semi cell. The chloroplast is elongated with 3–4 laminae.

Genus: Staurastrum (Meyen) Ralphs

32. Staurastrum zonatum Borges var. majus Presc. (Image 41)

Prescott, 1961, p.119, pl. 46, fig. 8

The semi cells consist of five long dentate ends with rings of granules and the apex biundulate with some tiny teeth. The cell body is 40–70 μ m long, 81–90 μ m wide and the isthmus is 13–16 μ m.

Order: Charales

Family: Characeae

Genus: Nitella C. Agardh

33. *Nitella furcata* (Roxburgh apud Bruzelius) Agardh (Image 42–45)

B.P. Pal et al., 1962, p.62, figs. 76-79

The plant is monoecious. The stem is $600-1,000 \ \mu m$ thick and antheridia is terminal, which is $200-250 \ \mu m$ in diameter. The oogonia are 1-2, together, which are $230-240 \ \mu m$ long and $210-310 \ \mu m$ in diameter. Spiral cells showing 7–8 convolutions and the coronula are 70–100 $\ \mu m$ high and 70 $\ \mu m$ at the base.

Class: Euglenineae Order: Euglenales Family: Astasiaceae Genus: Euglena Ehrenberg 34. Euglena elastica Prescott (Image 46)

Prescott, 1962, p. 392, pl.86, figs. 10–12

The cells have the potential to change shape regularly, when in motion. Usually the cells are spindle-shaped but often swollen in the mid-region and slightly tapered to the apices. The cell consists of many irregularly ovoid-shaped chloroplasts. The cell is 10–11 µm in diameter

and 80–90 μm long.

35. Euglena minuta Prescott (Image 47)

Prescott, 1962, p. 393, pl.85, figs. 23, 25

The cells are highly active, which are fusiform to pyriform in shape. The smooth membraned cell consists of one plate-like chloroplast with a pyrenoid. The cell is $14-16 \mu m$ long and $2-6 \mu m$ broad.

Genus: Phacus Dujardin

36. Phacus curvicauda Swirenko (Image 48)

Prescott, 1962, p.399, pl.87, fig. 14, pl.88, fig.21

The cells are ovoid and slightly spiral, which causes the caudus to curve slightly to the left. The cell consists of numerous ovoid chloroplasts. The cell is 40–48 μm in diameter and 48–60 μm long.

37. Phacus obolus Pochmann (Image 49)

Wolowski, 1998, p.78, figs. 270-272

The cells are broadly oval and slightly narrower at the anterior end with straight, conical cauda at the posterior end. The cell consists of numerous ovoid-globular chloroplasts. Cells are 34–42 μ m long and 22–35 μ m broad

38. *Phacus orbicularis* var. *caudatus* Skzortzow (Image 50)

Prescott, 1962, p.401, pl.87, fig. 12, pl.88, fig.15

Cells are ovoid with a long, straight , sharply pointed caudus. 1–2 paramylon bodies are present. Cells are 38–41 μ m in diameter and 50–70 μ m long.

Genus: Lepocinclis Perty

39. *Lepocinclis acus* (Muller) Marin & Melkonian (Image 51)

Wotowski & Hindak, 2005, p. 28, figs. 5–8 The cells are long, elongate, thin and spindle-shaped, gradually tapering to apices which forms a sharp tail. Numerous disc-shaped chloroplasts are present, and two paramylon bodies are present. The cells are 10–12 μ m diameter and 150 μ m long.

Genus: Trachelomonas Ehrenberg

40. *Trachelomonas hispida* var. *papillata* Skvortzow (Image 52)

Prescott, 1962, p. 414, pl. 84, fig. 7

The cell is $25-30 \ \mu m$ in diameter and $35-40 \ \mu m$ long. The wall is brown smooth except for a few minute spines near the flagellum aperture.

Jose & Xavier

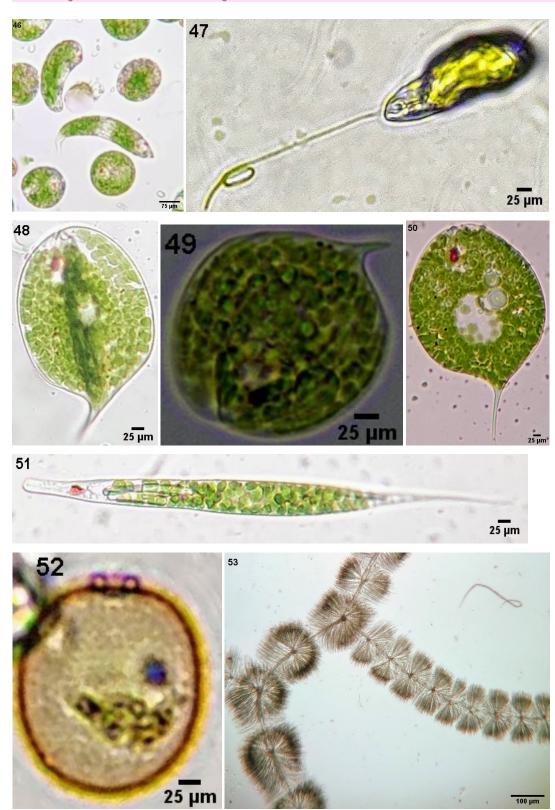


Image 46–53. 46—Euglena elastica | 47—Euglena minuta | 48—Phacus curvicauda | 49—Phacus obolus | 50—Phacus orbicularis var. caudatus | 51—Lepocinclis acus | 52—Trachelomonas hispida var. papillata | 53—Sheathia boryana. (© Joel Jose).

Class: Rhodophyceae Order: Batrachospermales Family: Batrachospermaceae Genus: *Batrachospermum* Roth

41. *Sheathia boryana* (Sirodot) Salomaki & M.L.Vis (Image 53; Image 54)

Prescott, 1962, p.567, pl. 136, fig. 4; Sheath & Hymes, 1980, p.1306, figs. 31–36; John & Francis, 2013, p. 237.

