

OPEN ACCESS



The Journal of Threatened Taxa (JoTT) is dedicated to building evidence for conservation globally by publishing peer-reviewed articles online every month at a reasonably rapid rate at www.threatenedtaxa.org. All articles published in JoTT are registered under [Creative Commons Attribution 4.0 International License](https://creativecommons.org/licenses/by/4.0/) unless otherwise mentioned. JoTT allows unrestricted use, reproduction, and distribution of articles in any medium by providing adequate credit to the author(s) and the source of publication.

Journal of Threatened Taxa

Building evidence for conservation globally

www.threatenedtaxa.org

ISSN 0974-7907 (Online) | ISSN 0974-7893 (Print)

COMMUNICATION

ON THE REPRODUCTIVE BIOLOGY OF THE INVASIVE ARMoured SAILFIN CATFISH *PTERYGOLICHTHYS PARDALIS* (CASTELNAU, 1855) (SILURIFORMES: LORICARIIDAE) FROM THE NATURAL DRAINAGES IN THIRUVANANTHAPURAM, INDIA

Smrithy Raj, Suvarna S. Devi, Amrutha Joy & A. Biju Kumar

26 August 2021 | Vol. 13 | No. 9 | Pages: 19263–19273

DOI: 10.11609/jott.7164.13.9.19263-19273



For Focus, Scope, Aims, and Policies, visit https://threatenedtaxa.org/index.php/JoTT/aims_scope

For Article Submission Guidelines, visit <https://threatenedtaxa.org/index.php/JoTT/about/submissions>

For Policies against Scientific Misconduct, visit https://threatenedtaxa.org/index.php/JoTT/policies_various

For reprints, contact [<ravi@threatenedtaxa.org>](mailto:ravi@threatenedtaxa.org)

The opinions expressed by the authors do not reflect the views of the Journal of Threatened Taxa, Wildlife Information Liaison Development Society, Zoo Outreach Organization, or any of the partners. The journal, the publisher, the host, and the partners are not responsible for the accuracy of the political boundaries shown in the maps by the authors.

Publisher & Host



**The challenges of the climate crisis are frustrating.
Learn to lead to make a positive change.**



The Ram Hattikudur Advanced Training in Conservation (RHATC) is a four-month residential course mentored by Indian and international experts. The course will bridge the gap between academics and on-ground conservation realities by equipping you with knowledge, tools, and an understanding of global conservation issues.

Challenge yourself

- Resolve conservation challenges.
- Develop skills in assessments and planning.
- Exposure to real-time conservation needs.
- A window into conservation NGOs.
- Potential opportunities for internship.
- Potential job opportunities with conservation organizations.
- Pursue conservation careers
- Potential to start your own organization.
- Exposure to conservation experts.
- Develop leadership skills.

Apply now!

Applications open: 09 August 2021
Application last date: 31 August 2021
Course start date: 12 October 2021

To know more visit:
www.rhatc.zooreach.org





On the reproductive biology of the invasive Armoured Sailfin Catfish *Pterygoplichthys pardalis* (Castelnau, 1855) (Siluriformes: Loricariidae) from the natural drainages in Thiruvananthapuram, India

Smrithy Raj¹ , Suvarna S. Devi² , Amrutha Joy³ & A. Biju Kumar⁴

¹⁻⁴ Department of Aquatic Biology & Fisheries, University of Kerala, Thiruvananthapuram, Kerala 695581, India.

¹smrithyaqb@gmail.com, ²suvarnaraja1995@gmail.com, ³amruthajoy2@gmail.com,

⁴bujukumar@keralauniversity.ac.in (corresponding author)

Abstract: The present paper deals with the breeding biology of the invasive fish *Pterygoplichthys pardalis* from the natural drainages of Thiruvananthapuram, Kerala. The specimens were collected from Amayizhanchan Thodu, a natural drainage running through the heart of the city. A total of 145 males and 142 females were collected from January to December 2018. The sex ratio was determined monthly as the percentage of males to females (M: F). Monthly mean gonadosomatic index (GSI) values were compared using R stat, and GSI was plotted monthly to identify the spawning seasons. The gonads were examined and different stages of maturity were noted using standard methods. The length at first maturity was also found out. The fish exhibit courtship behaviour and the eggs are deposited in burrows and also along the crevices in the granite walls; the burrows are guarded by the male fish till the young ones are hatched out. The sex ratio showed an average mean value of 1.04: 1 and showed no significant departure. The size at first maturity was 23.9 cm standard length. The ova diameter studies show the presence of ripe ovaries throughout the year, with peaks during March and April and between August and September and in December, indicating the fish is a batch spawner. The absolute fecundity ranges from 923 to 14,777 eggs, and the relative fecundity ranges 0.0142–0.0015. Regression analysis showed a significant relationship ($P < 0.001$) between absolute fecundity and the total length, the total body weight, and ovary weight. The strong breeding behaviour, the presence of accessory respiratory organs, the absence of natural enemies and parental care makes *Pterygoplichthys pardalis* a successful invader in the natural drainage. More biological studies are needed for the successful eradication of the species from the invaded ecosystem.

Keywords: Breeding biology, breeding behaviour, fecundity, invasive alien species, invasion biology.

Malayalam: കേരളത്തിൽ തിരുവനന്തപുരം നഗരത്തിലെ സാമാന്യ നിർമ്മാണങ്ങളിൽ ജീവിക്കുന്ന ആമസോൺ സക്കർ മത്സ്യത്തിന്റെ (പ്റ്റെറിഗോപ്ലിക്തിസ് പർഡാലിസ്) പ്രജനന ജീവശാസ്ത്രത്തെക്കുറിച്ചാണ് ഈ പ്രബന്ധത്തിൽ വിവരിക്കുന്നത്. നഗരത്തിന്റെ ഹൃദയഭാഗത്തുകൂടി ഒഴുകുന്ന പ്രകൃതിദത്ത ഡ്രെയിനേജ് ആയ അമയിച്ചോൽ തോട്ടിൽ നിന്നാണ് സാമ്പിളുകൾ ശേഖരിച്ചത്. 2018 ജനുവരി മുതൽ ഡിസംബർ വരെ മൊത്തം 145 ആൺ മത്സ്യങ്ങളെയും 142 പെൺ മത്സ്യങ്ങളെയും ശേഖരിച്ചു. ലിംഗ അനുപാതം പ്രതിമാസം ആൺ മത്സ്യങ്ങളുടെ ശതമാനം പെൺ മത്സ്യങ്ങളുടേതുമായി താരതമ്യം ചെയ്തു. പ്രതിമാസ ശരാശരി ഗോണോസോമാറ്റിക് സൂചിക (GSI) മൂല്യങ്ങൾ R സ്റ്റാറ്റ് ഉപയോഗിച്ച് താരതമ്യം ചെയ്തു. മുട്ടയിടുന്ന സീസണുകൾ തിരിച്ചറിയാൻ GSI പ്രതിമാസം ഗ്രാഫുകളിൽ രേഖപ്പെടുത്തി. ജനനഗ്രന്ഥികൾ (ഗോണാഡുകൾ) പരിശോധിക്കുകയും ശാസ്ത്രീയ രീതികൾ ഉപയോഗിച്ച് വളർച്ചയുടെ വിവിധ ഘട്ടങ്ങൾ പരിശോധിക്കുകയും ചെയ്തു. ആദ്യമായി പ്രായപൂർത്തിയാകുന്ന ഓർഗ്സവും കണ്ടെത്തി. മത്സ്യങ്ങൾ കോർട്ട്ഷിപ്പ് സജ്ജമാക്കിയിട്ടുള്ളതും മുട്ടകൾ മാറ്റങ്ങളിലും കരിങ്കൽ ഭിത്തികളിലും വിളക്കുകളിലും നിക്ഷേപിക്കുകയും ചെയ്യുന്നു. കുഞ്ഞുങ്ങൾ വിരിയുന്നതുവരെ ആൺമത്സ്യം കാവൽ നിന്ന് മുട്ടകളെ സംരക്ഷിക്കുന്നു. ലിംഗാനുപാതത്തിന്റെ ശരാശരി മൂല്യം 1.04:1 ആണ്. ആദ്യമായി പ്രായപൂർത്തിയാകുന്ന വലുപ്പം 23.9 സെന്റിമീറ്റർ സ്റ്റാൻഡേർഡ് നീളമായിരുന്നു. അണ്ഡത്തിന്റെ വ്യാസത്തെപ്പറ്റിയുള്ള പഠനങ്ങൾ വർഷം മുഴുവനും മാർച്ച് ഏപ്രിൽ മാസങ്ങളിലും ഓഗസ്റ്റ് മുതൽ സെപ്റ്റംബർ വരെയും ഡിസംബറിലും പൂർണ്ണവളർച്ചയെത്തിയ അണ്ഡത്തിന്റെ സാന്നിധ്യം കാണിക്കുന്നു. ഇത് മത്സ്യം വർഷം മുഴുവനും മുട്ടയിടാനുള്ള (ബാച്ച് സ്പോൺ) സാധ്യത സൂചിപ്പിക്കുന്നു. മുട്ടയുടെ കേവലമായ എണ്ണം 923 മുതൽ 14,777 മുട്ടകൾ വരെയാണ്; ആപേക്ഷികമായ മുട്ടയുടെ 0.0142-0.0015 വരെയാണ്. മൊത്തം മുട്ടകളുടെ എണ്ണവും, മൊത്തം ഓർഗ്സവും, മൊത്തം ശരീരഭാരവും അണ്ഡാശയ ഭാരവും തമ്മിൽ സാരവത്തായ ബന്ധം (പി < 0.001) നിലനിൽക്കുന്നു. ശക്തമായ പ്രജനനരീതികളും, സഹായക ശ്വാസോപാധങ്ങളുടെ സാന്നിധ്യവും, സാമാന്യ ശത്രുക്കളുടെ അഭാവവും രക്ഷാകർത്ഥ പരിചരണവും ആണ് ആമസോൺ സക്കർ മത്സ്യത്തെ പ്രകൃതിദത്ത നിർമ്മാണങ്ങളിൽ വിജയകരമായി ആധിപത്യം സ്ഥാപിക്കുന്നതിന് സഹായിച്ചിരിക്കുന്നത് അനുമാനിക്കാം. അധിനിവേശ ആവാസവ്യവസ്ഥയിൽ നിന്ന് ഇവയെ വിജയകരമായി തുടച്ചുനീക്കുന്നതിന് കൂടുതൽ ജീവശാസ്ത്രപരമായ പഠനങ്ങൾ ആവശ്യമാണ്.

