

The Journal of Threatened Taxa (JoTT) is dedicated to building evidence for conservation globally by publishing peer-reviewed articles online every month at a reasonably rapid rate at www.threatenedtaxa.org. All articles published in JoTT are registered under Creative Commons Attribution 4.0 International License unless otherwise mentioned. JoTT allows unrestricted use, reproduction, and distribution of articles in any medium by providing adequate credit to the author(s) and the source of publication.

Journal of Threatened Taxa

Building evidence for conservation globally

www.threatenedtaxa.org ISSN 0974-7907 (Online) | ISSN 0974-7893 (Print)

COMMUNICATION

SERO-DIAGNOSIS OF TUBERCULOSIS IN ELEPHANTS IN MAHARASHTRA, INDIA

Utkarsh Rajhans, Gayatri Wankhede, Balaji Ambore , Sandeep Chaudhari, Navnath Nighot, Vitthal Dhaygude & Chhaya Sonekar

26 June 2021 | Vol. 13 | No. 7 | Pages: 18713–18718 DOI: 10.11609/jott.5502.13.7.18713-18718





For Focus, Scope, Aims, and Policies, visit https://threatenedtaxa.org/index.php/JoTT/aims_scope For Article Submission Guidelines, visit https://threatenedtaxa.org/index.php/JoTT/about/submissions For Policies against Scientific Misconduct, visit https://threatenedtaxa.org/index.php/JoTT/policies_various For reprints, contact <ravi@threatenedtaxa.org>

The opinions expressed by the authors do not reflect the views of the Journal of Threatened Taxa, Wildlife Information Liaison Development Society, Zoo Outreach Organization, or any of the partners. The journal, the publisher, the host, and the partners are not responsible for the accuracy of the political boundaries shown in the maps by the authors.

Publisher & Host



Journal of Threatened Taxa | www.threatenedtaxa.org | 26 June 2021 | 13(7): 18713-18718

ISSN 0974-7907 (Online) | ISSN 0974-7893 (Print)

https://doi.org/10.11609/jott.5502.13.7.18713-18718

#5502 | Received 28 October 2019 | Final received 10 April 2021 | Finally accepted 27 April 2021

Sero-diagnosis of tuberculosis in elephants in Maharashtra, India

Utkarsh Rajhans 10, Gayatri Wankhede 20, Balaji Ambore 30, Sandeep Chaudhari 40, Navnath Nighot 50, Vitthal Dhaygude 6 k Chhaya Sonekar 7

^{1,2,3} Department of Veterinary Clinical Medicine Ethics and Jurisprudence, ⁶ Department of Veterinary Pathology, Krantisinh Nana Patil College of Veterinary Science (KNPCVS), Shirwal, Satara District, Maharashtra 412801, India. ^{4,7} Department of Veterinary Public Health and Epidemiology, Nagpur Veterinary College, Maharashtra Animal and Fisheries Sciences UniveristY, Maharashtra 440006, India.

⁵ Rajiv Gandhi Zoological Park and Wildlife Research Centre, Katraj, Pune, Maharashtra 411046, India.

¹utkarshrajhans01@gmail.com (corresponding author), ² gayatriwd87@gmail.com, ³balajiavet@gmail.com, ⁴vphsandeep@gmail.com, ⁵drnknighot@rediffmail.com, ⁶drvitthalp@gmail.com, ⁷chhayasonekar178@gmail.com

Abstract: Tuberculosis is a highly contagious zoonotic disease caused by Mycobacterium spp. A study was conducted to detect the presence of Mycobacterium in captive elephants. A total of 15 captive elephants were screened from various regions in Maharashtra. The blood and serum samples collected were subjected to rapid test kit, BacT/ALERT 3D system, Ziehl-Neelsen (ZN) staining and PCR. All the samples were found seronegative using rapid test kit and whole blood PCR. Whereas, all samples were signalled culture positive in BacT/ ALERT 3D system which were further subjected to PCR, only one amplicon was produced of 176bp of RD4 gene (Mycobacterium bovis) and no acid-fast organism was detected upon ZN. Due to the atypical nature of this organism, diagnosis of this disease in elephants using various tests is complicated unlike the diagnostic tests that are validated in domestic animals. Therefore, many tests have sub-optimal sensitivity and specificity in elephants. As TB is a zoonotic disease, transmission can occur between human-livestock-elephants interface. Therefore, the zoos and state forest authority should inculcate a protocol of periodic TB screening for Mahouts and elephants in captivity along with protocol of elephant-visitor interaction, thus helping in conservation of this endangered species in India.