The plant is 5–9 cm high and has a highly mucilaginous thallus, which is brown to green. The central axes are 90–105 μ m wide, and glomeruli are ellipsoidal to globular. The lateral branches have short internodes. The carpogonia are 4–5 μ m wide at the basal portion and 25–30 μ m long. The trichogyne are elongate, club-shaped and embrace the carpogonia. The carposporophyte is globular and scattered close to the periphery. The carposporophyte measures 14–150 μ m in diameter.

Class: Cyanophyceae Order: Chroococcales

Family: Chroococcaceae

Genus: Aphanocapsa Nag

42. Aphanocapsa pulchra (Kutz) Rabenh (Image 55)

T.V. Desikachary, 1959, p.132, pl. 21, fig. 2

The thallus is gelatinous and blue-green. The cells are spherical, loosely arranged in single or sometimes doubles with individual sheaths. The cells are $6-7 \mu m$ in diameter.

Genus: Microcystis Kutzing

43. Microcystis aeruginosa Kutz. (Image 56)

T.V Desikachary, 1959, p. 93, pl. 17, fig. 1, 2, 6

The colonies are free-floating and attaining a macroscopic size with a mucilaginous envelope. The cells in the colony are spherical with distinct hyaline colonial mucilage. The colonies are light brown and round with $5-7 \mu m$ in diameter. Gas vacuoles are present.

Order: Nostocales

Family: Microchaetaceae Genus: *Microchaete* Thuret

44. Microchaete uberrima Carter, N (Image 57, 58)

T.V. Desikachary, 1959, p.511, pl. 104, figs. 5-7, 10, 13, 16, 18

The trichomes were long up to 4 mm, with cylindrical cells having a firm sheath. The filaments were $10-15 \mu m$ broad with intercalary heterocyst.

Family: Oscillatoriaceae

Genus: Oscillatoria Vaucher

45. *Oscillatoria limosa* Agardh ex Gomont (Image 59) T.V. Desikachary, 1959, p.206, pl. 42, fig.11

The thallus is blue-green with a straight trichome that is slightly constricted. The trichomes are 12–13 μm broad and 2–4 μm long.

46. Oscillatoria subbrevis Schmidle (Image 60, 61)

T. V. Desikachary, 1959, p.207, pl. 37, fig. 2, pl. 40, fig. 1

The trichomes are single, straight and not attenuated with round cell, calyptra absent. The trichome is 5–6 μ m broad, and the cells are 3–4 μ m long. The trichomes are blue-green, and they exhibit an oscillating movement at the apex.

47. Oscillatoria vizagapattensis Rao, C.B. (Image 62)

T.V. Desikachary, 1959, p.205, pl. 39, figs. 16, 18.

The cells are much shorter than the broad and form a broadly rounded cap with a slightly thickened outer wall. The trichome is blue-green in colour and 8–10 μm broad.

Genus: Phormidium Kutz.

48. Phormidium abronema Skuja (Image 64)

T.V. Desikachary, 1959, p.257.

The thallus is blackish-green to light bluish. The trichomes consist of the hyaline mucilaginous sheath. The cells are cylindrical or barrel-shaped. The trichome is $3-4 \mu m$ broad and $16-17 \mu m$ long.

49. *Phormidium hansgirgi* Schmidle (Image 63; Image 65)

T.V. Desikachary, 1959, p.272, pl. 43, fig. 20

The filaments are straight with a very thin mucilaginous sheath. The trichomes are cylindrical and not capitate. The hormogones are short. The trichomes are $12-14 \mu m$ broad and $2-3 \mu m$ long.

50. Phormidium microtomum Skuja (Image 66)

T.V. Desikachary, 1959, p.257, pl. 43, fig. 16, 17

The trichomes are greyish-brown, straight with a thin colourless sheath. The ends of trichomes are attenuated, and cells are well constricted at the cross wall. The trichome is $6-8 \ \mu m$ broad with apical rounded hyaline calyptra.

51. *Phormidium molle* (Kutz.) Gomont (Image 67)

T.V. Desikachary, 1959, p.255, pl. 59, fig. 8

The trichomes are thin, straight, constricted at

52. Phormidium retzii (Ag.) Gomont (Image 68)

T.V. Desikachary, 1959, p.268, pl. 44, figs. 13-15

The filaments are straight with a thin mucilaginous sheath. The trichomes are blue-green with a thin sheath. The ends are not attenuated and not capitate. The trichomes are $11-13 \mu m$ broad and $8-10 \mu m$ long.

53. Phormidium truncicola Ghose (Image 70)

T.V. Desikachary, 1959, p.258, pl. 59, fig. 9

The trichomes consist of thin membrane and are constricted at cross walls. The calyptra is absent. The trichomes are 6–8 μ m broad and 2–3 μ m long.

54. Phormidium usterii Schmidle (Image 69)

T.V. Desikachary, 1959, p.257.

Trichomes with thin mucilaginous sheath. The cells are shorter than broad with short rectangular cells with broadly round ends. The trichome is $3-4 \mu m$ broad and $5-6 \mu m$ long.

Family: Nostocaceae

Genus: Anabaena Bory de Bornet & Flahault

55. Anabaena anomala Fritsch (Image 71)

T.V. Desikachary, 1959, p.398, pl. 73, fig. 2

The thallus is thin and gelatinous. The cells are spherical, and apical cells are rounded. The trichome is blue-green, consisting of densely or irregularly aggregated rounded cells. The cell is $2-5 \mu m$ in diameter.

Anabaena sphaerica Bornet et Flahault (Image 72) T.V. Desikachary, 1959, p.393, pl. 71, fig. 8

Gelatinous thin sheath present, Trichomes are pale blue-green in colour. Cells are barrel-shaped and 2–7 μ m long. End cells are rounded. Heterocysts are 9–11 μ m broad and 13–17 μ m long with a smooth yellow outer wall.

