

OPEN ACCESS The Journal of Threatened Taxa is dedicated to building evidence for conservation globally by publishing peer-reviewed articles online every month at a reasonably rapid rate at www.threatenedtaxa.org. All articles published in JoTT are registered under Creative Commons Attribution 4.0 International License unless otherwise mentioned. JoTT allows unrestricted use of articles in any medium, reproduction, and distribution by providing adequate credit to the authors and the source of publication.

# **Journal of Threatened Taxa**

Building evidence for conservation globally

www.threatenedtaxa.org ISSN 0974-7907 (Online) | ISSN 0974-7893 (Print)

# ΝΟΤΕ

# **INTRAGUILD PREDATION OF GREEN LACEWING LARVAE (NEUROPTERA:** CHRYSOPIDAE) ON SPIDER EGGS AND SPIDERLINGS

K.K. Srikumar, S. Smitha, B. Suresh Kumar & B. Radhakrishnan

26 July 2018 | Vol. 10 | No. 8 | Pages: 12133-12136 10.11609/jott.2555.10.8.12133-12136





For Focus, Scope, Aims, Policies and Guidelines visit http://threatenedtaxa.org/index.php/JoTT/about/editorialPolicies#custom-0 For Article Submission Guidelines visit http://threatenedtaxa.org/index.php/JoTT/about/submissions#onlineSubmissions For Policies against Scientific Misconduct visit http://threatenedtaxa.org/index.php/JoTT/about/editorialPolicies#custom-2 For reprints contact <info@threatenedtaxa.org>







Journal of Threatened Taxa | www.threatenedtaxa.org | 26 July 2018 | 10(8): 12133–12136

# INTRAGUILD PREDATION OF GREEN LACEWING LARVAE (NEUROPTERA: CHRYSOPIDAE) ON SPIDER EGGS AND SPIDERLINGS

# K.K. Srikumar<sup>1</sup>, S. Smitha<sup>2</sup>, B. Suresh Kumar<sup>3</sup> & B. Radhakrishnan<sup>4</sup>

 <sup>1-4</sup> UPASI Tea Research Foundation, Nirar Dam (P.O.), Valparai, Coimbatore, Tamil Nadu 642127, India
<sup>1</sup> Present address: PT Riau Andalan Pulp and Paper, AAA R & D, RAPP Ltd., Pangalan Kerinci 28312, Indonesia
<sup>1</sup> sreeku08@gmail.com (corresponding author), <sup>2</sup> smitazoology@ gmail.com, <sup>3</sup> bsksuresh999@gmail.com, <sup>4</sup> dr.radhaupasi@gmail.com

Tea, Camellia sinensis L. (O. Kuntze) plantation provides habitats for thousands of insect species including pests and their natural enemies like parasitoids and predators. The immense value of predators in pest suppression has been well understood by entomologists and there is a renewed interest in biological pest suppression. Classical biological control or periodic inundative release of natural enemies has been most effective in cropping systems where large-scale use of insecticides or their ecologically disruptive practices are minimal (David & Easwaramoorthy 1988). Green lacewings are known to have tolerance to commonly used pesticides (Bigler 1984), and they are relatively easy to rear in captivity (Tulisalo et al. 1984). Laboratory culture and augmentation of Mallada desjardinsi (= boninensis) is feasible through Corcyra cephalonica larvae and artificial diet (Vasanthkumar et al. 2012).

*Mallada desjardinsi* is an important predator of pests such as mealy bugs and aphids. In tea they are important predator of Red Spider Mite *Oligonychus coffeae* (Vasanthkumar et al. 2012). Distribution study in India showed Bengaluru, Karnataka, to have the highest density of *M.* desjardinsi population (26.6% and  $5.05 \pm 0.108$  per plant) in the areas sampled (Boopathi et al. 2016).