Keywords: Elephants, mycobacterium, serodiagnosis, Tuberculosis.

Editor: Bahar S. Baviskar, Wild-CER, Nagpur, India.

Date of publication: 26 June 2021 (online & print)

Citation: Rajhans, U., G. Wankhede, B. Ambore, S. Chaudhari, N. Nighot, V. Dhaygude & C. Sonekar (2021). Sero-diagnosis of tuberculosis in elephants in Maharashtra, India. Journal of Threatened Taxa 13(7): 18713–18718. https://doi.org/10.11609/jott.5502.13.7.18713-18718

Copyright: © Raihans et al. 2021. Creative Commons Attribution 4.0 International License. JoTT allows unrestricted use, reproduction, and distribution of this article in any medium by providing adequate credit to the author(s) and the source of publication.

Funding: (1) Department of Veterinary Public Health and Epidemiology, Nagpur Veterinary College, Maharashtra Animal and Fisheries Sciences University, India; (2) Department of Veterinary Medicine, Ethic and Jurisprudence, Department of Veterinary Pathology, Krantisinh Nana Patil College of Veterinary Science, Shirwal, Satara District, Maharashtra, Inida [Project Number: 10(14)/2014-EP & HS].

Competing interests: The authors declare no competing interests.

For Author details & Author contributions see end of this article.

Acknowledgements: This research work was possible due to the permission granted by central zoo authority (CZA), New Delhi and principal chief conservator of forests (PCCF), Maharashtra. Dr. G.D. Wankhede, assistant professor and Dr. B.N. Ambore, assistant professor and sectional head, Department of Veterinary Clinical of Medicine, Ethics and Jurisprudence, Krantisinh Nana Patil College of Veterinary Science, Shirwal or her valuable guidance moral support and constant encouragement during the entire course of studies. Dr. S.P. Chaudhari Professor and Head, Department of Veterinary Public Health and Epidemiology, Nagpur Veterinary College, Nagpur for his expertise to formulate the research methodology in particular and for providing chance to work at his departmental laboratory facility. I wish to record special thanks to Dr. N.K. Nighot, Deputy Director, Rajiv Gandhi Zoological Park and Wildlife Research Centre, Pune whose proficiency was invaluable in formulating the research topic and his timely guidance. This work would not have been possible without the support of my committee member Dr. V.S. Dhaygude.





INTRODUCTION

Elephants are the largest terrestrial mammals on the earth. Elephants belong to the family Elephantidae in animal kingdom. Two genera *Elephas* and *Loxodonta* and three species are present today – the Asian Elephant *Elephas maximus*, the African Bush Elephant *Loxodonta africana*, and the African Forest Elephant *Loxodonta cyclotis*.

Currently, a population of 27,312 elephants has been estimated from 23 states in India (Project Elephant Division, Government of India, 2017). In past decades, the population of elephants has drastically been reduced and since 1986, the Asian Elephant has been listed as 'Endangered' species on the IUCN Red List, as the wild population has declined by at least 50% (Choudhury et al. 2008). The Asian Elephant is placed in Schedule I and Part I of Indian Wildlife Protection Act (1972); conferring it the highest level of protection.

Tuberculosis is a highly contagious zoonotic disease in animals as well as humans. It is caused by highly pathogenic bacteria of Mycobacterium tuberculosis complex (MTBC) which are M. tuberculosis, M. bovis, and M. canetti. The M. tuberculosis and M. bovis are most pathogenic. Tuberculosis (TB) in elephants was first observed more than 2,000 years ago by ancient Ayurvedic physicians in Ceylon (Iyer 1937; McGaughey Transmission between human and captive 1961). animals has occurred following close and frequent contact (Kathleen et al. 2002). More frequent reporting of this disease occurs in Asian Elephants than in African Elephants may be due to closer human contact related to their use for performances, rides and in temple rituals. Mycobacterium tuberculosis is the predominating disease-causing agent in elephants, although TB cases have been caused by M. bovis (Mikota 2008). The reservoirs for M. tuberculosis and M. bovis are infected human and cattle (Hirsch 2004).