Genus: Cylindrospermum Kutz

21260

57. *Cylindrospermum stagnale* (Kutz.) Born.et Flah (Image 73, 74)

T.V. Desikachary, 1959, p.363, pl. 65, fig. 9

The thallus is blue-green with a mucilaginous sheath. The cells are constricted at the cross wall and nearly quadrant to cylindrical with spherical or oblong heterocyst. The trichomes are cylindrical and $2-5 \ \mu m$ broad.

Family: Rivulariaceae

Genus: Gloeotrichia Ag.

58. *Gloeotrichia echinulata* (J. E. Smith) P. Richter (Image 75, 76)

Prescott, 1962, p.557, pl. 134, figs. 1,2

The colonies are tiny macroscopic and opaque at the centre and translucent at the periphery. The colonies are free-floating, spherical and covered in a gelatinous sheath. The trichomes radiate from a common centre and are tapered from basal heterocyst to a fine hair-like end. The cells are cylindrical to barrel-shaped 6–9 μ m wide, and the cells are joined end to end to form long chains.

Genus: Scytonema Ag.

59. Scytonema ocellatum Lyngbye ex Born. et Flah (Image 77)

T.V Desikachary, 1959, p.467, pl.92, fig.3

The thallus is cushion-like, brownish to reddish with false branching. The trichomes are covered in a firm mucilaginous sheath. The filaments are 11–15 μ m broad.

60. Scytonema rivulare Borzi ex Born. et Flah (Image 78)

T.V Desikachary, 1959, p.452, pl.100, fig.2

The thallus is broad, with a thick mucilaginous sheath. The thallus is brownish to reddish with false branching. The cells are shorter than broad and $30 \,\mu\text{m}$ broad.

Order: Stigonematales

Family: Nostochopsidaceae

Genus: Nostochopsis Wood em. Geitler

61. *Nostochopsis lobatus Wood em. Geitler* (Image 79; Image 80, 81)

T.V. Desikachary, 1959, p.570, pl. 120, figs. 1-8

The thallus is irregularly lobed, blue-green with a thick mucilaginous matrix. The cells are barrel-shaped. The heterocyst are mostly lateral, spherical to ellipsoidal. The trichomes are 5–9 μ m wide and 6–10 μ m long.

DISCUSSION

The freshwater ecosystem holds the most biodiversity among all other ecosystem. The study of freshwater habitat is significant as it occupies only 0.5% of the earth surface, but is equally crucial because they are the cheapest natural source for domestic and industrial purposes (Norton et al. 1996).

The present study portraits the algal diversity of CWS. In our study, Chlorophyceae and Cyanophyceae

Jose & Xavier

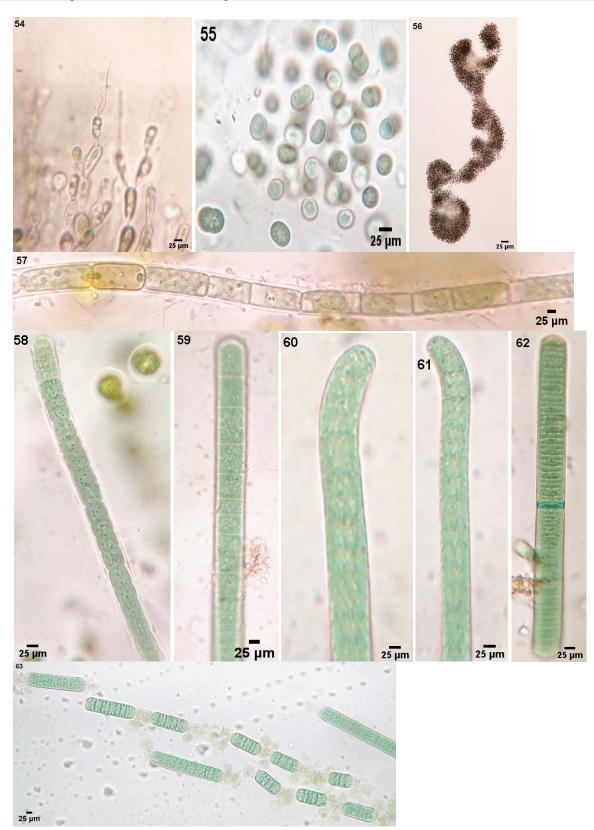


 Image 54–63. 54—Sheathia boryana | 55—Aphanocapsa pulchra | 56—Microcystis aeruginosa | 57, 58—Microchaete uberrima | 59—

 Oscillatoria limosa | 60, 61—Oscillatoria subbrevis | 62—Oscillatoria vizagapattensis | 63—Phormidium hansgirgi. (© Joel Jose)

Jose & Xavier

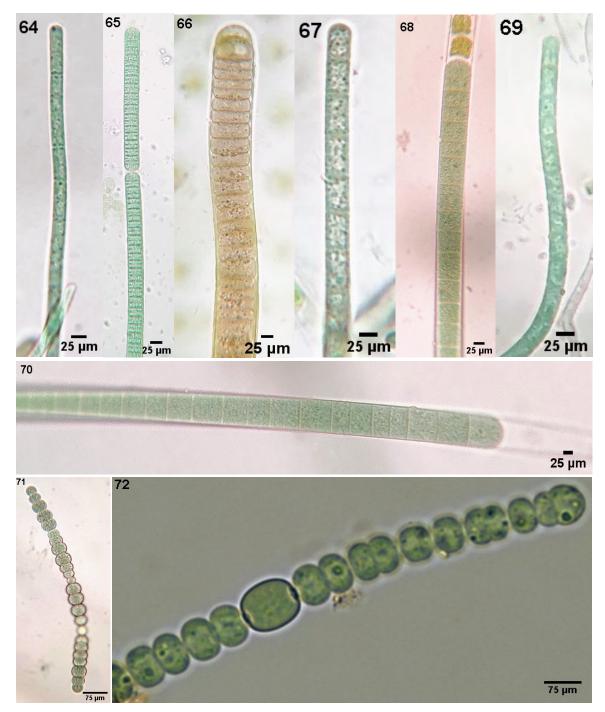


Image 64–72. 64–Phormidium abronema | 65–Phormidium hansgirgi | 66–Phormidium microtomum | 67–Phormidium molle | 68–Phormidium retzii | 69–Phormidium usterii | 70–Phormidium truncicola | 71–Anabaena anomala | 72–Anabaena sphaerica. (© Joel Jose)