The larvae of green lacewings are important predators largely used as biological agents. They feed on pest thrips, aphids, scales,



ISSN 0974-7907 (Online) ISSN 0974-7893 (Print)

**OPEN ACCESS** 



caterpillars, and spider mites infesting a variety of plants (McEwen et al. 2001). Adults of green lacewing generally are not predatory and feed on nectar, pollen or honeydew while a few of them are predatory (Coppel & Mertins 1977).

Mallada desjardinsi (Navas) (= boninensis) (Neuroptera: Chrysopidae), is reported as an important predator of red spider mite (RSM) Oligonychus coffeae Nietner (Acari: Tetranychidae) (Babu et al. 2004; Vasanthkumar et al. 2012). Mallada desjardinsi are also considered as generalist predators and are reported as important natural enemies of a variety of pests such as mealy bugs (Mani & Krishnamoorthi 1987), white flies (Selvakumaran et al. 1996), bollworms and aphids (Kabissa et al. 1996).

Generalist predators prefer to take prey of whatever size they can handle (Dong & Polis 1992; Finke 1994). If these prey include younger conspecifics or other predators, then control of the herbivore population is not guaranteed. Intraguild predation (IGP) is a combination of the killing and eating species that use similar, often limiting, resources and are thus potential competitors (Polis & Myers 1989); however, there has been little research on the IGP of chrysopids, especially on the *M*.

DOI: http://doi.org/10.11609/jott.2555.10.8.12133-12136

Editor: K. Rajmohana, Zoological Survey of India, Kolkata, India.

Date of publication: 26 July 2018 (online & print)

Manuscript details: Ms # 2555 | Received 02 January 2017 | Final received 05 July 2018 | Finally accepted 10 July 2018

Citation: Srikumar, K.K., S. Smitha, B.S. Kumar & B. Radhakrishnan (2018). Intraguild predation of green lacewing larvae (Neuroptera: Chrysopidae) on spider eggs and spiderlings. *Journal of Threatened Taxa* 10(8): 12133–12136; http://doi.org/10.11609/jott.2555.10.8.12133–12136

Copyright: © Srikumar et al. 2018. Creative Commons Attribution 4.0 International License. JoTT allows unrestricted use of this article in any medium, reproduction and distribution by providing adequate credit to the authors and the source of publication.

Funding: UPASI Tea Research Foundation.

Competing interests: The authors declare no competing interests.

Acknowledgements: The authors are thankful to UPASI Tea Research Foundation for providing financial assistance.



12133

## Predation of green lacewing larvae on spider eggs and spiderling

*desjardinsi* in tea ecosystem. Thus the seasonality, IGP on spider eggs and spiderlings and the relationship between the proportion of larvae population and spider population were studied to provide a theoretical foundation for future studies.

The current study was undertaken from December 2014 to November 2015 at UPASI Experimental Farm, Valparai (10.36666°N & 76.96666°E, 1,065m) in Anamallais province, Tamil Nadu, southern India. Daily field surveys were conducted randomly for a year during morning hours (08:00–10.00 hr) in tea plantations. Wild guava trees, (Psidium guajava L.) dispersed in tea plantations support a huge population of the predator Mallada desjardinsi. The identification of lacewing was done following standard reference (Babu et al. 2004). The green lacewing larvae were collected and recorded on guava trees. Recorded lacewing larvae were tabulated on monthly interval. The spider population was also assessed in the trees. Mallada desjardinsi larvae were found in and around spider egg sacs and spiderlings. The spiders were collected in small glass vials (5ml) with 90% alcohol, brought to the laboratory and identified using standard reference (Tikader 1987; Sebastian & Peter 2009).