Elephants with tuberculosis infection show clinical signs like weight loss, wasting and weakness, coughing or dyspnoea have been reported but appear to be uncommon. Exercise intolerance may be observed in working elephants (Mikota 2008). In some cases, ventral oedema has been reported, but other pathologic factors could be the initiating cause (Seneviratna et al. 1966). Majority of times elephants infected with TB do not have any clinical signs. In some cases, elephants manifest symptoms only in advance stage of disease or may not be diagnosed until necropsy (Paudel & Tsubota 2016).

The study presents the clinical, serological, and culture data from 15 elephants present in captivity thus

helping to diagnose and decrease TB risk to these wild

MATERIALS AND METHODS

animals.

Study animals and sample collection

Blood and serum samples were collected from the 15 elephants in captivity of Forest Camp areas of Gadchiroli (19.4290° N, 80.0563° E), Pune Zoo (18.452°N, 73.865°E), Mumbai Zoo (18.978°N, 72.835°E), Shegaon temple (20.789°N, 76.701°E) in Maharashtra. The elephants were included in the study irrespective of their health status, age, sex or habitat.

Serological testing

The Wild TB alert kit is a lateral flow chromatographic immunoassay for the detection of antibodies of mycobacterium tuberculosis complex antigenserum, plasma and whole blood of elephants. This kit contents a unique cocktail of tuberculosis specific recombinant proteins (ESAT-6, CFP-10, MPB83, MPB70) and crude protein impregnated on nitrocellulose membrane housed in a disposable plastic cassette. After adding sample to the well followed by addition of diluent they travel through the membrane by capillary action. If antibodies are present, they bind to the antigen and a red colour band is observed in test area.

BacT/ALERT 3D system

BacT/ALERT 3D system is an automated microbial detection system which offers microbiological culture of blood. This mycobacteria detection systems utilize a colorimetric sensor and reflected light to monitor the presence and production of carbon dioxide (CO₂) dissolved in the culture medium. BacT/ALERT MB are disposable culture bottles with a removable closure contain 10 ml of media and an internal sensor that detects carbon dioxide as an indicator of microbial growth. The media formulation consists of: Middlebrook 7H9 Broth (0.47% w/v), Pancreatic Digest of Casein (0.1% w/v), Bovine Serum Albumin (1.0% w/v), Catalyse (48 μ / ml), in purified water. Bottle reflectance is monitored and recorded by the instrument every 10 minutes. The growth curve enters lag phase then the bottle is flagged positive. At the time of detection, approximate colony forming units (CFUs) are 106–107 per ml.

Ziehl-Neelsen/Acid Fast staining

Bacterial culture smear was prepared from samples indicated positive in BacT/ALERT 3D system on clean

Sero-diagnosis of tuberculosis in elephants

and grease free slide, using standard protocol of Ziehl-Neelsen staining kit (Hi-Media Pvt. Ltd, India).

PCR detection of mycobacterium

DNA was extracted from blood samples and samples signaled positive in BacT/ALERT 3D system of 15 elephants using the extraction protocol described by Samrook et al. 1989 and Tissue Genomic DNA Extraction Mini Kit (FAVORGEN Biotech Corp, Taiwan). The extracted DNA was subjected to PCR by using the standard primer RD4 F 5'-AATGGTTTGGTCATGACGCCTTC-3': R 5'-CCCGTAGCGTTACTGAGAAATTGC-3' and RD1 5'-CCCTTTCTCGTGTTTATAGTTTGA-3' F R 5'-GCCATATCGTCCGGAGCTT-3' which was amplified 176 and 110 bp of Mycobacterium tuberculosis and Mycobacterium bovis. The PCR reaction was carried out at 94°C for 10 minutes followed by 35 cycles of denaturation at 94°C for 1 minute, annealing at 60°C for 30 seconds and extension at 72°C for 1 minute, with final extension at 72°C for 10 minutes. The PCR products were analysed by electrophoresis in 1.5% agarose gel at100 V for 45 minutes and documented. Amplicon of size 176bp and 110bp is specific for Mycobacterium genus.