algae were dominant. The preliminary study conducted in Kannam River, Kannur, Kerala for the diversity of algae has reported 40 algal species of which Chlorophyceae was dominant, followed by Cyanophycea (Girish et al. 2018). The algal population of Pennar River, Kottayam, has reported 61 algal species were Chlorophyceae was dominant (Joseph & Claramma 2010). In our study also, more algae were reported from the order Zygnematales, and *Spirogyra* was the most common genus. The algal species from order Nostocales of Cyanophyceae was dominant. A similar type of diversity was observed in the Gundur lake of Tamil Nadu. Out of 87 algal species reported from Gundur Lake, 37 species were Cyanophyta (Vijayan et al. 2014). The algae from Chlorophyceae and

Jose & Xavier

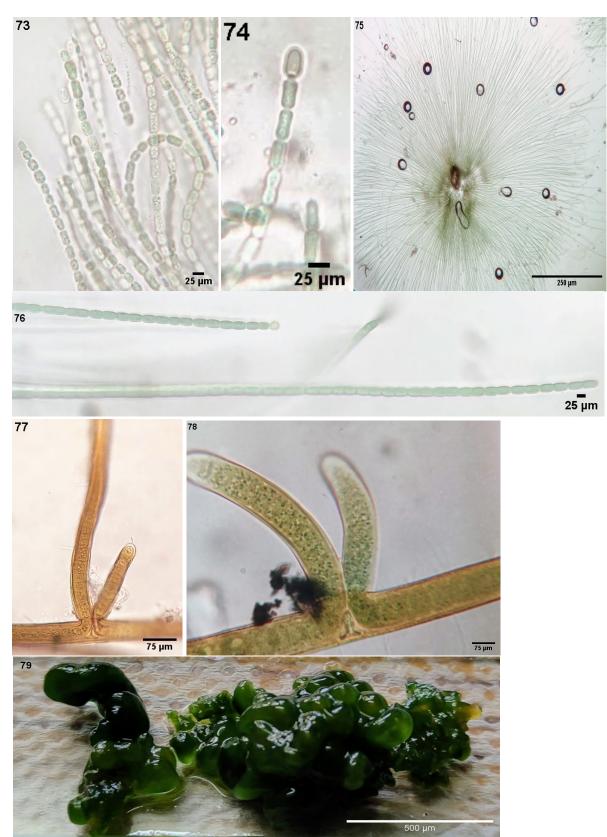


Image 73–79. 73, 74–Cylindrospermum stagnale | 75, 76–Gloeotrichia echinulata | 77–Scytonema ocellatum | 78–Scytonema rivulare | 79–Nostochopsis lobatus. (© Joel Jose)

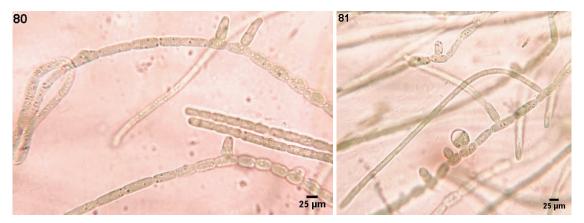


Image 80-81. 80, 81-Nostochopsis lobatus. (© Joel Jose)

Cyanophyceae were dominant in species composition compared to other classes.

CONCLUSION

Overall, the biodiversity study conducted in Chimmony Wildlife Sanctuary shows a good presence of algae. The study also revealed that *Spirogyra* was dominant from Chlorophyceae, *Phacus* was dominant from Euglenineae, and *Phormidium* was dominant from Cyanophyceae. The algal diversity directly depends on season and the physicochemical parameters of the freshwater ecosystem. Therefore, extensive seasonal studies are required for acquiring more knowledge about algal diversity.

REFERENCES

- Briji, G. (2005). Relevance and policy dimensions of research on biodiversity in freshwater ecosystems: a developing country perspective. *Hydrobiologia* 542(2): 19–21. https://doi.org/10.1007/ s10750-005-0890-1
- De Clerck, O., M.D. Guiry, F. Leliaert, Y. Samyn & H. Verbruggen. (2013). Algal Taxonomy: A Road to Nowhere? *Journal of Phycology* 49(2): 215–225. https://doi.org/10.1111/jpy.12020
- Desikachary, T.V. (1959). Cyanophyta. Indian Council of Agricultural Research, New Delhi, 686 pp.
- Girish, K.E., K.V. Thanzeeha, K. Sasikala, G. Pradeepkumar, K.K. Sivadasan & J.V. Abdul (2018). A preliminary study on the diversity of planktonic algae of Kaanam river, Kannur, Kerala, India. *Phykos* 48(2): 13–16.
- George, V.J. (2012). Management Plan Chimmony Wildlife Sanctuary. Kerala Forest Department, 107 pp.
- Hindak, F. (1977). Studies on the Chlorococcal Algae (Chlorophyceae)
 I. Veda Publishing house of the Slovak Academy of Sciences, Bratislava, 190 pp.
- Hindak, F. (1984). Studies on the Chlorococcal Algae (Chlorophyceae)
 III. Veda Publishing house of the Slovak Academy of Sciences, Bratislava, 299 pp.