Mallada desjardinsi's prey preference was derived from extensive field observations of spiderlings and egg sacs feeding and identification of prey carcasses (trash) taken from the larvae. The larvae of *M. desjardinsi* were collected in glass tubes (25×200 mm length) and brought to the laboratory and trash was examined using a stereomicroscope. Spider egg sacs were also examined in the laboratory. Egg sacs were opened and examined using a

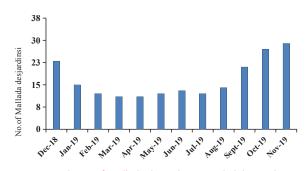


Figure 1. Population of *Mallada desjardinsi* recorded during the period of 2014–2015.

stereomicroscope to determine the number consumed by the predator. Any egg that appeared deflated was counted as consumed. The population abundance of *M. desjardinsi* and spiders (spiderlings and egg sacs) were correlated using Spearman's rank correlation (Siegel & Castellan 1988).

Mallada desjardinsi is a common chrysopid in tea plantations. The larvae are trash carriers and cover themselves with fluffy heaps of debris that conceal their body. The covering included remains of spider egg sacs. It is held in place by hooked spines or bristles on the larva's body. When in motion, the larva's legs and large mandibles can be seen on close inspection. The present study revealed that the fluctuation patterns of *M. desjardinsi* are more or less synchronized in different months. The population was higher during the months of September to December. A synchronized pattern of low population was observed during February to August (Fig. 1). According to regression analysis it seems that

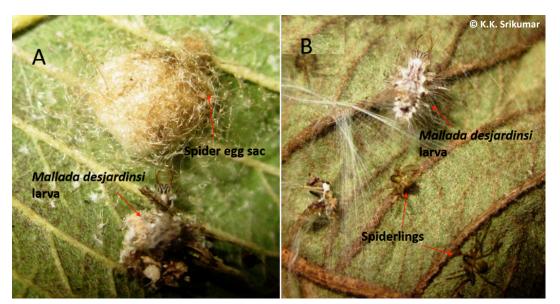


Image 1. Mallada desjardinsi larva predation on spider (A) egg sac and (B) spiderlings

#### Predation of green lacewing larvae on spider eggs and spiderlings

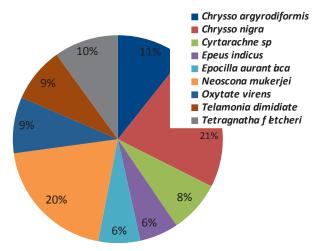


Figure 2. Spider egg sac preference by Mallada desjardinsi

the increase in number of species of lacewings correlates with increasing winter temperature, while they decrease with increasing summer precipitation (McEwen et al. 2001).

A total of nine species of spiders were recorded on the guava trees, viz., Epeus indicus Proszynski, Epocilla aurantiaca Simon, Chrysso nigra O.P. Cambridge, Chrysso argyrodiformis Yaginuma, Cyrtarachne sp., Neoscona mukerjei Tikader, Oxytate virens Thorell, Telamonia dimidiate Simon and Tetragnatha fletcheri Gravely. Neoscona mukerjei and Cyrtarachne sp. belongs to Araneidae (Orbweb spiders). Epeus indicus, Epocilla aurantiaca and Telamonia dimidiate are jumping spiders (Salticidae). Chrysso nigra, C. argyrodiformis and Cyrtarachne sp. belongs to Theridiiae (Comb-footed spiders). Oxytate virens commonly called as green crab spider (Thomisidae). Tetragnatha fletcheri Gravely (Long-jawed spiders) belongs to Tetragnathidae. These spiders construct small, irregular webs, typically on the underside of leaves and within the branches. The larvae of M. desjardinsi are voracious feeders on these spider egg sacs. The larvae actively seek a previously constructed spider egg sac that they enter through direct penetration (Image 1).

Preference was mostly for abandoned egg sacs and spiderlings. In the field it was observed that the fully grown larvae of *M. desjardinsi*, roamed near and consumed 22% eggs of *C. nigra*, 20% of *N. mukerjei*, 11% of *C. argyrodoformis* and 10% of *Cyrtarachne* sp. and below 10% of the other spider species (Fig. 2).