RESULTS

The Table 1 shows the results of various diagnostic tests used for diagnosis of mycobacterium in elephants. The serum samples collected from the 15 elephants

Table 1. Overall results of test applied (n= 15).

were seronegative by the rapid test kit as no coloured band was observed in the test area of the rapid test kit (Image 1). All the 15 samples were detected positive by the BacT/ALERT 3D system in 6 mean days. These samples were further subjected to ZN staining, no sample detected the presence of acid fast bacilli (Amer et al. 2016; Bapat et al. 2017) (Image 2). Isolates of DNA extracted from the blood samples of these 15 elephants were subjected to PCR which did not produce specific amplicon of 176bp and 110bp RD4 and RD1 gene. Similarly, the DNA isolates from the BacT/ALERT culture system did not produce amplicon of 176 and 110 bp but one isolate produced amplicon of 176bp of RD4 of targeted gene indicating presence of Mycobacterium bovis (BCG) (Bapat et al. 2017) as illustrated in Image 3 and 4.

DISCUSSION

Tuberculosis is a highly contagious zoonotic disease with high incidence and prevalence in human, domestic and wild animals of developing countries. Tuberculosis infection in captive elephants is ongoing and complex problem with respect to their conservation. Due to atypical nature of the mycobacteria that causes diseases, the diagnosis is rather complicated, apart from the fact that many diagnostic tests are developed for domestic species however, those are not validated for wild animals. Therefore, many tests have sub-optimal specificity and sensitivity.

Elephant No.	BacT/ALERT	ZN Staining	Blood PCR	BacT/ALERT + ve PCR	Rapid test
(E1)	Positive	Negative	Negative	Negative	Negative
(E2)	Positive	Negative	Negative	Negative	Negative
(E3)	Positive	Negative	Negative	Negative	Negative
(E4)	Positive	Negative	Negative	Negative	Negative
(E5)	Positive	Negative	Negative	Negative	Negative
(E6)	Positive	Negative	Negative	Negative	Negative
(E7)	Positive	Negative	Negative	Negative	Negative
(E8)	Positive	Negative	Negative	Positive	Negative
(E9)	Positive	Negative	Negative	Negative	Negative
(E10)	Positive	Negative	Negative	Negative	Negative
(E11)	Positive	Negative	Negative	Negative	Negative
(E12)	Positive	Negative	Negative	Negative	Negative
(E13)	Positive	Negative	Negative	Negative	Negative
(E14)	Positive	Negative	Negative	Negative	Negative
(E15)	Positive	Negative	Negative	Negative	Negative

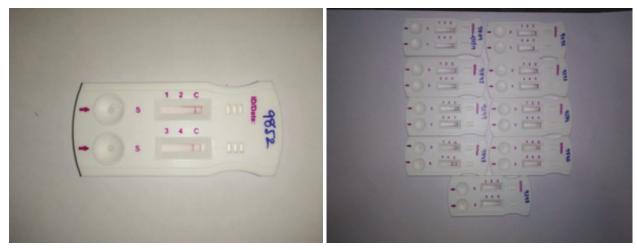


Image 1. Results of rapid TB test kit in elephants screened for tuberculosis.

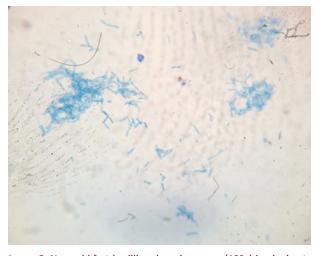
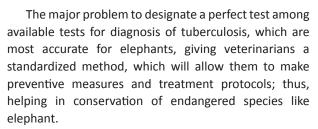


Image 2. Non-acid fast bacilli under microscope (100x) in elephants screened for tuberculosis.



These samples were subjected to diagnostic tests like BacT/ALERT 3D system, ZN staining, PCR, Rapid TB test kit. All 15 samples were signalled positive by BacT/ ALERT 3D system. This test is not yet used and validated in animals, like in humans. This was the first time when the test was used in detection of TB in wild animals. Therefore, the specificity still remains a question. On the other hand, other tests like ZN staining, Rapid TB test kit and blood PCR did not detect any mycobacteria in the

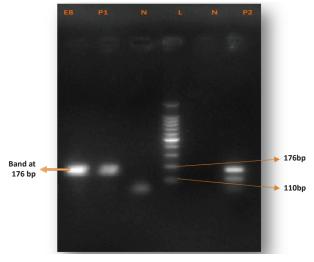


Image 3. PCR pattern of RD4 and RD1 gene at 176bp and 110bp of BacT/ALERT tuberculosis positive sample. Lane E8: positive sample showing band at 176bp of RD4 gene, Lane P1:

positive control (*M. bovis*), Lane P2: Postive control (*M. tuberculosis & M. bovis*), Lane N: negative control, Lane L: DNA ladder 100 bp.

samples.