- Hirose, H., K. Akiyama, K. Ioriya, H. Imahori, S. Kasaki, H. Kumano, Kobayashi, E. Takahashi, K. Tsumura, M. Hirano & T. Yamagishi (1977). Illustration of the Japanese freshwater algae. Uohida Rokakauho, Tokyo, Japan, 933 pp.
- John, J. & M.S. Francis (2013). An Illustrated algal flora of Kerala, vol. I – Idukki district. Green Carmel Scientific Books, Cochin, 279 pp.
- Joseph, P.V & J. Claramma (2010). Bacterial, fungal and algal population of Pennar river: a fresh water wet land in Kottayam district, Kerala. *Asian Journal of Chemistry* 22(6): 4286–4290.
- Koen, M. & H. Segers (2005). Taxonomy and systematics in biodiversity research. *Hydrobiologia* 542 (4):27–31. https://doi.org/10.1007/ s10750-005-0892-z
- Kouwets, F.A.C. & P.F.M. Coesel (1984). Taxonomic revision of the Conjugatophycean family Peniaceae on the basis of cell wall ultrastructure. *Journal of Phycology* 20(4): 555–562. https://doi. org/10.1111/j.0022-3646.1984.00555.x
- Norton, T.A., M. Melkonian & R.A. Andersen (1996). Algal biodiversity. *Phycologia* 35(4): 308–326. https://doi.org/10.2216/i0031-8884-35-4-308.1
- Pal, B.P. & B.C. Kundu (1962). Charophyta. Indian Council of Agricultural Research, New Delhi, 130 pp.
- Pandey, A.K. (1995). Conservation of biodiversity: Present status and future strategy, pp. 44–51. In: Pandey, A.K. (eds.). *Taxonomy and Biodiversity*. CBS Publishers and distributors, New Delhi, 232 pp.
- Philipose, M.T. (1967). Chlorococcales. Indian Council of Agricultural Research, New Delhi, 365 pp.
- Prasad, B.N. & P.K. Misra (1992). Fresh water algal flora of Andaman & Nicobar Islands. Vol. II. Bishen Singh Mahindhra Pal Singh, Dehra Dun, 284 pp.
- Prescott, G.W. (1961). Algae of the Western Great Lakes Area. Cranbrook Institute of Science, New York, 977 pp.
- Rai, L.C., D.K Har, H.M. Frieder & J.S. Carl (2000). Services of algae to the environment. *Journal of Microbiology and Biotechnology* 10(1): 36–119.
- Ralfs, J. (1848). The British Desmidieae. The drawings by Edward Jenner. Benham & Reeve, London, 266 pp.
- Ramanathan, K.R. (1964). Ulotrichales. Indian Council of Agricultural Research, New Delhi, 188 pp.
- Randhawa, M.S. (1959). *Zygnemaceae*. Indian Council of Agricultural Research, New Delhi, 478 pp.
- Sheath, R.G. & B.J. Hymes (1980). A preliminary investigation of the freshwater red algae in streams of southern Ontario, Canada. *Canadian Journal of Botany* 58: 1295–1318. https://doi. org/10.1139/b80-161
- Tessy, P.P. & R. Sreekumar (2017). Seasonal and spatial distribution of freshwater diatoms from Thrissur Kole lands (part of Vembanad -Kol, Ramsar site), Kerala. *Phykos* 47(2): 129–134.
- Turner, W.B. (1892). The fresh water algae (principally Desmidieae)

of East India. Kongl. Svenska Vetenskaps–Akademiens Handlingar 25(5): 1–187.

- Velayudhan, A., A. Mohanarangan, G. Chandy & S. Biju (2021). Does the size of the butterfly enhance detection? Factors influencing butterfly detection in species inventory surveys. *Journal of Threatened Taxa* 13(3): 17950–17962. https://doi.org/10.11609/ jott.6596.13.3.17950-17962
- Vijayan, D., K. Manivannam, S. Santhoshkumar, D. Pandiaraj, M.M. Imran, N. Thajuddin, K. Kala & M.H.M. Ilyas (2014). Depiction of Microalgal Diversity in Gundur Lake, Tiruchirappalli District, Tamil

Nadu, South India. Asian Journal of Biological Sciences 7(3): 111–121. https://doi.org/10.3923/ajbs.2014.111.121

- West, W. & G.S. West (1904). A Monograph of the British Desmidiaceae. The Ray Society, London, 311 pp.
- Wolowski, K. (1998). Taxonomic and environmental studies on Euglenophytes of the Krakow- Czestochowa upland (South Poland). Polish Academy of Sciences, Krakow, 192 pp.
- Wotowski, K. & F. Hindak (2005). Atlas of Euglenophytes. VEDA, Publishing House of the Slovak Academy of Sciences, Czechia, 136 pp.



Dr. John Noyes, Natural History Museum, London, UK

- Dr. Albert G. Orr, Griffith University, Nathan, Australia
- Dr. Sameer Padhye, Katholieke Universiteit Leuven, Belgium
- Dr. Nancy van der Poorten, Toronto, Canada Dr. Kareen Schnabel, NIWA, Wellington, New Zealand
- Dr. R.M. Sharma, (Retd.) Scientist, Zoological Survey of India, Pune, India
- Dr. Manju Siliwal, WILD, Coimbatore, Tamil Nadu, India
- Dr. G.P. Sinha, Botanical Survey of India, Allahabad, India
- Dr. K.A. Subramanian, Zoological Survey of India, New Alipore, Kolkata, India
- Dr. P.M. Sureshan, Zoological Survey of India, Kozhikode, Kerala, India
- Dr. R. Varatharajan, Manipur University, Imphal, Manipur, India Dr. Eduard Vives, Museu de Ciències Naturals de Barcelona, Terrassa, Spain
- Dr. James Young, Hong Kong Lepidopterists' Society, Hong Kong
- Dr. R. Sundararaj, Institute of Wood Science & Technology, Bengaluru, India

Dr. M. Nithyanandan, Environmental Department, La Ala Al Kuwait Real Estate. Co. K.S.C., Kuwait