In the laboratory studies Vanitha et al. (2009) showed that when egg sacs were offered to fully grown larvae of *Chrysoperla*, they consumed eggs of *Oxyopes javanus* and *Clubiona drassodes*, whereas no consumption was observed when the mother was present. The popula-

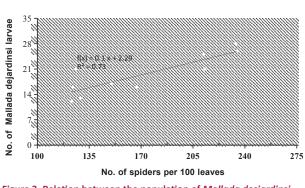


Figure 3. Relation between the population of *Mallada desjardinsi* larvae and spiders

tion of *M. desjardinsi* showed a positive correlation (R = 0.7347) with spider population. Thus, the larger the population of spiderlings and egg sacs, the greater the *M. desjardinsi* population (Fig. 3). Noppe et al. (2012) reported that green lacewing, *Chrysoperla carnea* was the superior intraguild predator, winning 88.9% when the experiment was repeated in petri dishes without plant material, regardless of whether green bugs or eggs of *Ephestia kuehniella* Zeller were offered as focal prey.

Intraguild predation by *M. desjardinsi* can be regarded as a mechanism for enabling survival when the red spider mite prey is scarce. Nevertheless, the intraguild predation of *M. desjardinsi* may reduce pest suppression in tea plantations.

#### References

- Babu, A., R. Selvasundaram, N. Muraleedharan & R. Sasidhar (2004). A new predator of red spider mites. *Newsletter*. *UPASI Tea Res. Foundation* 14: 1–2.
- Bigler, F. (1984) Biological Control by Chrysopids Integration with Pesticides, pp. 233–245. Canard, M., Y. Semeria & R.T. New (eds.). Biology of Chrysopidae. Dr. W. Junk Publishers, Boston.
- Boopathi, T., S. B. Singh, M. Ravi & T. Manju (2016). Distribution and biology of Mallada desjardinsi (Neuroptera: Chrysopidae) in India and its predatory potential against Aleurodicus dispersus (Hemiptera: Aleyrodidae). Journal of Economic Entomology 109: 1988–1994.
- Coppel, H.C. & J.W. Mertins (1977). Biological Insect Pest Suppression. Springer, Berlin, 314pp.
- David, H. & S. Easwaramoorthy (1988). Biocontrol Technology for Sugarcane Pest Management. The Seshan Printers, Coimbatore, 372pp.
- Dong, Q. & G.A. Polis (1992). The dynamics of cannibalistic populations: A foraging perspective, pp. 13–37. Cannibalism. Elgar, M.A. & B.J. Crespi (eds.). *Ecology and Evolution Among Diverse Taxa*. Oxford: Oxford Science Publications.
- Finke, O.M. (1994). Population regulation of a tropical damselfly in the larval stage by food limitation, cannibalism, intraguild predation and habitat drying. *Oecologia* 100: 118–127.
- Kabissa, J.C.B., J.C. Yarro, H.Y. Kayumbo & S.A. Juliano (1996). Functional responses of two chrysopid predators feeding on *Helicoverpa* armigera (Lep.: Noctuidae) and *Aphis gosssypii* (Hom.: Aphididae). Entomophaga 41: 141–151.
- Mani, M. & A. Krishnamoorthy (1987). Feeding potential and development of lacewings, *Mallada boninensis* (Okamota) on the grape

#### Predation of green lacewing larvae on spider eggs and spiderling

mealy bug, Maconellicoccus hirsutus (Green). Entomon 14: 19–21. McEwen, P., T.R. New & A.E. Whittington (2001). Lacewings. The Crop Environment. Cambridge University Press, Cambridge, 546pp.

- Noppe, C., J.P. Michaud & P. De Clercq (2012). Intraguild predation between lady beetles and lacewings: Outcomes and consequences vary with focal prey and arena of interaction. *Annals of the Entomological Society of America* 105(4): 562–571.
- Polis, G.A. & C.A. Myer (1989). The ecology and evolution of intraguild predation: potential competitors that eat each other. *Annual review of Ecology and Systematics* 20: 297–330.
- Sebastian, P.A. & K.V. Peter (2009). Spiders of India. 1st Edition. Universities Press, India, 615pp.
- Selvakumaran, S., M. Kallil, S. Devasahayam (1996). Natural enemies of two major species of scale insects infesting Black Pepper (*Piper nigrum* L.) in India. *Pest Management Horticulture Ecosystem* 2: 79–83.