Editor: Anonymity requested.

Date of publication: 26 August 2021 (online & print)

Citation: Raj, S., S.S. Devi, A. Joy & A.B. Kumar (2021). On the reproductive biology of the invasive Armoured Sailfin Catfish *Pterygoplichthys pardalis* (Castelnau, 1855) (Siluriformes: Loricariidae) from the natural drainages in Thiruvananthapuram, India. *Journal of Threatened Taxa* 13(9): 19263–19273. <https://doi.org/10.11609/jott.7164.13.9.19263-19273>

Copyright: © Raj et al. 2021. Creative Commons Attribution 4.0 International License. JoTT allows unrestricted use, reproduction, and distribution of this article in any medium by providing adequate credit to the author(s) and the source of publication.

Funding: This research was funded by the Directorate of Environment and Climate Change, Government of Kerala, India.

Competing interests: The authors declare no competing interests.

Author details, Author contributions and Acknowledgements: See end of this article



INTRODUCTION

Reproductive biology is one of the key features considered for the invasion biology studies of fish (Feiner et al. 2012; Gutkowsky & Fox 2012; Zahorska et al. 2013; Horkova & Kovac 2015). The suckermouth armoured catfishes of the family Loricariidae, especially the genus *Pterygoplichthys* T.N. Gill, native to inland water bodies of South America, is an emerging invasive fish globally (Orfinger & Goodding 2018), also, one of the most popular and commonly traded aquarium fish in India (Knight 2010). The sailfin armoured catfish species recorded from the natural water bodies of India include *P. anisitsi*, *P. disjunctivus*, *P. multiradiatus*, *P. pardalis*, and possible hybrids (Sinha et al. 2010; Singh 2014; Kumar et al. 2015; Soundararajan et al. 2015; Sandilyan et al. 2016; Hussan et al. 2018).

The high invasiveness shown by this species is primarily because of its unique biological features, which include the ability to survive in water with varying levels of flow regimes (Nico & Martin 2001; Welcomme & Vidthayanom 2003; Chavez et al. 2006; Nico et al. 2012), temperature and dissolved oxygen using accessory respiration with diverticula of the gastrointestinal tract (Armbruster 1998; da Cruz et al. 2013), and pH (Mendoza et al. 2009; Parente et al. 2017), a broad range of diet (German et al. 2010) and the absence of natural predators (Nico & Martin 2001; Gibbs et al. 2008), which enable them to survive in the invaded ecosystems. They are salt-tolerant and survive well in brackish water regions (Mendoza et al. 2009; Kumar et al. 2018). They also have efficient reproductive strategies, including parental care and deter predation by possessing bony plates that cover their body (Hoover et al. 2004; Liang et al. 2005; Wei et al. 2017). One such extreme measure of parental care is exhibited by the male members of the genus *Pterygoplichthys*. It is the burrows they dig out on the river banks, and the female will lay eggs that are guarded by males (Nico et al. 2009; Alamdin & Jumawan 2016). Thus, site selection for spawning, nest building and caring of eggs and the young ones are a complex set of reproductive behaviour among loricariids (Covain & Fisch-Muller 2007).

The reproductive traits of *Pterygoplichthys* spp such as size at maturity, spawning, sex-ratio, ova diameter, fecundity and reproductive plasticity are studied to have a better knowledge on the reproductive dynamics (Jumawan and Herrera, 2014; Gibbs et al., 2017) which is a prerequisite for the management measures of an invasive species (Hoover et al., 2005; Kopp et al. 2009). The reproductive patterns of *P. disjunctivus* from Florida

were extensively analysed by Gibbs et al. (2017) for 10 years. In contrast, studies on the particular aspects of reproduction have been published by Rueda-Jasso et al. (2013), Jumawa & Herrera (2014) on *P. disjunctivus* from Mexico and Philippines, Samat et al. (2016) on *P. pardalis* from Malaysian waters, and Cook-Hildreth et al. (2016) from Texas waters. Wei et al. (2017) studied the maturity of *Pterygoplichthys* spp., a hybrid from China and reported self-sustaining populations in the drainages of the area.

The negative impacts caused by *Pterygoplichthys* spp. in the invaded ecosystems, include siltation problems, bank erosion in rivers and streams, competition with native species for food and space, consumption of the eggs of native and threatened species, displacement of vegetation and disturbance to the breeding grounds of native fish and economic losses to the fishermen including damage to the fishing gears, are reported earlier by many researchers (see Bunkley-Williams et al. 1994; Hoover et al. 2004; Chavez et al. 2006; Wakida-Kusunoki et al. 2007; Hossain et al. 2008; Cook-Hildreth 2009; Krishnakumar et al. 2009; Mendoza et al. 2009; Nico et al. 2009; Capps & Flecker 2013).

The reproductive parameters of oviparous fish outside their natural range will supplement the evidence to comprehend their establishment in the invaded ecosystems (Samat et al. 2016). Such studies, especially on reproductive biology, are required to better understand the natural history and reproductive plasticity, which are necessary tools for effectively managing this emergent invasive species. Despite the increasing numbers of publications regarding the invasion range extension of *Pterygoplichthys* spp. in various biogeographic regions, a knowledge gap on the reproductive biology of this invasive fish persists in India. Hence to address this significant gap, we investigated the reproductive biology of invasive loricariid fish *Pterygoplichthys pardalis* with a description of their breeding behaviour in natural streams.

MATERIALS AND METHODS

Collection Site and Sampling

The fishes for the study were collected from Amayizhanchan Thodu, (8.484711 – 8.566293°N; 76.933348 – 76.949982°E), a natural drainage of 3.4 km² in Thiruvananthapuram city, Kerala (Figure 1; Image 1). A total of 145 males and 142 females were collected fortnightly from January to December 2018 using a cast net, 5-m long and 3.8-cm mesh size. The fishes were

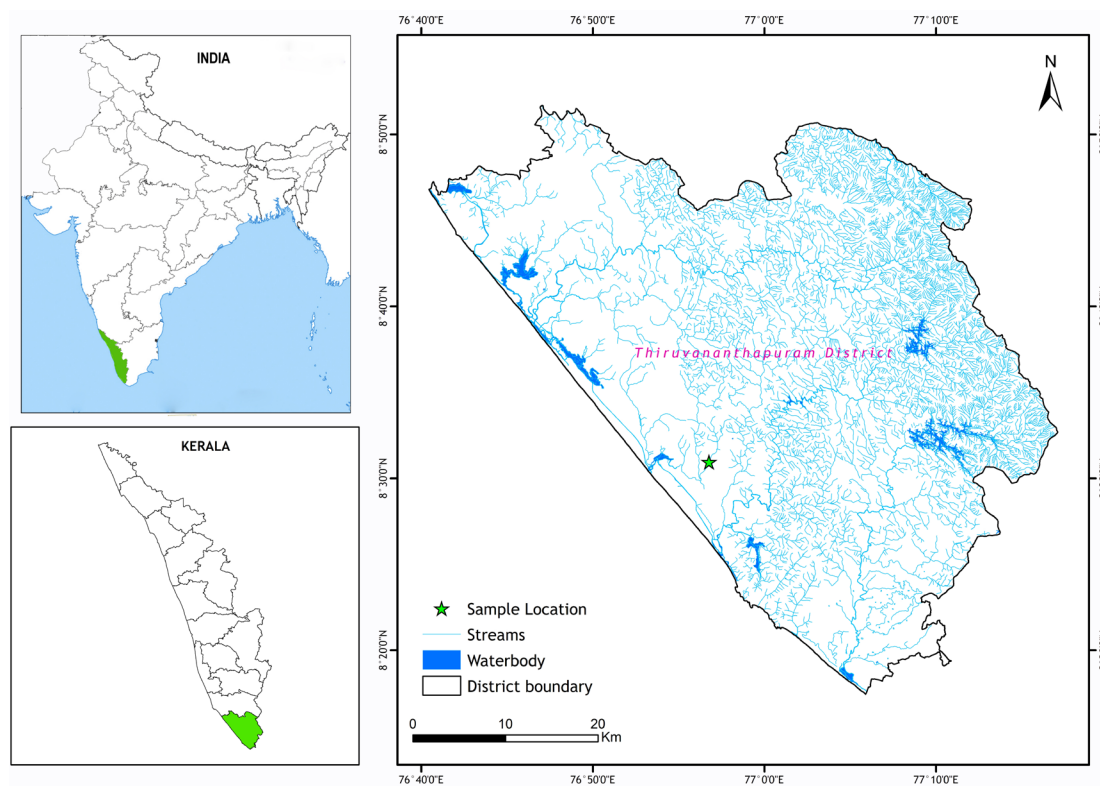


Figure 1. Sampling location at Amayinzhanthodu, Thiruvananthapuram, Kerala, India.