- Dr. Himender Bharti, Punjabi University, Punjab, India
- Mr. Purnendu Roy, London, UK
- Dr. Saito Motoki, The Butterfly Society of Japan, Tokyo, Japan Dr. Sanjay Sondhi, TITLI TRUST, Kalpavriksh, Dehradun, India
- Dr. Nguyen Thi Phuong Lien, Vietnam Academy of Science and Technology, Hanoi, Vietnam
- Dr. Nitin Kulkarni, Tropical Research Institute, Jabalpur, India
- Dr. Robin Wen Jiang Ngiam, National Parks Board, Singapore
- Dr. Lional Monod, Natural History Museum of Geneva, Genève, Switzerland.
- Dr. Asheesh Shivam, Nehru Gram Bharti University, Allahabad, India
- Dr. Rosana Moreira da Rocha, Universidade Federal do Paraná, Curitiba, Brasil Dr. Kurt R. Arnold, North Dakota State University, Saxony, Germany
- Dr. James M. Carpenter, American Museum of Natural History, New York, USA
- Dr. David M. Claborn, Missouri State University, Springfield, USA
- Dr. Kareen Schnabel, Marine Biologist, Wellington, New Zealand
- Dr. Amazonas Chagas Júnior, Universidade Federal de Mato Grosso, Cuiabá, Brasil
- Mr. Monsoon Jyoti Gogoi, Assam University, Silchar, Assam, India Dr. Heo Chong Chin, Universiti Teknologi MARA (UITM), Selangor, Malaysia
- Dr. R.J. Shiel, University of Adelaide, SA 5005, Australia
- Dr. Siddharth Kulkarni, The George Washington University, Washington, USA
- Dr. Priyadarsanan Dharma Rajan, ATREE, Bengaluru, India
- Dr. Phil Alderslade, CSIRO Marine And Atmospheric Research, Hobart, Australia
- Dr. John E.N. Veron, Coral Reef Research, Townsville, Australia
- Dr. Daniel Whitmore, State Museum of Natural History Stuttgart, Rosenstein, Germany.
- Dr. Yu-Feng Hsu, National Taiwan Normal University, Taipei City, Taiwan
- Dr. Keith V. Wolfe, Antioch, California, USA
- Dr. Siddharth Kulkarni, The Hormiga Lab, The George Washington University, Washington, D.C., USA
- Dr. Tomas Ditrich, Faculty of Education, University of South Bohemia in Ceske Budejovice, Czech Republic
- Dr. Mihaly Foldvari, Natural History Museum, University of Oslo, Norway
- Dr. V.P. Unival, Wildlife Institute of India, Dehradun, Uttarakhand 248001, India
- Dr. John T.D. Caleb, Zoological Survey of India, Kolkata, West Bengal, India
- Dr. Priyadarsanan Dharma Rajan, Ashoka Trust for Research in Ecology and the Environment (ATREE), Royal Enclave, Bangalore, Karnataka, India

Fishes

- Dr. Neelesh Dahanukar, IISER, Pune, Maharashtra, India
- Dr. Topiltzin Contreras MacBeath, Universidad Autónoma del estado de Morelos, México
- Dr. Heok Hee Ng, National University of Singapore, Science Drive, Singapore
- Dr. Rajeev Raghavan, St. Albert's College, Kochi, Kerala, India Dr. Robert D. Sluka, Chiltern Gateway Project, A Rocha UK, Southall, Middlesex, UK
- Dr. E. Vivekanandan, Central Marine Fisheries Research Institute, Chennai, India
- Dr. Davor Zanella, University of Zagreb, Zagreb, Croatia Dr. A. Biju Kumar, University of Kerala, Thiruvananthapuram, Kerala, India
- Dr. Akhilesh K.V., ICAR-Central Marine Fisheries Research Institute, Mumbai Research
- Centre, Mumbai, Maharashtra, India
- Dr. J.A. Johnson, Wildlife Institute of India, Dehradun, Uttarakhand, India

Amphibians

- Dr. Sushil K. Dutta, Indian Institute of Science, Bengaluru, Karnataka, India
- Dr. Annemarie Ohler, Muséum national d'Histoire naturelle, Paris, France

Reptiles

cal Records.

NAAS rating (India) 5.64

- Dr. Gernot Vogel, Heidelberg, Germany
- Dr. Raju Vyas, Vadodara, Gujarat, India
- Dr. Pritpal S. Soorae, Environment Agency, Abu Dubai, UAE.
- Prof. Dr. Wayne J. Fuller, Near East University, Mersin, Turkey
- Prof. Chandrashekher U. Rivonker, Goa University, Taleigao Plateau, Goa. India

Journal of Threatened Taxa is indexed/abstracted in Bibliography of Systematic Mycology, Biological Abstracts, BIOSIS Previews, CAB Abstracts, EBSCO, Google Scholar, Index Copernicus, Index Fungorum, JournalSeek,

National Academy of Agricultural Sciences, NewJour, OCLC WorldCat,

SCOPUS, Stanford University Libraries, Virtual Library of Biology, Zoologi-

- Dr. S.R. Ganesh, Chennai Snake Park, Chennai, Tamil Nadu, India
- Dr. Himansu Sekhar Das, Terrestrial & Marine Biodiversity, Abu Dhabi, UAE

Birds

- Dr. Hem Sagar Baral, Charles Sturt University, NSW Australia
- Mr. H. Byju, Coimbatore, Tamil Nadu, India
- Dr. Chris Bowden, Royal Society for the Protection of Birds, Sandy, UK Dr. Priya Davidar, Pondicherry University, Kalapet, Puducherry, India
- Dr. J.W. Duckworth, IUCN SSC, Bath, UK
- Dr. Rajah Jayapal, SACON, Coimbatore, Tamil Nadu, India
- Dr. Rajiv S. Kalsi, M.L.N. College, Yamuna Nagar, Haryana, India
- Dr. V. Santharam, Rishi Valley Education Centre, Chittoor Dt., Andhra Pradesh, India
- Dr. S. Balachandran, Bombay Natural History Society, Mumbai, India Mr. J. Praveen, Bengaluru, India
- Dr. C. Srinivasulu, Osmania University, Hyderabad, India
- Dr. K.S. Gopi Sundar, International Crane Foundation, Baraboo, USA
- Dr. Gombobaatar Sundev, Professor of Ornithology, Ulaanbaatar, Mongolia
- Prof. Reuven Yosef, International Birding & Research Centre, Eilat, Israel
- Dr. Taej Mundkur, Wetlands International, Wageningen, The Netherlands
- Dr. Carol Inskipp, Bishop Auckland Co., Durham, UK
- Dr. Tim Inskipp, Bishop Auckland Co., Durham, UK Dr. V. Gokula, National College, Tiruchirappalli, Tamil Nadu, India
- Dr. Arkady Lelej, Russian Academy of Sciences, Vladivostok, Russia Dr. Simon Dowell, Science Director, Chester Zoo, UK
- Dr. Mário Gabriel Santiago dos Santos, Universidade de Trás-os-Montes e Alto Douro, Quinta de Prados, Vila Real, Portugal
- Dr. Grant Connette, Smithsonian Institution, Royal, VA, USA
- Dr. M. Zafar-ul Islam, Prince Saud Al Faisal Wildlife Research Center, Taif, Saudi Arabia