- Siegel, S. & N.J. Castellan (1988). Nonpara-metric Statistics for the Behavioral Sciences. 2<sup>nd</sup> Edition. McGraw-Hill International Editions, Singapore.
- Tikader, B.K. (1987). Handbook of Indian Spiders. Zoological Survey of India, Calcutta, 251pp.
- Tulisalo, U., M. Canard, Y. Séméria & T.R. New (1984). Biology of Chrysopidae - Biological Control in the Greenhouse. Dr. W. Junk Publishers, Boston, 228–232pp.
- Vanitha, K.P., P. Sivasubramanian, Z. Kavitharaghavan, C. Vijayaraghavan & K. Samiayyan (2009). Prey preference, cross predation and impact of some cultural practices on spiders and their abundance in cotton. *Karnataka Journal of Agricultural Science* 22: 548–551.
- Vasanthakumar, D., R.A. Kumar, J.V. Rahman, P. Kumar, C. Sundaravadivelan & A. Babu (2012). Enhancement of reproductive potential of *Mallada boninensis* Okamoto (Neuroptera: Chrysopidae), a predator of red spider mite infesting tea: an evaluation of artificial diets. Archives of Biological Science 64: 281–285.







OPEN ACCESS The Journal of Threatened Taxa is dedicated to building evidence for conservation globally by publishing peer-reviewed articles online every month at a reasonably rapid rate at www.threatenedtaxa.org. All articles published in JoTT are registered under Creative Commons Attribution 4.0 International License unless otherwise mentioned. JoTT allows unrestricted use of articles in any medium, reproduction, and distribution by providing adequate credit to the authors and the source of publication.

### ISSN 0974-7907 (Online); ISSN 0974-7893 (Print)

### July 2018 | Vol. 10 | No. 8 | Pages: 11999-12146 Date of Publication: 26 July 2018 (Online & Print) DOI: 10.11609/jott.2018.10.8.11999-12146

#### www.threatenedtaxa.org

#### Communications

Habitat suitability and threat analysis of Greater One-horned Rhinoceros Rhinoceros unicornis Linnaeus, 1758 (Mammalia: Perissodactyla: Rhinocerotidae) in Rautahat District, Nepal

-- Saru Rimal, Hari Adhikari & Shankar Tripathi, Pp. 11999–12007

Camera-trapping survey to assess diversity, distribution and photographic capture rate of terrestrial mammals in the aftermath of the ethnopolitical conflict in Manas National Park, Assam, India

-- Dipankar Lahkar, M. Firoz Ahmed, Ramie H. Begum, Sunit Kumar Das, Bibhuti Prasad Lahkar, Hiranya K. Sarma & Abishek Harihar, Pp. 12008–12017

In plain sight: Bacular and noseleaf morphology supports distinct specific status of Roundleaf Bats Hipposideros pomona Andersen, 1918 and Hipposideros gentilis Andersen, 1918 (Chiroptera: Hipposideridae) -- Bhargavi Srinivasulu & Chelmala Srinivasulu, Pp. 12018–12026

#### The amphibian diversity of selected agroecosystems in the southern Western Ghats, India

-- M.S. Syamili & P.O. Nameer, Pp. 12027-12034

Taxonomic status and additional description of White's Stalked-eyed Fly Cyrtodiopsis whitei (Curran, 1936) (Diptera: Diopsidae) from India with a key to the allied species and note on its habitat -- Basant Kumar Agarwala, Pp. 12035-12043