Image 1. *Pterygoplichthys pardalis*: A—lateral view | B—catch in cast net | C—specimens caught in single cast netting. © Biju Kumar

dissected, but before that, they were kept for four hours in a deep freezer. The sex ratio was determined monthly as the percentage of males to females (M: F). The burrow structure was captured with a digital camera, and its width, diameter, depth, the maximum height of entrance, shape of the tunnel, condition of the burrow, and occupancy of burrows were noted (Image 2).

Measurements of total length (TL, cm), standard length (SL), total weight (TW, g) and gonad weight (GW, g) were taken for male and female specimens. Total length and standard length were measured to the nearest 0.1 cm, and weight to the nearest gram 0.1 g using a digital balance. Gonadosomatic index (GSI) was calculated as $GSI = [(gonad\ weight)/(total\ weight\ of\ fish)] \times 100$ (Vazzoler, 1996). Monthly mean GSI values were compared using R stat, and GSI was plotted monthly to identify the spawning seasons.

The fishes were dissected to remove their gonads. Stages of maturity were determined following (Mazzoni & Caramaschi 1997) and ova diameter observations. The number of fishes in various stages include: immature (16), maturing (33), ripe (45), regressing spent (18), and recovering spent (30). Ovaries were excised, weighed to the nearest gram, and preserved in 10 per cent formalin for later assessment. Spawning type was designated according to the stage of the ripe and spent ovaries. Testis was characterized using a visual-based macro scale of maturity based on testicular size, colour, and swelling (Lowerre-Barbieri et al. 2011).

The length at first maturity (L_{50}) was defined as the total length, where 50% of all individuals are predicted to be adults (Vazzoler 1996). For ova diameter studies, the diameter of maturing and mature eggs (Vitellogenic oocytes) was measured to the nearest 0.01 mm with digital calipers and from the photos taken using Leica stereo zoom dissecting microscope.

To evaluate the absolute fecundity (AF), ripe ovaries which are in late-maturing stages to ripe ones (mid to late vitellogenic phase- Patiño & Sullivan 2002) were taken for the studies, and 100 mg sub-samples of eggs from the anterior, middle, and posterior regions were weighed and counted under a binocular stereo microscope. The counted eggs were averaged and extrapolated for the entire ovaries using the formula: $F = nG/g$ where F is the fecundity, n is the number of eggs in the sub-sample, G, ovary weight and g the subsample weight. The number and size of eggs were determined under a binocular dissecting microscope. The gonadal cycle also has been estimated based on macroscopic observation, and five stages have been described (Araújo et al. 1998; Duarte & Araújo 2002). To evaluate the relative fecundity (RF), the

absolute fecundity is divided by the total weight of the fish (Bagenal 1978). The relationship between relative fecundity (RF) and the variables total body length (TL), the total fish weight (FW), and ovary weight (OW) were estimated.

RESULTS

Breeding behaviour

Pterygoplichthys pardalis that invaded the natural drainages in Thiruvananthapuram city (Images 1A–C) excavate burrows for breeding <<https://www.youtube.com/watch?v=h5VZ-SVw7Wc>>. Our observations reveal that the male fish excavate burrows (Image 2A, B) before spawning for laying eggs.

The burrows (older/used) above the water level are small and triangular to circular, measuring 10–20 cm in diameter. In contrast, those below water are larger without definite shape, measuring 30–50 cm in width. The horizontal burrows are 120–140 cm deep, with the slope extending downwards into the bank. Courtship behaviour was exhibited in the form of circular movements near the burrows (Images 2C–E), by rubbing their bodies with the flashing of water, and in a few cases, multiple males take part in the process, and the eggs are guarded till the young ones emerge from the nest (Image 2F).

It was also observed that this species also selects crevices in the granite walls of the stream to deposit eggs, which may be one of the reasons for their higher rate of survival in the drainages of Thiruvananthapuram city in Kerala.

Sex ratio

A total of 145 males and 142 females of *P. pardalis* collected from the study site showed sex ratio (M: F) of an average mean value of 1.04: 1. The ratio was tested by chi-square analysis for differences from hypothetical ratio 1: 1, which showed no significant departure.

Stages of the reproductive cycle

Females (N= 142) ranging from SL 17.2 cm (TL 24.4 mm) to 45.6 cm (TL 58.4 mm) were considered for ascertaining reproductive stages. Mature ovaries exhibited asymmetry, whereas immature ones were symmetrical. It was observed that the larger the size of the individual, the greater would be the occupancy of the ripe ovary in the abdominal cavity and vice versa. Based on macroscopic and microscopic examination (Table 1) and ova diameter studies, five developmental stages of

Table 1. Macroscopic and microscopic characteristics of ovarian maturity in *Pterygoplichthys pardalis*.

Stages of maturity	Macroscopic and microscopic features
1. Immature	Tiny ovaries, ranging 11.08–32.5mm, SL mostly less than 33.4 cm occupying only a tiny percentage of the body cavity. Ovary thin transparent to light pink, no visible oocytes
2. Maturing	Size of the ovary ranges between 33–59.36 mm with tiny granules to less yolky oocytes with SL around 35 cm, colour opaque to pale yellow
3. Ripe	Highly vascularised, thin-walled large asymmetrical ovaries occupying mostly half of the body cavity, brightly orange coloured fully yolked oocytes, size (1.88–2.81 mm)
4. Regressing spent	Large flaccid thick-walled ovaries usually with very few or no vitellogenic oocytes, vascularisation still visible but less, thick brush-like fimbriae projects from the ovarian wall into the lumen.
5. Recovering spent	Ovaries purple to dark pink with thick inner ovarian walls with slight vascularisation with small oocytes of different diameters, absence of ripe oocytes

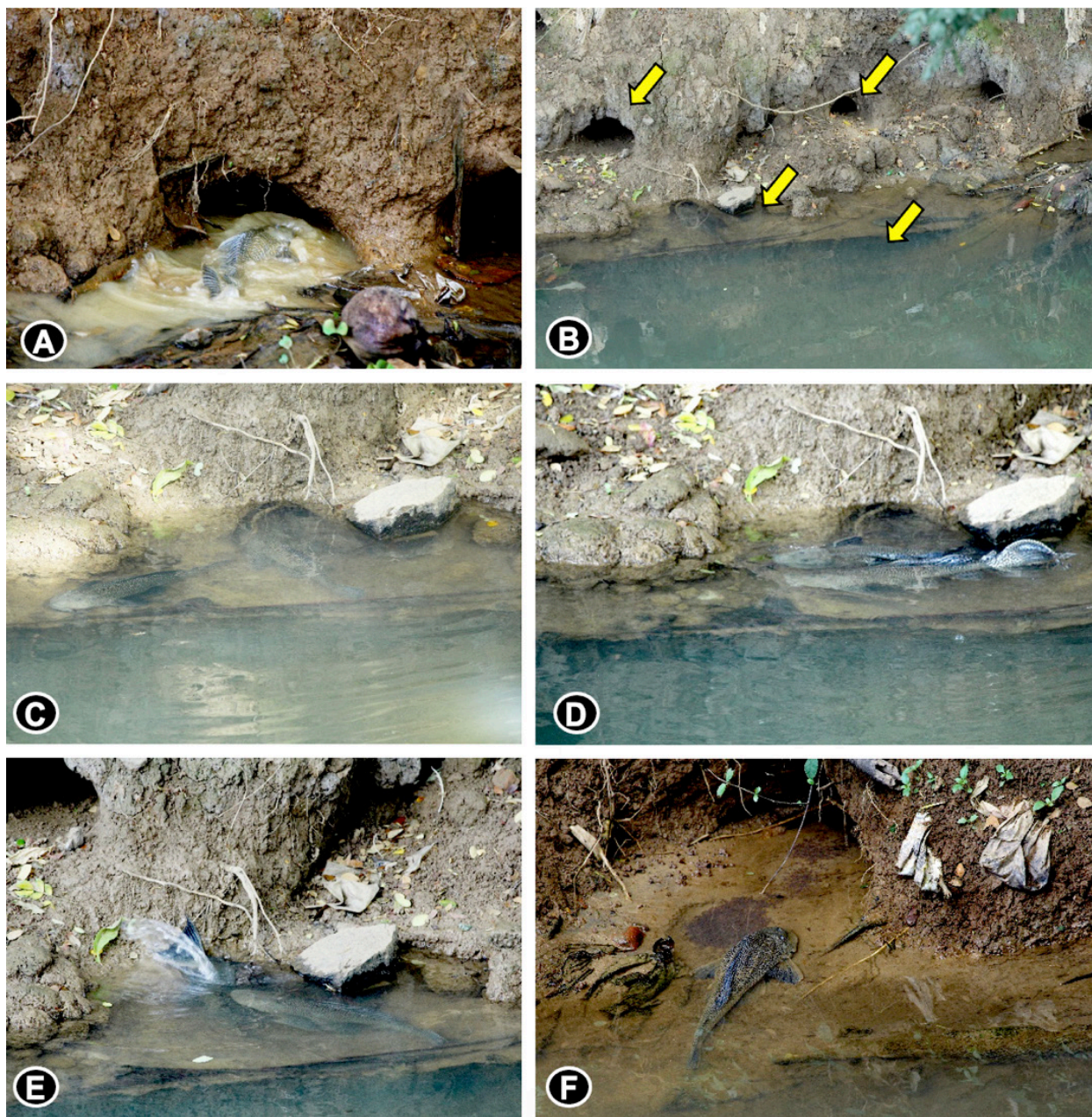


Image 2. Breeding behaviour of *Pterygoplichthys pardalis*: A—*P. pardalis* making burrows in canal margins | B—old and new burrows above and below the water level | C, D, E—courtship behaviour of *P. pardalis* around the burrow opening | F—*P. pardalis* guarding the burrow. <<https://www.youtube.com/watch?v=h5VZ-SVw7Wc>> © Biju Kumar

gonads (immature, maturing, ripe, regressing spent, and recovering spent) were identified in female fish (Table 1, Image 3 A–D, 4 A–F). In males, three maturity stages were identified: immature, maturing and mature, depending upon the colour and size of testes (Image 5 A–F).