Mammals

- Dr. Giovanni Amori, CNR Institute of Ecosystem Studies, Rome, Italy
- Dr. Anwaruddin Chowdhury, Guwahati, India
- Dr. David Mallon, Zoological Society of London, UK
- Dr. Shomita Mukherjee, SACON, Coimbatore, Tamil Nadu, India Dr. Angie Appel, Wild Cat Network, Germany
- Dr. P.O. Nameer, Kerala Agricultural University, Thrissur, Kerala, India
- Dr. Ian Redmond, UNEP Convention on Migratory Species, Lansdown, UK
- Dr. Heidi S. Riddle, Riddle's Elephant and Wildlife Sanctuary, Arkansas, USA

Dr. Justus Joshua, Green Future Foundation, Tiruchirapalli, Tamil Nadu, India

Dr. Jim Sanderson, Small Wild Cat Conservation Foundation, Hartford, USA

Dr. David Mallon, Manchester Metropolitan University, Derbyshire, UK

Dr. Brian L. Cypher, California State University-Stanislaus, Bakersfield, CA Dr. S.S. Talmale, Zoological Survey of India, Pune, Maharashtra, India

Prof. Karan Bahadur Shah, Budhanilakantha Municipality, Kathmandu, Nepal

Dr. Hemanta Kafley, Wildlife Sciences, Tarleton State University, Texas, USA

Dr. Aniruddha Belsare, Columbia MO 65203, USA (Veterinary)

Dr. Ulrike Streicher, University of Oregon, Eugene, USA (Veterinary)

Dr. Hari Balasubramanian, EcoAdvisors, Nova Scotia, Canada (Communities) Dr. Rayanna Hellem Santos Bezerra, Universidade Federal de Sergipe, São Cristóvão, Brazil Dr. Jamie R. Wood, Landcare Research, Canterbury, New Zealand Dr. Wendy Collinson-Jonker, Endangered Wildlife Trust, Gauteng, South Africa

Dr. Rajeshkumar G. Jani, Anand Agricultural University, Anand, Gujarat, India

Dr. Rupika S. Rajakaruna, University of Peradeniya, Peradeniya, Sri Lanka

Due to pausity of space, the list of reviewers for 2018-2020 is available online.

The opinions expressed by the authors do not reflect the views of the Journal of Threatened Taxa, Wildlife Information Liaison Development Society, Zoo Outreach Organization, or any of the partners. The journal, the publisher, the host, and the partners are not responsible for the accuracy of the political

Dr. Bahar Baviskar, Wild-CER, Nagpur, Maharashtra 440013, India

boundaries shown in the maps by the authors.

Print copies of the Journal are available at cost. Write to:

c/o Wildlife Information Liaison Development Society, No. 12, Thiruvannamalai Nagar, Saravanampatti - Kalapatti Road,

Saravanampatti, Coimbatore, Tamil Nadu 641035, India

Dr. O.N. Tiwari, Senior Scientist, ICAR-Indian Agricultural Research Institute (IARI), New

Dr. L.D. Singla, Guru Angad Dev Veterinary and Animal Sciences University, Ludhiana, India

Dr. Susan Cheyne, Borneo Nature Foundation International, Palangkaraja, Indonesia

Dr. Mandar S. Paingankar, University of Pune, Pune, Maharashtra, India (Molecular) Dr. Jack Tordoff, Critical Ecosystem Partnership Fund, Arlington, USA (Communities)

Dr. H. Raghuram, The American College, Madurai, Tamil Nadu, India

Dr. Spartaco Gippoliti, Socio Onorario Società Italiana per la Storia della Fauna "Giuseppe

- Dr. Karin Schwartz, George Mason University, Fairfax, Virginia.
- Dr. Lala A.K. Singh, Bhubaneswar, Orissa, India
- Dr. Mewa Singh, Mysore University, Mysore, India
- Dr. Paul Racey, University of Exeter, Devon, UK Dr. Honnavalli N. Kumara, SACON, Anaikatty P.O., Coimbatore, Tamil Nadu, India

Dr. Paul Bates, Harison Institute, Kent, UK

Altobello", Rome, Italy

Other Disciplines

Delhi, India

Reviewers 2019-2021

The Managing Editor, JoTT,

ravi@threatenedtaxa.org

Dr. Nishith Dharaiya, HNG University, Patan, Gujarat, India

Dr. Dan Challender, University of Kent, Canterbury, UK





The Journal of Threatened Taxa (JoTT) is dedicated to building evidence for conservation globally by publishing peer-reviewed articles online every month at a reasonably rapid rate at www.threatenedtaxa.org. All articles published in JoTT are registered under Creative Commons Attribution 4.0 International License unless otherwise mentioned. JoTT allows allows unrestricted use, reproduction, and distribution of articles in any medium by providing adequate credit to the author(s) and the source of publication.