Molecular detection (duplex PCR) of the samples that signalled positive in BacT/ALERT 3D system was carried out using RD4 and RD1 gene primer with amplicon size of 176bp and 110bp respectively as described by Bapat et al. (2017). Only one sample was positive detecting the presence of *M. bovis* (BCG) at 176bp of RD4 gene.

During the study it was not possible to calculate the specificity of various diagnostic tests used. Development and use of new and more species specific diagnostic methods are needed at the moment, as it will help in early and accurate diagnosis that might permit early application of preventive measures and will ensure

Sero-diagnosis of tuberculosis in elephants

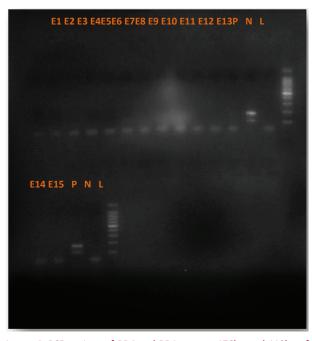


Image 4. PCR pattern of RD4 and RD1 gene at 176bp and 110bp of blood samples

Lane E1-15: negative elephant DNA isolates, Lane P: Positive control (*M. tuberculosis* & *M. bovis*), Lane N: negative control, Lane L: DNA ladder 100bp.

Specimen/DNA Museum Information:

Specimen: Blood. Museum: Niche Area of Excellence, Centre for Zoonoses, Indian Council of Agriculture Research (Central India), Nagpur Veterinary College, Nagpur. Voucher Number: NAE9299.

safety of endangered species as well as human staff involved. Moreover, this mycobacterial disease requires long term surveillance plans in order to be effective, as this organism has prolonged incubation and latency.

Although, the reported case of TB in elephant in present study was caused by M. bovis (BCG) which is vaccine strain, its species predilection is still unidentified. Moreover, this animal should be screened multiple times over the period of time to confirm the disease. Cultural isolation of mycobacterium is currently the only gold standard test for TB diagnosis in elephants, but ancillary tests like PCR, BacT/Alert 3D system, rapid TB test kit etc. may be useful. The molecular method (PCR) used in diagnosis of mycobacterium in present study is not a confirmatory test due to its possibility of cross contamination (false positive) and inability to determine the pathogenicity of the organism. As this is a zoonotic disease, transmission of TB can occur between humans, livestock and elephants. Elephants are at risk of contracting TB from infected human (Mahouts). Therefore, Mahouts (handlers) and elephants should undergo periodic TB screening to minimize the risk of animals' health. Zoos and forest elephant camp areas should be encouraged to incorporate protocol for elephant-visitor interactions and periodic screening of animals for tuberculosis.

This study highlights the potential usefulness and efficacy of ante-mortem diagnostic methods. Use of multiple tests helps to achieve high possibility (sensitivity) of tuberculosis detection in elephants rather than using single test; however, it is important to evaluate and validate the test regime and will require addition of more animals in to the study; expectantly allowing in better understanding of tuberculosis in elephants, thus contributing to undertake control measures by state forest department and zoo authorities for conservation of this endangered species.