The macroscopic and ova diameter studies showed *P. pardalis* has an intermittent spawning period with ripe ovaries throughout the year, maximum during March and April. Immature stages were noticed from May onwards, followed by maturing ones from June to November with a peak in July. The ripe ovaries were present in all months except February, with a maximum during March and April. Accordingly, the spent ovaries (regressive and recovering ones) and immature stages were noticed in the subsequent months, which mean the fish spawns during the rains (as summer rains occur during May). The second set of a large number of ripe ovaries was visible in August and December, with regressing and recovering spent in September and January, respectively showing the extended spawning season for the fish.

Length at first maturity

The minimum length to attain sexual maturity with vitellogenic oocytes was noticed in females at a standard length of 23.9 cm. The length at which 50% of the fish gets matured is at 36.56 cm (Figure 3).

Fecundity

To understand the absolute fecundity (AF), the ovary (left lobe) of ripe fishes were dissected, and the ripe ova were fully counted and extrapolated. The absolute fecundity of fish ranges from 923 (TL 393 mm; SL 294 mm) to 14,777 eggs (TL 516 mm; SL 414mm), and the relative fecundity ranges from 0.0142 (TL 459 mm) to 0.0015 (TL 393 mm). In mature fish, both the ovaries (left and right lobe) showed a clear asymmetry inside the abdominal cavity (Image 4A).

Gonado-Somatic index and ova diameter

The GSI of females showed three peaks, with the first one in March–April, the second in August–September and a third in December, which indicates an extended batch spawning nature of the fish (Figure 4). Similar to GSI, three peaks could also be observed with regard to the mean ova diameter, confirming an extended spawning season for *P. pardalis* (Figure 5). The maximum ova diameter obtained in the present study was 3.75 mm. While comparing ova diameter frequencies corresponding to different maturity stages, maximum oocyte diameter (mean) was noticed in the ripe stage (Figure 6).

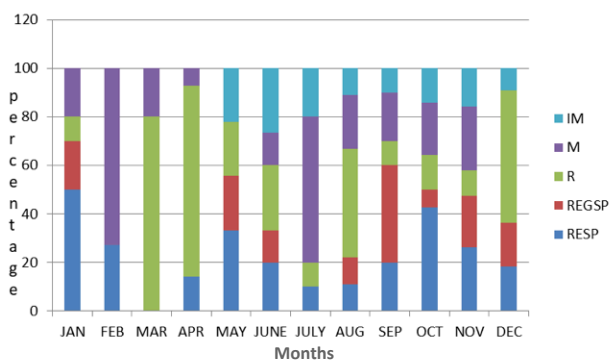


Figure 2. The maturity stages of female *Pterygoplichthys pardalis* in various months under study: IM—Immature | M—Maturing | R—Ripe | REGSP—Regressing spent | RESP—Recovering spent.

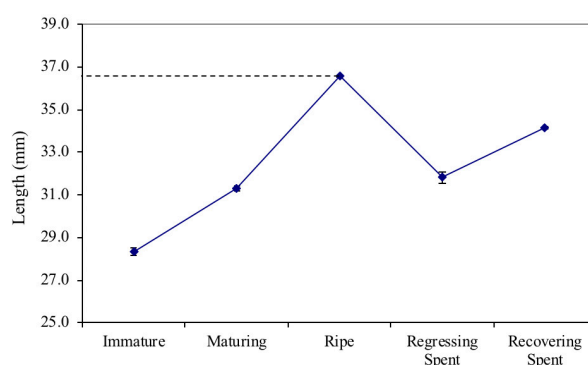


Figure 3. *P. pardalis*: Length at first maturity of female in the present study.

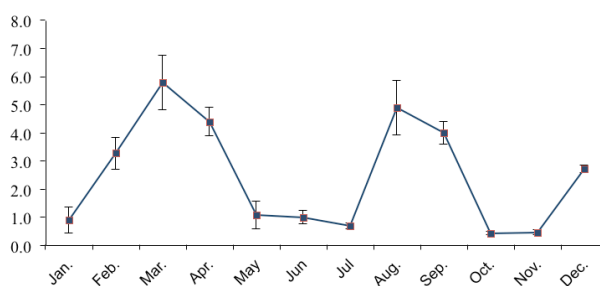


Figure 4. Monthly variations of GSI in *P. pardalis* (female) from the present study. Error bars represent SD.

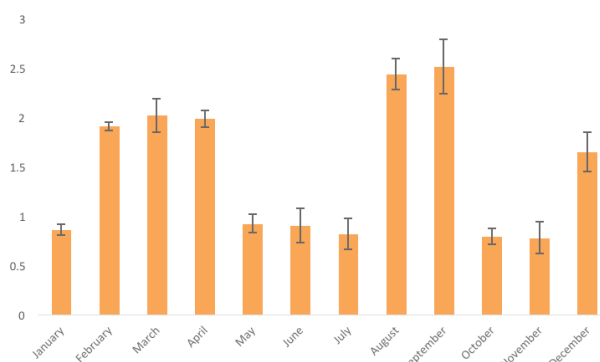


Figure 5. Mean ova diameter (mm) of *P. pardalis* in different months during the present study. Error bars represent SD.



Image 3. Maturity stages of female *Pterygoplichthys pardalis*: A—immature ovary in situ | B—immature ovary | C—maturing ovary in situ | D—maturing ovary. © Smrithy Raj

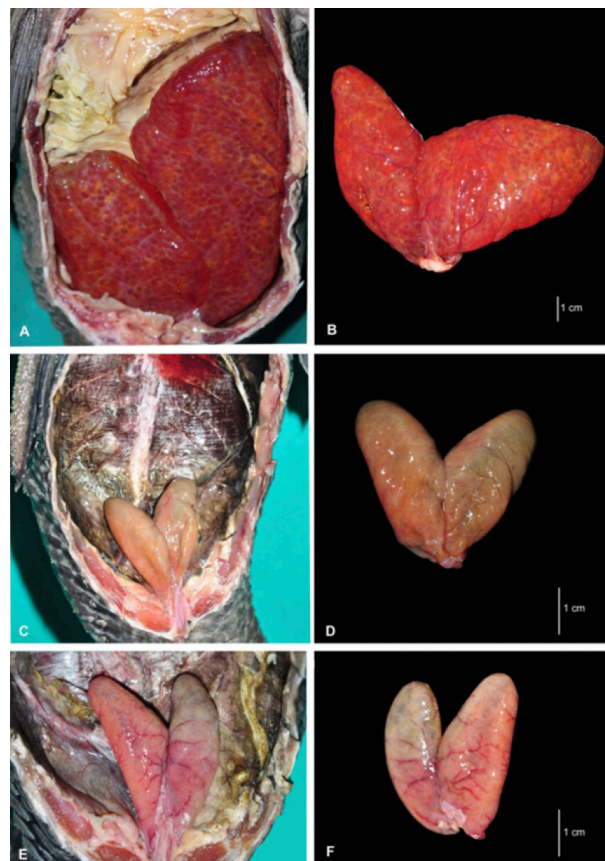


Image 4. Maturity stages of female *Pterygoplichthys pardalis*: A—ripe ovary in situ | B—ripe ovary | C—spent/regressing ovary in situ | D—spent/regressing ovary | E—recovering spent ovary in situ | F—recovering spent ovary. © Smrithy Raj

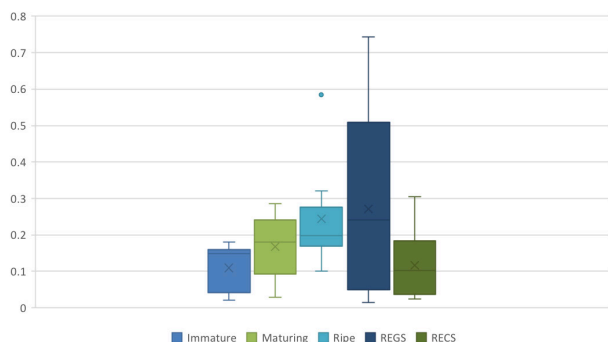


Figure 6. Ova diameter (mm) and maturity stages of *P. pardalis* in the present study. Error bars represent SD.

Regression analysis

Regression analysis showed a significant relationship ($P < 0.001$) between absolute fecundity and the total length, the total body weight and ovary weight (Figures 7–9). As the total length, total body weight, and ovary weight of fish increases, the fecundity does not increase correspondingly due to lesser 'b' value (2.0482, 0.8214, 0.6944).