ISSN 0974-7907 (Online) | ISSN 0974-7893 (Print)

June 2022 | Vol. 14 | No. 6 | Pages: 21127–21330 Date of Publication: 26 June 2022 (Online & Print) DOI: 10.11609/jott.2022.14.6.21127-21330

www.threatenedtaxa.org

Article

Identification of confiscated pangolin for conservation purposes through molecular approach

- Wirdateti, R. Taufiq P. Nugraha, Yulianto & Gono Semiadi, Pp. 21127-21139

Communications

The trade of Saiga Antelope horn for traditional medicine in Thailand – Lalita Gomez, Penthai Siriwat & Chris R. Shepherd, Pp. 21140–21148

The occurrence of Indochinese Serow Capricornis sumatraensis in Virachey National Park, northeastern Cambodia

- Gregory McCann, Keith Pawlowski & Thon Soukhon, Pp. 21149–21154

Attitudes and perceptions of people about the Capped Langur Trachypithecus pileatus (Mammalia: Primates: Cercopithecidae): a preliminary study in Barail Wildlife Sanctuary, India

 – Rofik Ahmed Barbhuiya, Amir Sohail Choudhury, Nazimur Rahman Talukdar & Parthankar Choudhury, Pp. 21155–21160

Feather characteristics of Common Myna Acridotheres tristis (Passeriformes: Sturnidae) from India

 – Swapna Devi Ray, Goldin Quadros, Prateek Dey, Padmanabhan Pramod & Ram Pratap Singh, Pp. 21161–21169

Population and distribution of Wattled Crane *Bugeranus carunculatus*, Gmelin, 1989 at lake Tana area, Ethiopia

- Shimelis Aynalem Zelelew & George William Archibald, Pp. 21170-21178

Waterbird assemblage along Punatsangchhu River, Punakha and Wangdue Phodrang, Bhutan

– Nima & Ugyen Dorji, Pp. 21179–21189

Freshwater fishes of the Chimmony Wildlife Sanctuary, Western Ghats, India – P.S. Eldho & M.K. Saieevan, Pp. 21190–21198

Butterflies of Eravikulam National Park and its environs in the Western Ghats of Kerala, India

Kalesh Sadasivan, Toms Augustine, Edayillam Kunhikrishnan & Baiju Kochunarayanan, Pp. 21199–21212

The dragonflies and damselflies (Insecta: Odonata) of Shendurney Wildlife Sanctuary, southern Western Ghats, India

- Kalesh Sadasivan, Vinayan P. Nair & K. Abraham Samuel, Pp. 21213-21226

A pioneering study on the spider fauna (Arachnida: Araneae) of Sagar District, Madhya Pradesh, India

- Tanmaya Rani Sethy & Janak Ahi, Pp. 21227-21238

Taxonomy and threat assessment of *Lagotis kunawurensis* Rupr (Plantaginaceae), an endemic medicinal plant species of the Himalaya, India – Aijaz Hassan Ganie, Tariq Ahmad Butt, Anzar Ahmad Khuroo, Nazima Rasool, Rameez

Ahmad, Syed Basharat & Zafar A. Reshi, Pp. 21239–21245

The study of algal diversity from fresh water bodies of Chimmony Wildlife Sanctuary, Kerala, India

- Joel Jose & Jobi Xavier, Pp. 21246-21265

Review

A checklist of herpetofauna of Telangana state, India

- Chelmala Srinivasulu & Gandla Chethan Kumar, Pp. 21266–21281

Viewpoint

Comments on "The Dragonflies and Damselflies (Odonata) of Kerala – Status and Distribution" – A. Vivek Chandran & K. Muhamed Sherif, Pp. 21282–21284

Short Communications

Landings of IUCN Red Listed finfishes at Chetlat Island of Lakshadweep, southeastern Arabian Sea

- Davood Nihal, N.M. Naseem, N. Abhirami & M.P. Prabhakaran, Pp. 21285-21289

First report of the termite *Glyptotermes ceylonicus* (Blattodea: Isoptera: Kalotermitidae) from India: an example of discontinuous distribution

– Edwin Joseph, Chinnu Ipe, Nisha P. Aravind, Sherin Antony & Jobin Mathew, Pp. 21290–21295

Authentic report of the emesine bug *Gardena melinarthrum* Dohrn, 1860 (Hemiptera: Heteroptera: Reduviidae) from India

– Sangamesh R. Hiremath, Santana Saikia & Hemant V. Ghate, Pp. 21296–21301

Reappearance of stomatopod *Gonodactylus platysoma* (Wood-Mason, 1895) after an era from the intertidal region of Chota Balu, South Andaman, India

– N. Muthu Mohammed Naha, Limaangnen Pongener & G. Padmavati, Pp. 21302–21306

Range extension of earthworm *Drawida impertusa* Stephenson, 1920 (Clitellata: Moniligastridae) in Karnataka, India – Vivek Hasyagar, S. Prasanth Narayanan & K.S. Sreepada, Pp. 21307–21310

Pelatantheria insectifera (Rchb.f.) Ridl. (Orchidaceae): a new generic record for Eastern Ghats of Andhra Pradesh. India

– V. Ashok Kumar, P. Janaki Rao, J. Prakasa Rao, S.B. Padal & C. Sudhakar Reddy, Pp. 21311–21314

Notes

New breeding site record of Oriental White Ibis Threskiornis melanocephalus (Aves: Threskiornithidae) at Thirunavaya wetlands, Kerala, India – Binu Chullakattil, Pp. 21315–21317

Rediscovery of Gardena melinarthrum Dohrn from Sri Lanka – Tharindu Ranasinghe & Hemant V. Ghate, Pp. 21318–21320

A report on the occurrence of the cicada *Callogaeana festiva* (Fabricius, 1803) (Insecta: Cicadidae) from Mizoram, India

- Khawlhring Marova, Fanai Malsawmdawngliana, Lal Muansanga & Hmar Tlawmte Lalremsanga, Pp. 21321–21323

New distribution records of two species of metallic ground beetles of the genus *Chlaenius* (Coleoptera: Carabidae: Chlaeniini) from the Western Ghats, India – Duraikannu Vasanthakumar & Erich Kirschenhofer, Pp. 21324–21326

Report of *Euphaea pseudodispar* Sadasivan & Bhakare, 2021 (Insecta: Odonata) from Kerala, India

- P.K. Muneer, M. Madhavan & A. Vivek Chandran, Pp. 21327-21330