#### Community structure of benthic macroinvertebrate fauna of river Ichamati, India

-- Arnab Basu, Indrani Sarkar, Siddartha Datta & Sheela Roy, Pp. 12044–12055

#### Conservation status of Mascarene Amaranth Aerva congesta Balf.F. Ex Baker (Eudicots: Caryophyllales: Amaranthaceae): a Critically Endangered endemic herb of the Mascarenes, Indian Ocean

-- Kersley Bruno Pynee, David Harold Lorence & Poojanraj Khurun, Pp. 12056-12063

#### Vegetative and reproductive phenology of Aquilaria malaccensis Lam. (Agarwood) in Cachar District, Assam, India

-- Birkhungur Borogayary, Ashesh Kumar Das & Arun Jyoti Nath, Pp. 12064-12072

#### **Conservation Application**

Taking the first steps: Initial mapping of the human-wildlife interaction of the Mauritius Fruit Bat Pteropus niger (Mammalia: Chiroptera: Pteropodidae) in Mauritius by conservation organizations -- Brandon P. Anthony, Vikash Tatayah & Deborah de Chazal, Pp. 12073–12081

#### Peer Commentary

The term human-wildlife conflict creates more problems than it resolves: better labels should be considered -- Priya Davidar, Pp. 12082-12085

#### **Short Communications**

First photographic evidence of Snow Leopard Panthera uncia (Mammalia: Carnivora: Felidae) outside current protected areas network in Nepal Himalava

-- Rinzin Phunjok Lama, Tashi R. Ghale, Madan K. Suwal, Rishi Ranabhat & Ganga Ram Regmi, Pp. 12086–12090





Small carnivores of Silent Valley National Park, Kerala, India -- Devika Sanghamithra & P.O. Nameer, Pp. 12091–12097

Status survey and conservation of the House Sparrow Passer domesticus (Aves: Passeriformes: Passeridae) through public participation in Kannur, Kerala, India

-- R. Roshnath, C.P. Arjun, J. Ashli, D. Sethu & P. Gokul, Pp. 12098–12102

The ecology and distribution of percoid fish Dario neela from Wayanad in the Western Ghats of Kerala, India -- Dencin Rons Thampy & C.P. Shaji, Pp. 12103–12107

A checklist of the ornamental fishes of Himachal Pradesh, the western Himalaya, India

-- Indu Sharma & Rani Dhanze, Pp. 12108–12116

Odonate diversity of Nalsarovar Bird Sanctuary - a Ramsar site in Gujarat, India

-- Darshana M. Rathod & B.M. Parasharya, Pp. 12117–12122

Root holoparasite Balanophora polyandra Griff. (Balanophoraceae) in eastern Himalaya (Sikkim, India): distribution, range, status and threats -- Prem K. Chhetri, Alexander R. O'Neill & Bijoy Chhetri, Pp. 12123–12129

#### Notes

Transfer of Storena gujaratensis Tikader & Patel, 1975 to the genus Suffasia Jocqué, 1991 (Araneae: Zodariidae)

-- Reshma Solanki, Manju Siliwal & Dolly Kumar, Pp. 12130–12132

#### Intraguild predation of green lacewing larvae (Neuroptera: Chrysopidae) on spider eggs and spiderlings

-- K.K. Srikumar, S. Smitha, B. Suresh Kumar & B. Radhakrishnan, Pp. 12133-12136

Rediscovery, extended distribution and conservation assessment of Cinnamomum goaense (Lauraceae) in the Western Ghats, India -- M.P. Geethakumary, S. Deepu & A.G. Pandurangan, Pp. 12137–12139

#### Coltriciella dependens (Berk. & M.A. Curtis) Murrill, a new addition to wood-rotting fungi of India

-- Ayangla S. Pongen, Kuno Chuzho, N.S.K. Harsh, M.S. Dkhar & Manoj Kumar, Pp. 12140-12143

#### **Book Review**

The need of conservation laws coherent with communities for complete success

-- S. Suresh Ramanan & Lalit Upadhyay, Pp. 12144–12145

#### Miscellaneous

**National Biodiversity Authority** 