DISCUSSION

Although the suckermouth armoured catfish of the genus *Pterygoplichthys* is an emerging global invader, the details of its invasion biology are also being studied from its extended invasion ranges (Orfinger & Goodding 2018), the unique breeding behaviour of *Pterygoplichthys* spp. by excavating burrows in river banks have been documented from Florida (Nico et al. 2009), Mexico (Lienart et al. 2013) and the Philippines (Almadin & Jumawan 2016). Similar breeding behaviour was also observed in the present study. The females use the burrows dug by the males to deposit eggs and are guarded by the males till the young ones emerge from the nest; similar behaviour was also noted by Mazzoni et al. (2002), Power (2003), and Liang et al. (2005), which establish that the males of this fish exhibit parental care by building nests, protecting eggs and as well as the juveniles. Lienart et al. (2013) observed egg clutches frequently inside active nests, and such observation was

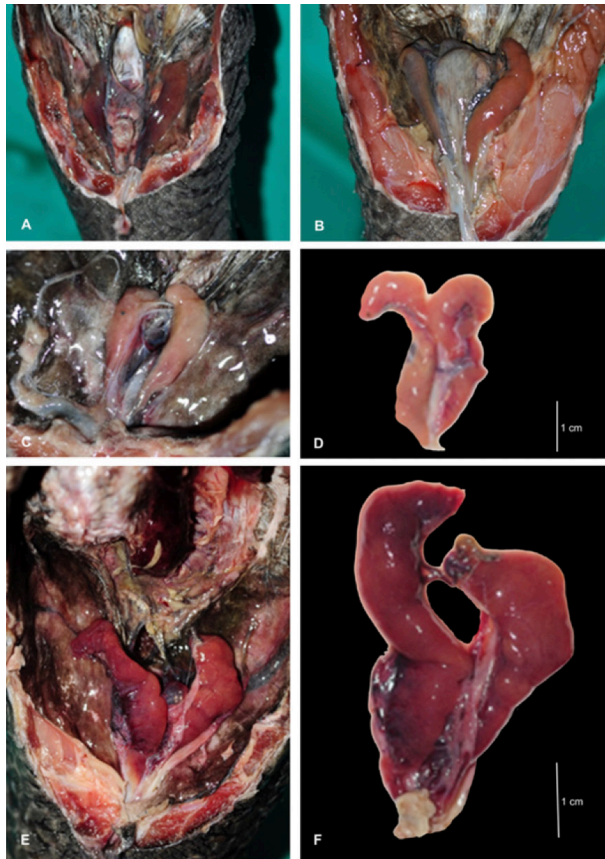


Image 5 A–F. Maturity stages of male *Pterygoplichthys pardalis*: A, B—immature testis | C, D—maturing testis | E, F—mature testis. © Smrithy Raj

not observed in the present study. The present study observes that *Pterygoplichthys pardalis* use the crevices in the granite wall for breeding, which was the first record of breeding behaviour of the species in the Indian water bodies. The benefit of additional natural space also compounded the reproductive behaviour of the fish enhanced its invasive nature in the invaded ecosystem. The sex ratio (M: F) of *P. pardalis* showed an average mean value of 1.04: 1, indicating no bias, showing an equal representation of both sexes in the population.

The current study on the macroscopic and the ova diameter clearly showed different reproductive strategies, showing an extended spawning period with ripe ovaries throughout the year, mostly during March, April, August, and December. Spent ovaries (regressive and recovering ones) and immature stages were noticed in the subsequent months. Such reproductive plasticity was also reported for *P. disjunctivus* by Gibbs et al. (2017) from Volusia Blue Spring for a decade. The peak breeding season reported in *P. pardalis* by Wakida-Kusunoki, & Amador-del Angel (2011) was from June to

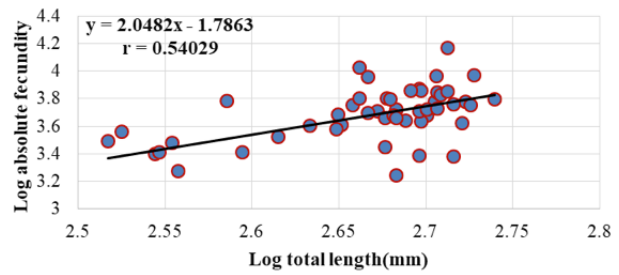


Figure 7. Logarithmic relationship between absolute fecundity and the total length of *P. pardalis* from the present study.

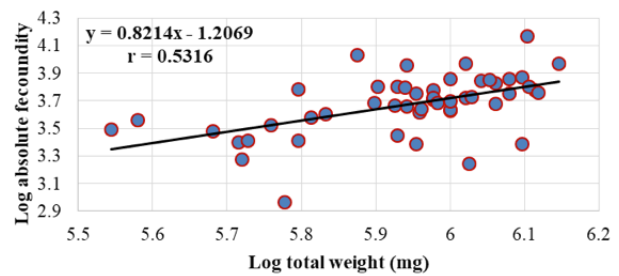


Figure 8. Logarithmic relationship between absolute fecundity and the total weight of *P. pardalis* from the present study.

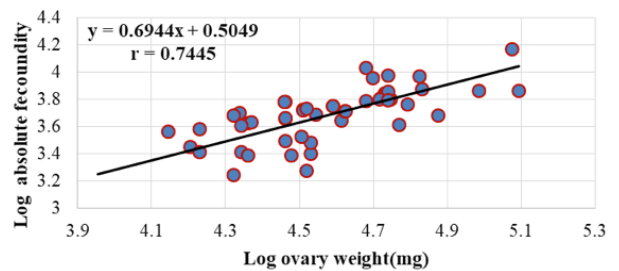


Figure 9. Logarithmic relationship between absolute fecundity and ovary weight of *P. pardalis* from the present study.

September. GSI and ova diameter also showed the same results, which coincided with these months. Fish with ripe gonads were obtained throughout our study period except for February, which is consistent with the studies of Rueda-Jasso et al. (2013) for *P. disjunctivus* from Mexico. In the ripe ovaries itself, we came across oocytes with different diameters, consisting of several immature and maturing ova at the same time, which proves the fish as a determinate batch spawner. Similar to other congeners, *P. pardalis* also spawn in batches (Suzuki et al. 2000; Duarte & Araujo 2002). Studies on *P. disjunctivus* from the Philippines by Jumawan & Herrera (2014) also support this view. Batch spawning in *P. pardalis* and its congeners like *Loricariichthys platymetopon*, *Loricariichthys* sp., and *Loricaria* sp. (Suzuki et al. 2000),

and *Hypostomus affinis* (Duarte & Araujo 2002) have been reported, where the mature ovaries are seen along with immature and maturing ones with pre-vitellogenic eggs. All Loricariids do not spawn in batches, as *Hypostomus ternetzi*, *Megalancistrus aculeatus*, and *Rhinelepis aspera* are total spawners (Suzuki et al. 2000).

Changes in climatic events could disrupt the reproductive process in fishes (Yoneda & Wright 2005; Pankhurst & Munday 2011). Stable water temperature in tropical rivers is considered best instead of fluctuating waters in subtropical rivers and a shift in temperature could confine the spawning period as well (Samat et al. 2016). According to Humphries et al. (1999), flooding is likely the dominant factor in the breeding behaviour of fish. *Pterygoplichthys* may be adapted to take advantage of flooding by initiating reproduction before or at the time of the flood, which allows fry to feed and grow within inundated floodplain habitats (Kramer 1978; Humphries et al. 1999; Lienart et al. 2013). Based on the results of this work, it may be presumed that *P. pardalis* inhabiting the natural drainages without noticeable temperature variations is a batch spawner and temperature may not be an important limiting factor for spawning of fish that live in a habitat with stable or less fluctuating water temperature. The population assessment of *P. pardalis* from natural drainages of Thiruvananthapuram indicated rapid growth, high-performance index and continuous recruitment, which resulted in their successful invasion (Raj et al. 2020). Maximum reproductive activity of *P. disjunctivus* and *L. multiradiatus* were reported from July to September (Liang et al. 2005; Rueda-Jasso et al. 2013).

The size at first maturity of *P. pardalis* in our study was 23.9 cm standard length (TL 33.5 cm). The minimum size at sexual maturity with highly vitellogenic ova in *P. disjunctivus* was reported as 26.7 cm SL onwards by Jumawan & Herrera (2014) from the Philippines. The case of precocious maturation was also reported in smaller females during the peak spawning time. A report of sexual maturity in *P. disjunctivus* by Gibbs et al. (2008) was of 300 mm SL. The length at which 50% of fish gets mature is 36.56 cm. Gonadal development of male and females in the present study also corroborates with the observations of researchers from other parts of the world.

Absolute fecundity of fish ranges from 923 to 14,777 in the present study, and the fecundity reported for *P. pardalis* from Malaysian waters ranged between 1,297 and 18,791 (Samat et al. 2016). A linear relationship is also exhibited between fecundity and TL, TW, and GW of the fish (Bagenal 1978; Mazzoni & Caramaschi 1995;

Duarte & Araujo 2002). The highest degree of correlation was exhibited in the present study between fecundity and the total length of the fish ($r = 0.7445$).

The maximum ova diameter obtained in the present study was 3.75 mm, which was in accordance with the ova diameter of 3.3 mm for *P. pardalis* by Samat et al. (2016), 3.8 mm and 3.6 mm obtained for *P. disjunctivus* by Gibbs et al. (2008) and Jumawan & Herrera (2014) respectively. The largest mean ova diameter was 2.327 for ripe ova, as the ova samples contain vitellogenic oocytes of different sizes from the anterior, middle and posterior regions.