REFERENCES

- Bapat, P.R., A.S. Satav, S.D. Shekhawat, S.D. Manke, A.A. Husain, A.R. Nayak, A.P. Kawle, L.R. Singh, H.F. Daginawala & R.S. Kashyap (2017a). Molecular diagnosis of zoonotic *Mycobacterium bovis* infection in Melghat, India. *Journal of Zoonotic Diseases* 2(2): 2–16.
- Bapat, P.R., R.S. Dubey, S.D. Shekhawat, A.A. Husain, A.R. Nayak, A.P. Kawle, H.F. Daginawala, L.R. Singh & R.S. Kashyap (2017b). Prevalence of zoonotic tuberculosis and associated risk factors in Central Indian populations. *Journal of Epidemiology and Global Health* 7: 277–283.
- Choudhury, L., D.K. Desai, A. Duckworth, J.W. Easa, P.S. Johnsingh, A.J.T. Fernando, P. Hedges, S. Gunawardena, M. Kurt, F. Karanth, U. Lister, A. Menon, V. Riddle, H. Rubel & A.E. Wikaranayake (2008). *Elephas maximus*. The ICUN Red List of Threatened Species [online] https://doi.org/10.2305/IUCN.UK.2008.RLTS.T7140A12828813.en [viewed 6 March 2019].
- Hirsch, D.C., N.. MacLachian & R.L. Walker (eds.) (2004). Veterinary Microbiology. 2nd edition. Blackwell Publishing Ltd., USA, 223pp.
- Iyer, A.K. (1937). Veterinary science in India, Ancient and modern with special reference to tuberculosis. *Agriculture Livestock India* 7: 718–724.
- Kathleen, A.A., E. Pleydell, M.C. Williams, E.P. Lane, J.F.C. Nyange & A.L. Michel (2002). Mycobacterium tuberculosis: an emerging disease of free-ranging wildlife. *Emerging Infectious Diseases* 8(6): 598–601.
- Mikota, S.K. (ed.) (2008). Zoo and Wild Animal Medicine 6th Edition. Elsevier Saunders Inc., USA, 355pp.
- Mc Gaughey C.A. (1961). Diseases of elephants Part 3. Ceylon Veterinary Journal 9: 94–98.
- Paudel, S. & T. Tsubota (2016). Tuberculosis in elephants: a zoonotic disease at the human-elephant interface. *Japanese Journal of Zoo* and Wildlife Medicine 21(3): 65–69.
- Project Elephant Division Goverment of India (2017). [online] http:// www.moef.nic.in/division/project-elephant/
- Seneviratna, P., S.G. Wttimuny & D. Senevirtna (1966).Fatal tuberculosis pneumonia in an elephant. Veterinary Medicine, Small Animal Clinician 60: 129–132.
- Samrook J., E.F. Fritsch & T. Maniatis (1989). Molecular cloning: alaboratory manual. Cold Spring Harbor Laboratory Press, New York. *Brazilian Journal of Biology* 62(3): 387–408.

Sero-diagnosis of tuberculosis in elephants

Author details: UTKARSH RAJHANS is currently working as Veterinarian and is Managing director at Fauna Healthcare Veterinary Clinic, Pune, Maharashtra. He has also worked as Veterinarian and CT scan consultant at Cessna Lifeline Veterinary Hospital, Bangalore, Karnataka for 2 years. GAYATRI WANKHEDE is currently working as Assistant Professor. Author is young veterinarian & academician having experience of 8yrs of teaching, research & extension activities. She has published research more than 10 articles in reputed national journals. BALAJ AMBORE is currently working as Assistant Professor & Head. Author has total 24 years of experience in teaching, research, extension and Clinician in the field of Veterinary Science. He has published more than 30 research articles in national and international journals of repute. Dr. SANDEEP CHAUDHARI is currently working as Professor & Head, having experience of 21 years in teaching, research and extension activities has published more than 80 research articles in reputed international and national journals. Dr. NAVNATH KESHAV NIGHOT, MVSc (Vet. Medicine), Formarly Deupty Director, Rajiv Gandhi Zoological Park and Wildlife Research Center, Pune. Having more 16yrs experience of wild animal medicine and therapeutics. VITTHAL DYAYGUDE is currently working as Assistant Professor and Head. Author has 13 years of experience as an academician and He has published more than 30 research papers and articles in journal of national and international reputes. Dre. CHHYAA P. SONEKAR, PhD scholar in the Department of Veterinary Public Health, Nagpur Veterinary College, Nagpur. Currently working on tuberculosis in livestock animals, in and around the Nagpur region along with the zoonotic aspect in humans.

Author contributions: Dr. Utkarsh Rajhans, designed and conducted study on Sero-diagnosis of tuberculosis in elephants in Maharashtra, India. Dr. Gayatri Wankhede and Dr. Balaji Ambore helped in coordinated and guided in the research and manuscript writeup. Dr. Sandeep Chaudhari, Dr. Vitthal Dhaygude and Dr. Chhaya Sonekar designed, performed and analyzed the diagnostic procedures and data. The manuscript was written by Dr. Utkarsh Rajhans and commented by all authors.







The Journal of Threatened Taxa (JoTT) is dedicated to building evidence for conservation globally by publishing peer-reviewed articles online every month at a reasonably rapid rate at www.threatenedtaxa.org. All articles published in JOTT are registered under Creative Commons Attribution 4.0 International License unless otherwise mentioned. JoTT allows allows unrestricted use, reproduction, and distribution of articles in any medium by providing adequate credit to the author(s) and the source of publication.