A clear case of asymmetry in the ripe ovary with the left lobe larger than the right in the abdominal cavity was observed in the present study. Similar asymmetry in the mature ovary was also observed in *P. disjunctivus* (Gibbs et al. 2008). This asymmetry may be due to the large area occupied by the intestine and the bulk of fat in the abdominal cavity as reported by Rounsefell (1957) in three female salmonids species.

There are no predators for *P. pardalis* in the natural drainages where they have established. This, coupled with the availability of plenty of detritus in the polluted drainages as food, provide them with a competitive edge over indigenous species, better reproductive strategies, including the ability to breed in burrows along the stream banks and crevices in the granite walls, accessory respiratory organs, batch spawning abilities and parental care assisted *P. pardalis* to colonize the system successfully. The population assessment of *P. pardalis* from the same habitat indicated that rapid growth, high growth performance index, and continuous recruitment are the reasons for their successful invasion, and targeting the young individuals would help in controlling the population of the invasive fish (Raj et al. 2020). We recommend more research on the invasion biology of invasive *P. pardalis*, incorporating long-term studies to fully understand the long-term strategies for their establishment and plasticity in the reproductive behaviour.

REFERENCES

- Almadin, F.J.F. & J.C. Jumawan (2016). Description of breeding burrows of janitor fish (*Pterygoplichthys* spp.) in Agusan River, Butuan City, Philippines. *International Journal of Advances in Chemical Engineering and Biological Sciences* 3(2): 187–190. <https://doi.org/10.15242/IJACEBS.AE0516116187>
- Araújo, F.G., I. Fichberg & S. Duarte (1998). Ciclo reprodutivo de *Loricariichthys spixii* (Steindachner, 1882) (Pisces Loricariidae) na Reservatório de Lajes, RJ. *Acta Biologica Leopoldensia* 20: 309–318.
- Armbruster, J.W. (1998). Modifications of the digestive tract for

- p holding air in loricariid and scoloplacid catfishes.
- Copeia*
- 3: 663–675.
- <https://doi.org/10.2307/1447796>
- Bagenal, T.B. (1978). Aspects of fish fecundity. In: Gerting, S.D. (ed.). *Ecology of freshwater fish production*. Blackwell Scientific, Oxford, 300 pp.
- Bunkley-Williams, L., E.H. Williams Jr., C.G. Lilystrom, I. Corujo-Flores, A.J. Zerbi & C. Aliaume (1994). The South American Sailfin Armored Catfish, *Liposarcus multiradiatus* (Hancock), a new exotic established in Puerto Rican fresh waters. *Caribbean Journal of Science* 30: 90–94.
- Capps, K.A. & A.S. Flecker (2013). Invasive fishes generate biogeochemical hotspots in a nutrient-limited system. *PLoS ONE* 8(1): e54093. <https://doi.org/10.1371/journal.pone.0054093>
- Chavez, J.M., R.M. De La Paz, S.K. Manohar, R.C. Pagulaya & J.R. Carandang VI (2006). New Philippine record of South American sailfin catfishes (Pisces: Loricariidae). *Zootaxa* 1109: 57–68. <https://doi.org/10.11646/zootaxa.1109.1.6>
- Cook-Hildreth, S.L. (2009). Exotic armored catfishes in Texas: Reproductive biology, and effects of foraging on egg survival of native fishes (*Etheostoma fonticola*, Endangered and Diodia diabolic, Threatened). Master in Science Dissertation. Texas State University? San Marcos, 63pp.
- Cook-Hildreth, S.L., T.H. Bonner & D.G. Huffman (2016). Female reproductive biology of an exotic suckermouth armored catfish (Loricariidae) in the San Marcos River, Hyas, Co., Texas, with observations on environmental triggers. *BiolInvasions Records* 5(3): 173–183. <https://doi.org/10.3391/bir.2016.5.3.09>
- Covain, R. & S. Fisch-Muller (2007). The genera of the Neotropical armored catfish subfamily Loricariinae (Siluriformes: Loricariidae): a practical key and synopsis. *Zootaxa* 1462: 1–40. <https://doi.org/10.11646/zootaxa.1462.1.1>
- da Cruz, A.L., H.R. da Silva, L.M. Lundstedt, A.R. Schwantes, G. Moraes, W. Klein & M.N. Fernandes (2013). Air-breathing behavior and physiological responses to hypoxia and air exposure in the airbreathing loricariid fish, *Pterygoplichthys anisitsi*. *Fish Physiology and Biochemistry* 39: 243–256. <https://doi.org/10.1007/s10695-012-9695-0>
- Duarte, S. & F.G. Araújo (2002). Fecundity of the *Hypostomus affinis* (Siluriformes, Loricariidae) in the Lajes Reservoir, Rio de Janeiro, Brazil. *Revista de Biologia Tropical* 50(1): 193–197.
- Feiner, Z.S., D.D. Aday & J.A. Rice (2012). Phenotypic shifts in white perch life history strategy across stages of invasion. *Biological Invasions* 14: 2315–2329. <https://doi.org/10.1007/s10530-012-0231-z>
- German, D.P., D.T. Neuberger, M.N. Callahan, N.R. Lizardo & D.H. Evans (2010). Feast to famine: the effects of food quality and quantity on the gut structure and function of a detritivorous catfish (Teleostei: Loricariidae). *Comparative Biochemistry and Physiology* 155: 281–293.
- Gibbs, M.A., J.H. Shields, D.W. Lock, K.M. Talmadge & T.M. Farrell (2008). Reproduction in an invasive exotic catfish *Pterygoplichthys disjunctivus* in Volusia Blue Spring, Florida, U.S.A. *Journal of Fish Biology* 73: 1562–1572. <https://doi.org/10.1111/j.1095-8649.2008.02031.x>
- Gibbs, M., P. Watson, K. Johnson-Sapp & C. Lind (2017). Reproduction revisited—a decade of changes in the reproductive strategies of an invasive catfish, *Pterygoplichthys disjunctivus* (Weber, 1991), in Volusia Blue Spring, Florida. *Aquatic Invasions* 12(2): 225–239. <https://doi.org/10.3391/ai.2017.12.2.10>
- Gutkowsky, L.F.G. & M.G. Fox (2012). Intra-population variability of lifehistory traits and growth during range expansion of the invasive round goby, *Neogobius melanostomus*. *Fisheries Management and Ecology* 19: 78–88. <https://doi.org/10.1111/j.1365-2400.2011.00831.x>
- Hoosain, M.Y., M.M. Rahman, Z.F. Ahmed, J. Ohtomi & A.B.M.S. Islam (2008). First record of South America sailfin catfish *Pterygoplichthys multiradiatus* in Bangladesh. *Journal of Applied Ichthyology* 24: 718–720.
- Hoover, J.J., K.J. Killgore & A.F. Cofrancesco (2004). Suckermouth catfishes: threat to aquatic ecosystem of the United States? *Aquatic Nuisance Species Research Program Bulletin* 4: 1–8.
- Horkova, K. & V. Kovac (2015). Ontogenetic phenomena, temporal aspect, and ecological factors in the successful invasion of round goby *Neogobius melanostomus* in the River Danube. *Aquatic Invasions* 10: 227–235. <https://doi.org/10.3391/ai.2015.10.2.11>
- Humphries, P., A.J. King & J.D. Koehn (1999). Fish, flows and flood plains: links between freshwater fishes and their environment in the Murray-Darling River system, Australia. *Environmental Biology of Fishes* 56: 129–151.
- Hussan, A., R.N. Mandal, F. Hoque, A. Das, P.P. Chakrabarti & Adhikari (2018). Dominance of intergrades of invasive suckermouth armored catfishes *Pterygoplichthys* spp. (Siluriformes: Loricariidae) in coastal wetlands of West Bengal, India. *Journal of Indian Society of Coastal Agricultural Research* 36(1): 84–92.
- Jumawan, J.C. & A.A. Herrera (2014). Ovary Morphology and Reproductive Features of the Female Suckermouth Sailfin Catfish, *Pterygoplichthys disjunctivus* (Weber 1991) from Marikina River, Philippines. *Asian Fisheries Science* 27: 75–89.
- Knight, J.D.M. (2010). Invasive ornamental fish: a potential threat to aquatic biodiversity in peninsular India. *Journal of Threatened Taxa* 2(2): 700–704. <https://doi.org/10.11609/JoTT.o2179.700-4>
- Kopp, D., J. Syväranta, J. Figuerola, A. Compín, F. Santoul & R. Céréghino (2009). Environmental effects related to the local absence of exotic fish. *Biological Conservation* 142: 3207–3212. <https://doi.org/10.1016/j.biocon.2009.07.030>
- Kramer, D.L. (1978). Reproductive seasonality in the fishes of a tropical stream. *Ecology* 59: 976–985.
- Krishnakumar, K., R. Raghavan, G. Prasad, A.B. Kumar, M. Sekharan, B. Pereira & A. Ali (2009). When pets become pests—exotic aquarium fishes and biological invasions in Kerala, India. *Current Science* 97: 474–476.
- Kumar, A.B., S. Raj, U. Sureshkumar & S. George (2015). Invasion of South American suckermouth armored catfishes *Pterygoplichthys* spp. (Loricariidae) in Kerala, India - a case study. *Journal of Threatened Taxa* 7(3): 6987–6995. <https://doi.org/10.11609/JoTT.o4133.6987-95>
- Kumar, A.B., P.J. Schofield, S. Raj & S. Sima (2018). Salinity tolerance of non-native suckermouth armored catfish (Loricariidae: *Pterygoplichthys* sp.) from Kerala, India. *Management of Biological Invasions* 9(1): 49–57. <https://doi.org/10.3391/mbi.2018.9.1.05>
- Liang, S.H., H.P.L. Wu & B.S. Shied (2005). Size structure, reproductive phenology and sex ratio of an exotic sailfin catfish (*Liposarcus multiradiatus*) in the Kaoping river of southern Taiwan. *Zoological Studies* 44: 252–259.
- Lienart, G.D.H., R. Rodiles-Hernandez & K.A. Capps (2013). Nesting burrows and behaviour of nonnative catfishes (Siluriformes: Loricariidae) in the Usumacinta-Grijalva watershed, Mexico. *The Southwestern Naturalist* 58(2): 238–243.
- Lowerre-Barbieri, S.K., N.J. Brown-Peterson, H. Murua, J. Tomkiewicz, D.M. Wyanski & F. Saborido-Rey (2011). Emerging issues and methodological advances in fisheries reproductive biology. *Marine and Coastal Fisheries* 3(1): 32–51. <https://doi.org/10.1080/19425120.2011.555725>
- Mazzoni, R. & E.P. Caramaschi (1995). Size, structure, sex ratio and onset of sexual maturity of two species of *Hypostomus*. *Journal of Fish Biology* 47: 841–849.
- Mazzoni, R. & E.P. Caramaschi (1997). Spawning season, ovarian development and fecundity of *Hypostomus affinis* (Osteichthyes, Loricariidae). *Revista Brasileira de Biologia* 57: 455–462.
- Mazzoni, R., E.P. Caramaschi & N. Fenerich-Verani (2002). Reproductive biology of a characidiinae (Osteichthyes, Characidae) from the Ubatiba River, Marica-RJ. *Brazilian Journal of Biology* 62(3): 487–494.
- Mendoza-Alfaro, R.E., B. Cudmore, R. Orr, J.P. Fisher, S. Contreras-Balderas, W.R. Courtenay, P.K. Osorio, N. Mandrak, P. Alvarez-Torres, M. Arroyo-Damian, C. Escalera-Gallardo, A. Guevara-Sanguines, G. Greene, D. Lee, A. Orbe-Mendoza, C. Ramirez-Martinez & O. Strabridis-Arana (2009). Trinational risk assessment



- guidelines for aquatic alien invasive species: test cases for the snakeheads (Channidae) and armored catfishes (Loricariidae) in North American inland waters. Commission for Environmental Cooperation, Montreal, Canada, 98pp.
- Nico, L.G. & R.L. Martin (2001). The South American suckermouth armored catfish introductions in the American Southwest. *The Southwestern Naturalist* 46: 98–104.
- Nico, L.G., H.L. Jelks & T. Tuten (2009). Non-native suckermouth armored catfishes in Florida: description of nest burrows and burrow colonies with assessment of shoreline conditions. *ANSRP Bulletin* 9: 1–30.
- Nico, L.G., P.L. Butt, G.R. Johnson, H.L. Jelks, M. Kail & S.J. Walsh (2012). Discovery of the South American Suckermouth Armoured Catfish (Loricariidae, *Pterygoplichthys* spp.) in the Santa Fe River drainage, Suwannee River basin, USA. *Bioinvasion Records* 1: 179–200. <https://doi.org/10.3391/bir.2012.1.3.04>
- Orfinger, A.B. & D.D. Goodding (2018). The Global Invasion of the Suckermouth Armored Catfish Genus *Pterygoplichthys* (Siluriformes: Loricariidae): Annotated List of Species, Distributional Summary, and Assessment of Impacts. *Zoological Studies* 57:7 (2018), 1–16. <https://doi.org/10.6620/ZS.2018.57-07>
- Pankhurst, N.W. & P.L. Munday (2011). Effects of climate change on fish reproduction and early life history stages. *Marine and Freshwater Research* 62(9): 1015–1026
- Parente, T.E., D.A. Moreira, M.G.P. Magalhaes, P.C.C. de Andrade, C. Furtado, B.J. Haas, J.J. Stegeman & M.E. Hahn (2017). The liver transcriptome of suckermouth armoured catfish (*Pterygoplichthys anisitsi*, Loricariidae): identification of expansions in defense gene families. *Marine Pollution Bulletin* 115: 352–361. <https://doi.org/10.1016/j.marpolbul.2016.12.012>
- Patino, R. & C.V. Sullivan (2002). Ovarian follicle growth, maturation, and ovulation in teleost fish. *Fish Physiology and Biochemistry* 26: 57–70.
- Power, M.E. (2003). Life cycles, limiting factors, and behavioural ecology of four loricariid catfishes in a Panamanian River. In: Arratia, G., B.G. Kapoor, M. Chardon & R. Dlogo (eds.). *Catfishes*. Science Publishers, Inc., Enfield, NH.
- Radhakrishnan, R.C., K. Roshni & M. K. Balakrishna (2020). Reproductive biology of the endemic cyprinid fish *Hypselobarbus thomassi* (Day, 1874) from Kallada River in the Western Ghats, India. *Journal of Applied Ichthyology* 2020;00: 1–9. <https://doi.org/10.1111/jai.14064>
- Raj, S., A.B. Kumar, R. Raghavan & N. Dahanukar (2020). Amazonian invaders in an Asian biodiversity hotspot: Understanding demographics for the management of the armoured sailfin catfish, *Pterygoplichthys pardalis* in Kerala, India. *Journal of Fish Biology* 96: 549–553. <https://doi.org/10.1111/jfb.14243>
- Rounsefell, G.A. (1957). Fecundity of the North American Salmonidae. *Fisheries Bulletin* 57: 451–468.
- Rueda-Jasso, R.A., A. Campos-Mendoza F. Arreguín-Sánchez, E. Díaz- Pardo & C.A. Martínez-Palacios (2013). The biological and reproductive parameters of the invasive armoured catfish *Pterygoplichthys disjunctivus* from Adolfo López Mateos El Infiernillo Reservoir, Michoacán-Guerrero, Mexico. *Revista Mexicana de Biodiversidad* 84: 318–326. <https://doi.org/10.7550/rmb.26091>
- Samat, A., F.M. Yusoff, A. Arshad, M.A. Ghaffar, S.M. Nor, A.L.B. Magalhaes & S.K. Das (2016). Reproductive biology of the introduced sailfin catfish *Pterygoplichthys pardalis* (Pisces: Loricariidae) in Peninsular Malaysia. *Indian Journal of Fisheries* 63(1): 35–41. <https://doi.org/10.21077/ijf.2016.63.1.44937-05>
- Sandilyan, S (2016). Occurrence of ornamental fishes: a looming danger for inland fish diversity of India. *Current Science* 110: 2099–2104.
- Singh, A.K. (2014). Emerging alien species in Indian aquaculture: prospects and threats. *Journal of Aquatic Biology & Fisheries* 2: 32–41.
- Sinha, R.K., R.K. Sinha, U.K. Sarkar & W.S. Lakra (2010). First record of the southern sailfin catfish, *Pterygoplichthys anisitsi* Eigenmann and Kennedy, 1903 (Teleostei: Loricariidae), in India. *Journal of Applied Ichthyology* 26: 606–608.
- Soundararajan, N., M.R. Raj, N. Kamaladhasan, I.R. Saidanyan & S. Chandrasekaran (2015). On-line trade of aesthetic exotic organisms: sword of Damocles? *Current Science* 109: 1404–1410.
- Suzuki, H.I., A.A. Agostinho & K.O. Winemiller (2000). Relationship between oocyte morphology and reproductive strategy in loricariid catfishes of the Parana River, Brazil. *Journal of Fish Biology* 57: 791–807. <https://doi.org/10.1006/jfbi.2000.1352>
- Vazzoler, A.E. (1996). *Biologia da reprodução de peixes teleosteos: Teoria e prática*. Maringá, Nupelia, 169 pp.
- Wakida-Kusunoki, A.T., R. Ruiz-Carus & Y.L.E Amador-del Angel (2007). Amazon sailfin catfish *Pterygoplichthys pardalis* (Castelnau, 1855) (Loricariidae), another exotic species established in Southeastern Mexico. *The Southwestern Naturalist* 52: 141–144.
- Wakida-Kusunoki, A.T & E. Amador-del Angel (2011). Aspectos biológicos del pleco invasor *Pterygoplichthys pardalis* (Teleostei: Loricariidae) en el río Palizada, Campeche, México. *Revista Mexicana de Biodiversidad* 82: 870–878.
- Wei, H., G.H. Copp, L. Vilizzi, F. Liu, D. Gu, D. Luo, M. Xu, X. Mu & Y. Hu (2017). The distribution, establishment and life-history traits of non-native sailfin catfishes *Pterygoplichthys* spp. in the Guangdong Province of China. *Aquatic Invasions* 12: 241–249. <https://doi.org/10.3391/ai.2017.12.2.11>
- Welcomme, R. & C. Vidithayanom (2003). The impacts of introductions and stocking of exotic species in the Mekong Basin and policies for their control. MRC Technical Paper No. 9, Mekong River Commission, Phnom Penh, 35pp.
- Yoneda, M. & P.J. Wright (2005). Effect of temperature and food availability on reproductive investment of first-time spawning male Atlantic cod, *Gadus morhua*. *ICES Journal of Marine Science* 62(7): 1387–1393. <https://doi.org/10.1016/j.icesjms.2005.04.018>
- Zahorska, E., K. Svolikova & V. Kovac (2013). Do invasive populations of topmouth gudgeon (*Pseudorasbora parva*, Temminck and Schlegel) from disturbed and undisturbed habitats follow different life-histories? *International Reviews in Hydrobiology* 98: 61–70. <https://doi.org/10.1002/iroh.201201446>

Author details: Smrithy Raj (SR) is a PhD candidate in the Department of Aquatic Biology & Fisheries, University of Kerala. He works on invasive fishes, invasion biology and taxonomy of freshwater crabs. Suvarna Devi (SD) works as guest faculty in the Department of Aquatic Biology & Fisheries, University of Kerala and she is specialised in fish biology and taxonomy of brachyuran crabs. Amrutha Joy (AJ) is an M.Phil. student in the Department of Aquatic Biology & Fisheries, University of Kerala. Biju Kumar (ABK) is the professor and head in the Department of Aquatic Biology & Fisheries, University of Kerala. His research interests include taxonomy of aquatic organisms, invasion biology, fisheries management and conservation policies.