ISSN 0974-7907 (Online) | ISSN 0974-7893 (Print)

June 2021 | Vol. 13 | No. 7 | Pages: 18679-18958 Date of Publication: 26 June 2021 (Online & Print) DOI: 10.11609/jott.2021.13.7.18679-18958

Short Communications

www.threatenedtaxa.org

Communications

Persistence of Trachypithecus geei (Mammalia: Primates: Cercopithecidae) in a rubber plantation in Assam, India

– Joydeep Shil, Jihosuo Biswas, Sudipta Nag & Honnavalli N. Kumara, Pp. 18679–18686

Population assessment of the endangered Western Hoolock Gibbon Hoolock hoolock Harlan, 1834 at Sheikh Jamal Inani National Park, Bangladesh, and conservation significance of this site for threatened wildlife species

– M. Tarik Kabir, M. Farid Ahsan, Susan M. Cheyne, Shahrul Anuar Mohd Sah, Susan Lappan, Thad Q. Bartlett & Nadine Ruppert, Pp. 18687-18694

Assessment of changes over a decade in the patterns of livestock depredation by the Himalayan Brown Bear in Ladakh. India

Aishwarva Maheshwari, A. Arun Kumar & Sambandam Sathvakumar, Pp. 18695–18702

Habitat selection of Himalayan Musk Deer Moschus leucogaster (Mammalia: Artiodactyla: Moschidae) with respect to biophysical attributes in Annapurna Conservation Area of Nepal - Bijaya Neupane, Nar Bahadur Chhetri & Bijaya Dhami, Pp. 18703-18712

Sero-diagnosis of tuberculosis in elephants in Maharashtra, India

– Utkarsh Rajhans, Gayatri Wankhede, Balaji Ambore , Sandeep Chaudhari, Navnath Nighot, Vitthal Dhaygude & Chhaya Sonekar, Pp. 18713–18718

Avian species richness in traditional rice ecosystems: a case study from upper Myanmar - Steven G. Platt, Myo Min Win, Naing Lin, Swann Htet Naing Aung, Ashish John & Thomas R. Rainwater, Pp. 18719–18737

Conservation status, feeding guilds, and diversity of birds in Daroji Sloth Bear Sanctuary, Karnataka. India

- M.N. Harisha, K.S. Abdul Samad & B.B. Hosetti, Pp. 18738–18751

Birds of Surat-Dangs: a consolidated checklist of 75 years (1944–2020) with special emphasis on noteworthy bird records and bird hotspots from northern Western Ghats of Gujarat, India - Nikunj Jambu & Kaushal G. Patel, Pp. 18752-18780

Identification of a unique barb from the dorsal body contour feathers of the Indian Pitta Pitta brachyura (Aves: Passeriformes: Pittidae) - Prateek Dey, Swapna Devi Ray, Sanjeev Kumar Sharma, Padmanabhan Pramod & Ram Pratap

Singh, Pp. 18781-18791

Underestimated diversity of Cnemaspis Strauch, 1887 (Sauria: Gekkonidae) on karst landscapes in Sarawak, East Malaysia, Borneo

- Izneil Nashriq & Indraneil Das, Pp. 18792-18799

Aborichthys barapensis, a new species of river loach (Cypriniformes: Nemacheilidae) from Arunachal Pradesh, the eastern Himalaya, India – P. Nanda & L. Tamang, Pp. 18800–18808

A study on the community structure of damselflies (Insecta: Odonata: Zygoptera) in Paschim Medinipur, West Bengal, India

– Pathik Kumar Jana, Priyanka Halder Mallick & Tanmay Bhattacharya, Pp. 18809–18816

New distribution and range extension records of geometrid moths (Lepidoptera: Geometridae) from two western Himalayan protected areas

- Pritha Dey & Axel Hausmann, Pp. 18817-18826

Butterfly diversity of Putalibazar Municipality, Syangja District, Gandaki Province, Nepal - Kismat Neupane & Mahamad Sayab Miya, Pp. 18827-18845

New records and distribution extension of Nassarius persicus (Martens, 1874) and N. tadjallii Moolenbeek, 2007 (Mollusca: Gastropoda: Nassariidae) to India - Sayali Nerurkar & Deepak Apte, Pp. 18846-18852

Flowering plants of Agumbe region, central Western Ghats, Karnataka, India - G.S. Adithya Rao & Y.L. Krishnamurthy, Pp. 18853-18867