Author contributions: SR—field work, data collection, photography, drafting manuscript, revisions at different stages. SD—methodology design, data analysis and interpretation, write up, review, revisions at different stages. AJ—data collection, practical biology studies, writing of preliminary draft. ABK—conceptualization, funding acquisition, photography, manuscript review, editing.

Acknowledgements: The authors thank the Directorate of Environment and Climate Change, Government of Kerala, for the financial support of the project on aquatic exotic species in Kerala. We are grateful to Madhu Vellayani, the fisherman who was our close companion and helped us in fish collection in all field surveys. We thank the support of Mr Mosab Ali Mohamed Ali Al-zahaby for his help in the regression analyses. SR is funded for his PhD programme by the Rajiv Gandhi National Fellowship (RGNF), Government of India





www.threatenedtaxa.org

OPEN ACCESS



The Journal of Threatened Taxa (JoTT) is dedicated to building evidence for conservation globally by publishing peer-reviewed articles online every month at a reasonably rapid rate at www.threatenedtaxa.org. All articles published in JoTT are registered under [Creative Commons Attribution 4.0 International License](https://creativecommons.org/licenses/by/4.0/) unless otherwise mentioned. JoTT allows unrestricted use, reproduction, and distribution of articles in any medium by providing adequate credit to the author(s) and the source of publication.

ISSN 0974-7907 (Online) | ISSN 0974-7893 (Print)

August 2021 | Vol. 13 | No. 9 | Pages: 19191–19390

Date of Publication: 26 August 2021 (Online & Print)

DOI: 10.11609/jott.2021.13.9.19191-19390

Articles

On the impact of earthquake-induced landslides on Red Panda *Ailurus fulgens* (Mammalia: Carnivora: Ailuridae) habitat in Langtang National Park, Nepal
– Yogesh Rana Magar, Man Kumar Dhamala, Ajay Mathema, Raju Chauhan & Sijar Bhatta, Pp. 19191–19202

Rhesus Macaque *Macaca mulatta* (Mammalia: Primates: Cercopithecidae) in a human-modified landscape: population, activity budget, and societal perceptions in Bangladesh
– Sufia Akter Neha, Mohammad Ashraf Ul Hasan, Mohammad Abdul Baki & Subrina Sehrin, Pp. 19203–19211

Factors affecting the species richness and composition of bird species in a community managed forest of Nepal
– Bishow Poudel, Bijaya Neupane, Rajeev Joshi, Thakur Silwal, Nirjala Raut & Dol Raj Thanet, Pp. 19212–19222

Communications

A large mammal survey in Koyli Alpha Community Wildlife Reserve and its surroundings in the Great Green Wall extension area in Senegal
– Anna Niang & Papa Ibnou Ndiaye, Pp. 19223–19231

Blackbuck *Antelope cervicapra* (Mammalia: Cetartiodactyla: Bovidae) estimates in human-dominated landscape in Aligarh, Uttar Pradesh, India
– Mujahid Ahamad, Jamal A. Khan & Satish Kumar, Pp. 19232–19238

Diet of Leopards *Panthera pardus fusca* inhabiting protected areas and human-dominated landscapes in Goa, India
– Bipin S. Phal Desai, Avelyno D'Costa, M.K. Praveen Kumar & S.K. Shyama, Pp. 19239–19245

First record of interspecies grooming between Raffles' Banded Langur and Long-tailed Macaque
– Zan Hui Lee, Andie Ang & Nadine Ruppert, Pp. 19246–19253

Photographic evidence of Red Panda *Ailurus fulgens* Cuvier, 1825 from West Kameng and Shi-Yomi districts of Arunachal Pradesh, India
– Moktan Megha, Sylvia Christi, Rajesh Gopal, Mohnish Kapoor & Ridhima Solanki, Pp. 19254–19262

On the reproductive biology of the invasive Armoured Sailfin Catfish *Pterygoplichthys pardalis* (Castelnau, 1855) (Siluriformes: Loricariidae) from the natural drainages in Thiruvananthapuram, India
– Smrithy Raj, Suvarna S. Devi, Amrutha Joy & A. Biju Kumar, Pp. 19263–19273

On the high bird diversity in the non-protected regions of Trashiyangtse District in Bhutan
– Lam Norbu, Phuntsho Thinley, Tandin Wangchuck, Ugyen Dechen, Lekey Dorji, Tshering Choephel & Pasang Dorji, Pp. 19274–19292

Population status and distribution of the Critically Endangered Bengal Florican *Houbaropsis bengalensis* in the grassland of Koshi Tappu Wildlife Reserve, Nepal
– Ritika Prasai, Hemanta Kafley, Suraj Upadhaya, Swosthi Thapa, Pratistha Shrestha, Alex Dudley & Yajna Prasad Timilsina, Pp. 19293–19301

Is habitat heterogeneity effective for conservation of butterflies in urban landscapes of Delhi, India?
– Monalisa Paul & Aisha Sultana, Pp. 19302–19309

A preliminary checklist of moths (Lepidoptera: Heterocera) from Gangajalghati, Bankura, West Bengal, India
– Ananya Nayak, Pp. 19310–19323

First report of three species of the genus *Diaphanosoma* (Crustacea: Cladocera: Sididae) from Jammu waters (J&K), India
– Nidhi Sharma & Sarbjeet Kour, Pp. 19324–19337

Review

Wild ungulates in Jordan: past, present, and forthcoming opportunities
– Ehab Eid & David Mallon, Pp. 19338–19351

Viewpoint

The captive population of the Lion-tailed Macaque *Macaca silenus* (Linnaeus, 1758). The future of an endangered primate under human care
– Nilofer Begum, Werner Kaumanns, Alexander Sliwa & Mewa Singh, Pp. 19352–19357

Short Communication

Jaguar *Panthera onca* (Linnaeus, 1758) (Mammalia: Carnivora: Felidae) presumably feeding on Flathead Catfish *Pylodictis olivaris* (Rafinesque, 1818) (Actinopterygii: Siluriformes: Ictaluridae) at Aros and Yaqui rivers, Sonora, Mexico
– Juan Pablo Gallo-Reynoso, Pp. 19358–19362

Notes

Life near a city: activity pattern of Golden Jackal *Canis aureus* Linnaeus, 1758 (Mammalia: Carnivora: Canidae) in a habitat adjoining Bhubaneswar, India
– Subrat Debata, Pp. 19363–19366

Chemical immobilisation of a Eurasian Lynx *Lynx lynx* (Linnaeus, 1758) (Mammalia: Carnivora: Felidae) with ketamine-dexmedetomidine mixture in Ladakh, India
– Animesh Talukdar & Pankaj Raina, Pp. 19367–19369

White-bellied Heron *Ardea insignis* in Hkakabo Razi Landscape, northern Myanmar
– Myint Kyaw, Paul J.J. Bates, Marcela Suarez-Rubio, Bran Shaung, Han Nyi Zaw, Thein Aung, Sai Sein Lin Oo & Swen C. Renner, Pp. 19370–19372

Range extension of the Common Slug Snake *Pareas monticola* (Cantor, 1839) (Reptilia: Squamata: Pareidae): a new family record for Nepal
– Dipa Rai, Manoj Pokharel & Tapil P. Rai, Pp. 19373–19375

First record of *Mantispilla indica* (Westwood, 1852) (Neuroptera: Mantispidae) from the Western Ghats, India
– T.B. Suryanarayanan & C. Bijoy, Pp. 19376–19379

A new distribution record of the Western Ghats endemic damselfly *Melanoneura bilineata* Fraser, 1922 (Insecta: Odonata) from Maharashtra, India
– Yogesh Koli & Akshay Dalvi, Pp. 19380–19382

A new record of the Emerald Striped Spreadwing *Lestes viridulus* Rambur, 1842 (Zygoptera: Lestidae) from Nepal
– Manoj Sharma, Pp. 19383–19385

Rediscovery of the Bhutan Primrose *Primula jigmediana* W.W. Smith (Angiosperms: Primulaceae) after 87 years in Bumdeling Wildlife Sanctuary, Bhutan
– Tez B. Ghalley, Tshering Dendup, Karma Sangay & Namgay Shacha, Pp. 19386–19388

First report of *Golovinomyces* sp. causing powdery mildew infection on *Dyschoriste nagchana* in Western Ghats of India
– Sachin Vasantrao Thite, Pp. 19389–19390

Publisher & Host