Population assessment and habitat distribution modelling of the threatened medicinal plant Picrorhiza kurroa Royle ex Benth, in the Kumaun Himalaya, India - Naveen Chandra, Gajendra Singh, Shashank Lingwal, M.P.S. Bisht & Lalit Mohan Tewari, Pp. 18868-18877

Occurrence of gilled fungi in Puducherry, India

- Vadivelu Kumaresan, Chakravarthy Sariha, Thokur Sreepathy Murali & Gunasekaran Senthilarasu, Pp. 18878-18887

First photographic evidence and distribution of the Indian Pangolin Manis crassicaudata (Mammalia: Pholidota: Manidae) in Sariska Tiger Reserve, Rajasthan, India Hemant Singh, Gobind Sagar Bhardwaj, N. Gokulakannan, Saket Agasti & K. Aditya, Pp. 18888-18893

Population and conservation threats to the Greater Flamingos Phoenicopterus roseus (Aves: Phoenicopteriformes: Phoenicopteridae) at Basai Wetland and Najafgarh Jheel Bird Sanctuary, Haryana, India

– Amit Kumar & Sarita Rana, Pp. 18894–18898

First report on the occurrence of Sargassum Weed Fish Histrio histrio (Lophiliformes: Antennariidae) in Nigeria deep water, Gulf of Guinea - Abdul-Rahman Dirisu, Hanson S. Uyi & Meshack Uyi, Pp. 18899-18902

A new distribution record of stomatopods Odontodactylus japonicus (De Haan, 1844) and Lysiosquilla tredecimdentata (Holthuis, 1941) from the Puducherry coastal waters, east coast of India

- S. Nithya Mary, V. Ravitchandirane & B. Gunalan, Pp. 18903-18907

New records of Agriocnemis keralensis Peters, 1981 and Gynacantha khasiaca MacLachlan, 1896 (Insecta: Odonata) from Maharashtra, India – Yogesh Koli, Akshay Dalvi & Dattaprasad Sawant, Pp. 18908–18919

A new distribution record of the Horn Coral Caryophyllia grandis Gardiner & Waugh, 1938 (Anthozoa: Scleractinia) from the Karnataka Coast, India – J.S. Yogesh Kumar & C. Raghunathan, Pp. 18920–18924

Re-collection, extended distribution, and amplified description of Vaccinium paucicrenatum Sleumer (Ericaceae) from the Arunachal Himalaya in India - Subhasis Panda, Pp. 18925-18932

Notes

Photographic record of the Rusty-spotted Cat Prionailurus rubiginosus (I. Geoffroy Saint-Hilaire, 1831) (Mammalia: Carnivora: Felidae) in southern Western Ghats, India - Devika Sanghamithra & P.O. Nameer, Pp. 18933-18935

Natural history notes on the highly threatened Pinto's Chachalaca Ortalis remota (Aves: Cracidae) - Carlos Otávio Araujo Gussoni & Marco Aurélio Galvão da Silva, Pp. 18936-18938

Black-bellied Coral Snake Sinomicrurus nigriventer (Wall, 1908) (Elapidae): an extended distribution in the western Himalaya, India

- Sipu Kumar, Jignasu Dolia, Vartika Chaudhary, Amit Kumar & Abhijit Das, Pp. 18939-18942

First record of the Afghan Poplar Hawkmoth Laothoe witti Eitschberger et al., 1998 (Sphingidae: Smerinthinae) from India: a notable range extension for the genus

– Muzafar Riyaz, Pratheesh Mathew, Taslima Shiekh, S. Ignacimuthu & K. Sivasankaran, Pp. 18943– 18946

The tribe Cnodalonini (Coleoptera: Tenebrionidae: Stenochiinae) from Maharashtra with two new records

- V.D. Hegde & D. Vasanthakumar, Pp. 18947-18948

Do predatory adult odonates estimate their adult prey odonates' body size and dispersal ability to proceed with a successful attack?

- Tharaka Sudesh Priyadarshana, Pp. 18949–18952

Rediscovery of Ophiorrhiza incarnata C.E.C. Fisch. (Rubiaceae) from the Western Ghats of India after a lapse of 83 years

– Perumal Murugan, Vellingiri Ravichandran & Chidambaram Murugan, Pp. 18953–18955

Response

Comments on the "A checklist of mammals with historical records from Darjeeling-Sikkim Himalaya landscape, India" Publisher & Host

- P.O. Nameer, Pp. 18956-18958

